#### **REVIEW**



### New strategies for managing adult gliomas

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#### **Abstract**

Gliomas are hard to treat. Their prognosis has improved little over the past few decades. Fundamental therapeutic challenges such as treatment resistance, malignant progression, and tumour recurrence persist. New strategies are needed to advance the management and treatment of gliomas. Here, we focus on where those new strategies could emerge. We consider how recent advances in our understanding of the biology of adult gliomas are informing new approaches to their treatment.

Keywords Glioblastoma · Cancer · Seizure · Treatment resistance · Tumour recurrence · Malignant progression

#### Introduction

Gliomas are the most frequent primary brain tumour in adults [1]. Unlike many other cancers, the prognosis for gliomas has improved little over recent decades [2]. The standard of care is maximal surgical resection with adjuvant chemotherapy and radiotherapy [3–7]. Disappointingly, the 1- and 5-year relative survival rates for patients of all ages with glioblastoma are 41% and 7%, respectively [8]. The age at presentation combined with their poor prognosis means that people with gliomas suffer the greatest loss of "years of potential life" for any adult cancer [9].

Two features of gliomas mean that they are particularly hard to treat and make them prone to recurrence. First, the majority of gliomas are highly invasive [10]. Consequently, the glioma has no clear boundary, which makes it very hard for surgeons to remove the entire tumour. Even removing an entire cerebral hemisphere may not cure [11]. Second, a glioma is not composed of a single clone of cells. Instead, each glioma contains multiple cell types at varying developmental stages and with different genetic and epigenetic signatures [12–14]. Hence, gliomas do not present a well-defined target for (neo)adjuvant chemoradiotherapy.

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In this review, we focus on emerging strategies to manage and treat adult gliomas. The 2016 WHO classification of brain tumours has transformed brain tumour diagnosis by combining molecular and histological features of gliomas. Since then, there have been several excellent recent reviews on different types of gliomas [15–17], standard of care for gliomas with management guidelines [6, 7], and treatment biomarkers [16]. Here, we consider how recent advances in our understanding of the biology of gliomas are informing new and different clinical approaches to tackle treatment resistance, tumour recurrence, and malignant progression.

#### **Genesis of gliomas**

Advances in our understanding of gliomagenesis have led to new strategies for treating gliomas. Two influential ideas have been clonal evolution and the glioma stem cell hypothesis, which is an example of the more general cancer stem cell hypothesis.

Clonal evolution is thought to be a feature of many cancers [18, 19]. Essentially, tumour cells have changes in their DNA, which enable them to replicate without regulation. The clones of tumour cells that are well adapted to their environment proliferate. The emergence of multiple clones of tumour cells and clonal selection can explain the heterogeneity of cell types in tumours, treatment resistance, and tumour recurrence.

The glioma stem cell hypothesis is a more recent idea. Glioma stem cells are a small, sub-population of the glioma cells, which are able to proliferate, self-renew, and propagate



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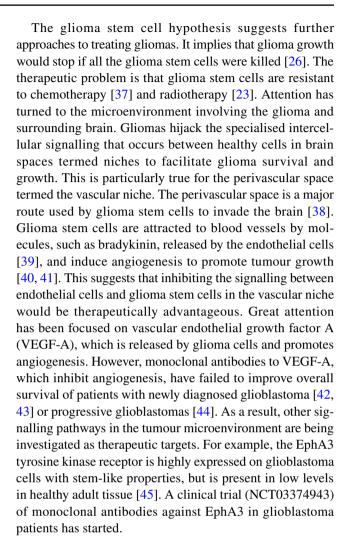
the entire tumour when transplanted [20]. They have been isolated from human glioblastomas [20–22]. The signalling pathways in glioma stem cells are modified to suppress apoptosis and enhance DNA repair, which fosters tumour growth [23]. The progeny of the glioma stem cells develop to varying extents, which generates the phenotypic variability of the glioma cells that form the bulk of glioblastomas [21]. It has been harder to show that glioma stem cells are present in less malignant adult gliomas. Evidence for their existence has come from single-cell RNA sequencing of lower grade gliomas, but it has not yet been shown that glioma cells with stem-like RNA signatures can generate an entire tumour when transplanted [24, 25].

Clonal variation and the cancer stem cell hypothesis were initially thought to be distinct explanations for tumour heterogeneity [26], but others have argued that they should be integrated [27, 28]. The combined hypothesis approach has been applied to gliomagenesis. A first step is that clonal evolution selects glioma cells that have some of the properties of stem cells. Taking this further, glioma stem cells have been proposed to be an end product of clonal variation [29]. The progeny of the glioma stem cells take different differentiation pathways, which contributes to glioma cell heterogeneity [25].

# Clinical significance: gliomagenesis and therapies

Knowledge of the genetic mutations and the resulting tumour proteins implicated in gliomagenesis has opened avenues for the development of new precision medicine treatments, some of which are undergoing trials. For example, mutations in the isocitrate dehydrogenase 1 (IDH1) gene have been identified as driver mutations in many gliomas [30, 31]. Small molecule inhibitors of the mutant IDH1 protein have been developed to target glioma cells [32].

Immunotherapies have taken advantage of our molecular understanding of gliomas too. Gliomas present fragments of tumour-specific proteins called neoantigens on their cell surface. Peptide vaccines have been developed to prime the immune system to attack cells presenting tumour neoantigens. Neoantigens derived from mutant IDH1 were an early target in preclinical models [33]. More recent peptide vaccines trialled in humans target multiple surface proteins specific to the patient's glioma to boost the immune response [34, 35]. These vaccines have generated promising results in early studies, but have yet to be validated in large clinical trials. Similarly, in another immunotherapy, chimeric antigen receptor (CAR) T cells have been modified to express a molecule, chlorotoxin, that binds specifically to glioblastoma cells and boosts the treatment response [36].



### Glioma cellular heterogeneity

The ability to study the molecular profiles of large numbers of single cells enables heterogeneity within gliomas to be probed in detail. Primary glioblastomas (WHO grade IV) are highly aggressive tumours. Molecular profiling using genomic, epigenomic, and single-cell transcriptomic information indicates that individual glioblastoma cells converge on one of four main cellular states: astrocyte-like, mesenchymal-like, oligodendrocyte-like, and neural progenitor-like [46]. Each glioblastoma contains cells from all four states, with one state predominating. Notably, glioblastoma cells can change their molecular profiles and move between states [46]. Hence, killing all the glioblastoma cells in one cellular state will have limited benefit, because the remaining glioblastoma cells will repopulate the "lost" state when treatment ceases. Instead, effective treatment requires that all states are targeted.



### Clinical significance: cellular heterogeneity

Glioma cell heterogeneity has led to a re-evaluation of treatment biomarkers. Temozolomide is an alkylating agent that is the backbone of chemotherapy treatment for gliomas. The effect of Temozolomide is reversed by the endogenous enzyme,  $O^6$ -methylguanine DNA methyltransferase (MGMT). The methylation status of the MGMT gene promoter controls MGMT protein expression and, therefore, the response of gliomas to alkylating chemotherapy agents [47]. The methylation status of the MGMT promoter is measured to determine whether Temozolomide should be given. However, the methylation status of single cells varies within individual gliomas [48]. Importantly, alkylating chemotherapy agents can benefit gliomas with "borderline" methylation status [49]. These findings suggest that multiple biopsies from a glioma are required to fully characterise its methylation status and to guide treatment [50]. This proposal needs to be tested in a clinical trial to see whether patient outcomes are affected.

# Glioma: brain interaction modifies the tumour microenvironment

Glioma cells change their cellular state and molecular profile in response to the microenvironment within and around the tumour [46]. The glioma microenvironment is not just created by ions and molecules released by glioma cells. The brain contributes too by adapting to the glioma growing within it, for example, by activating neuronal plasticity mechanisms. Hence, the glioma environment is a function of both the glioma and the surrounding brain.

The neocortex around gliomas becomes hyperexcitable leading to seizures [51, 52]. It has been proposed that neuronal activity increases the proliferation of high-grade gliomas [53, 54]. Experiments in mouse glioma models suggest that the mechanism is based on the release of the ectodomain of a postsynaptic cell-adhesion molecule, neuroligin-3 (Nlgn3), which acts as a growth factor. Synaptic Nlgn3 is cleaved in response to synaptic activity. The extracellular domain is shed into the extracellular space and diffuses to the glioma cells where it activates glioma signalling pathways that result in cellular proliferation [53, 54]. Blocking the enzyme, ADAM10 that cleaves synaptic Nlgn3 is one therapeutic approach to disrupt this mechanism for glioma growth.

The idea of a direct interaction between a glioma and the surrounding brain has developed further with the finding that neurons form putative synapses on glioma cells. Ultrastructural analysis of human glioma cells transplanted into mouse brains revealed that the glioma cells form structures with neighbouring axons [55–57]. These structures are similar to the excitatory glutamatergic synapses formed between

neurons. Glioma cells express a variety of synaptic transmitter receptors [58–60], which enable paracrine signalling to glioma cells. The existence of functional neuron-to-glioma communication was tested by activating the peritumoural cortex. The glioma cells exhibited a rapid depolarisation with a prominent AMPA-receptor-mediated component, recapitulating excitatory neural transmission [55]. Inhibition of AMPA receptors in xenograft models increased overall survival of the mice by reducing migration and proliferation [55, 56]. In a proportion of glioma cells, the depolarising current spread through gap junctions between glioma cells. The depolarization wave is accompanied by a transient increase in calcium concentration within the glioma cells, which promotes glioma cell invasiveness [55]. These findings suggest that glioma cells integrate themselves into neural circuits. The consequence is that neuronal activity may promote glioma proliferation and invasion.

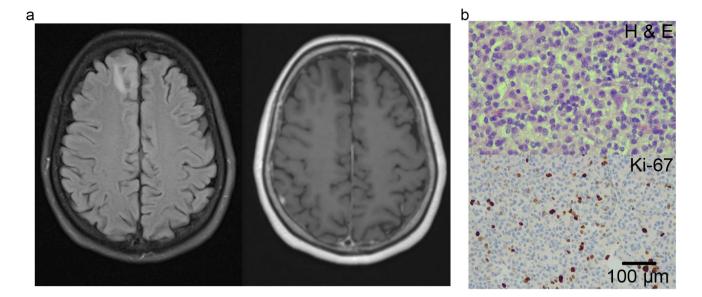
The evidence above considered how neural circuitry in the brain affects gliomas. Newly emerging evidence suggests that the reverse happens; that is, gliomas modify neural circuitry. An early idea was that gliomas caused seizures, because glutamate release from glioma cells resulted in greater excitotoxic cell death of inhibitory interneurons than of excitatory pyramidal cells in the peritumoural cortex [51, 52, 61]. A more nuanced idea has emerged from a mouse glioblastoma model that incorporates mutations in the RTK-RAS-PI3K pathway. Two mutations in the catalytic subunit of the PI3K enzyme, PI3KCA, were identified that altered the expression of synapse-associated genes [62]. Mice with these two mutations developed seizures due to reduced inhibitory synapses and increased excitatory synaptic synapses between neurons in peritumoural cortex. One of the mutations increased the expression of glypican, which promotes the formation of excitatory synapses [63]. When glypican production in the glioma cells is blocked, the mice had fewer seizures and survived longer [62].

Collectively, the recent evidence suggests that a subset of gliomas forms a positive feedback loop with the surrounding peritumoural cortex. The therapeutic challenge now is to break this feedback loop in a way that does not disrupt neural activity and impair cognition.

## Clinical significance: glioma-associated seizures

Epileptic seizures are the most common presenting symptom in patients with glioma [64, 65]. This is concerning, because the recent experimental evidence suggests that aberrant neuronal activity in the surrounding peritumoural cortex facilitates glioma growth [53, 61]. Low-grade glioma patients that present with seizures have longer overall survival if their seizures cease [66]. Furthermore, effective seizure control





**Fig. 1** Predicting malignant transformation in low-grade gliomas. **a** Patient presented with a generalised seizure. Left panel, FLAIR MRI, lesion in the right superior frontal gyrus. Right panel, T1-weighted MRI, lesion showed no contrast enhancement. No increase in perfusion (data not shown). Radiological diagnosis, low-grade glioma.

**b** Neuropathological diagnosis: IDH2 mutant, 1p/19q co-deleted, anaplastic oligodendroglioma (WHO grade III). Top panel, H&E stain exhibiting mitotic figures and apoptotic nuclei, no necrosis, or microvascular proliferation. Bottom panel, Ki-67-positive nuclei stain brown. Proliferation rate  $\sim 10\%$ . Scale bar;  $100 \, \mu m$ 

is the best predictor of quality of life in glioma patients [67]. Consequently, effective seizure control is critically important for people for gliomas. However, glioma-associated seizures frequently respond poorly to anticonvulsant medication [68]. Surgical removal of the epileptic focus during glioma surgery controls seizures in some patients, but not all [69]. New anticonvulsants are needed.

### **Malignant progression**

Some gliomas have an indolent course initially and then behave more aggressively. The terminology around this has been re-evaluated. Malignant transformation was used to describe low-grade (WHO grade I–II) gliomas developing features of high-grade (WHO grade III–IV) tumours. The practise has been undermined by molecular profiling of gliomas, which gives a more accurate prognosis [15]. Increasingly, WHO grade II and grade III gliomas with the same molecular profile are grouped together, because they have a similar prognosis and are referred to as lower grade gliomas [15, 70, 71]. The change from indolent to more aggressive behaviour is better termed malignant progression.

We do not understand what causes malignant progression and where it occurs. Genetic changes associated with malignant progression have been identified [71, 72]. However, there has been less progress in understanding how those genetic changes underpin more aggressive glioma behaviour. In large part, this is due to the lack of suitable

experimental models. True lower grade cell lines have been hard to obtain. Glioma stem cells have been isolated from adult higher grade gliomas, but are challenging to isolate from adult lower grade gliomas. As a result, standard in vitro or xenograft models have been challenging to set up [32, 73, 74]. New model systems are needed.

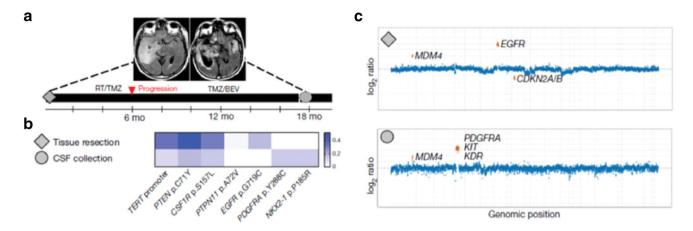
### Clinical significance: malignant progression

Malignant progression is a key clinical issue in the management of gliomas, particularly as lower grade gliomas commonly affect young adults. Malignant progression is frequently detected neuroradiologically by contrast enhancement or increased perfusion on MRI scans. However, these findings are not always present (Fig. 1). The issue is that the neuroradiological measures focus on vascular changes and not on the glioma cells. Consequently, they detect malignant progression when it is established rather than at the earliest stages. New biomarkers to predict early stage malignant progression are needed.

### **Liquid biopsy**

Tissue biopsies give a snapshot of a glioma. However, the molecular profile of glioma cells is not static. It evolves. Ideally, treatment would change to mirror progression of the glioma. A minimally invasive biomarker based on the





**Fig. 2** Cell-free DNA analysis of CSF to follow tumour evolution. **a** Fluid attenuated inversion recovery MRI showing glioblastoma at the time of resection (grey diamond) and time of CSF collection (grey circle). **b** CSF showed a new platelet-derived growth factor

receptor alpha (PDGFRA) mutation. c Copy-number variation plots. CSF showed PDGFRA gene amplification and loss of the epidermal growth factor receptor (EGFR) gene amplification found in the original sample. Figure adapted from Miller et al. Nature 2019 [79]

molecular profile of glioma cells could provide valuable information about tumour evolution. Such a biomarker could also be used to diagnose chemoradiotherapy-induced changes in brain imaging termed pseudoprogression and to monitor response of the glioma to treatment.

There has been huge interest in studying body fluids, such as blood or cerebrospinal fluid, to detect evidence of cancer. This is referred to as liquid biopsy. Blood biomarkers include circulating tumour cells, extracellular vesicles, and cell-free tumour DNA (ctDNA) [75]. Low frequencies of both circulating tumour cells and extracellular vesicles have been identified in the blood of glioma patients. The detection of circulating tumour cells in the blood of glioma patients is highly variable due to the use of different isolation techniques [75]. Extracellular vesicles have been shown to be significantly higher in gliomas with true progression than in patients with pseudoprogression or stable disease [76].

Blood biomarkers have the huge advantage of ease of collection. However, the blood-brain barrier limits access of tumour tissue to the blood. Tests of blood-derived extracellular vesicles and circulating tumour cells have suffered with low sensitivity [77]. As a result, they have not yet reached the clinic.

Liquid biopsy based on cerebrospinal fluid offers an approach that circumvents the blood-brain barrier. Analysis of ctDNA in the CSF successfully identified glioma in approximately half of patients [78, 79]. The probability of identifying ctDNA in the CSF was more successful in higher grade gliomas, suggesting that the presence of ctDNA in CSF could be used to study tumour progression (Fig. 2) [79].

Liquid biomarkers offer a minimally invasive alternative to tissue biopsies. Currently, blood- and CSF-derived biomarkers lack diagnostic specificity, due to the short half-life, penetrance, and dilution of the biomarker [75]. These

limitations may be overcome with interventions such as transcranial-focused ultrasound that enhances the release of the biomarkers into the blood or CSF [80].

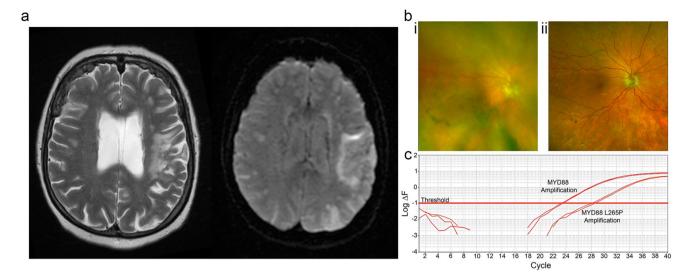
# Clinical significance: liquid biopsy to diagnose glioma mimics?

Liquid biopsy could be used to diagnose glioma mimics. A common diagnostic problem for neuro-oncology services is to differentiate glioblastoma from primary CNS lymphoma. Current practise is that tumour tissue is required from both glioblastoma and primary CNS lymphoma to diagnose the condition and to plan treatment [81]. Acquiring the tumour tissue is almost invariably done with a brain biopsy. Liquid biopsy may soon play a role here. The diagnosis of brain tumours is increasingly based on molecular features [82]. Glioblastoma and primary CNS lymphoma have different mutational landscapes [83, 84]. Therefore, it should be possible to differentiate the two conditions using liquid biopsy. This can be done with a vitreous biopsy as primary CNS lymphoma can involve the eyes (Fig. 3). Analysis of cell-free DNA in the cerebrospinal fluid has great potential, but has not yet reached the clinic.

# Translating basic science findings into glioma management

There has been considerable progress in our understanding of the biology of gliomas. However, many outstanding questions remain (Box 1). First, the finding that glioma growth and invasion is facilitated by aberrant neuronal activity suggests that effective





**Fig. 3** Diagnosis of primary CNS lymphoma with vitreous biopsy. **a** Patient presented with right-sided weakness with stuttering progression over several months. Left panel, T2-weighted MRI, increased signal in the territory of the left middle cerebral artery. Right panel, Diffusion-weighted imaging, increased signal suggesting restricted diffusion in left-hemisphere white matter. **b** Photographs of right fundus for patient in **a**: (i) before vitreous biopsy, retina obscured by cloudy vitreous; (ii) post-vitreous biopsy, retina visible, no sub-retinal

deposits identified. **c** Detection of MYD88 L265P mutation by allelespecific PCR. The test compares the patient's DNA with a positive control that has MYD88 L265P at 0.625%. The test relies on a failure of extension of the primers when there is mismatch between the primer and the extracted DNA. Any difference between the patient's sample and the positive control is amplified by repeated PCR cycles and is measured ( $\Delta F$ ). The presence of the MYD88 L265P mutation is reported when the difference crosses a threshold

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control of seizures is vital for glioma patients. Novel agents to diminish neuronal hyperexcitability will need careful titration to avoid inhibiting neuronal plasticity, as this could impair cognitive function. Enzyme-inducing anticonvulsants need to be avoided as they can increase metabolism of some chemotherapy agents. Blood and CSF liquid biopsy have great potential as methods to diagnose glioma mimics and tumour pseudoprogression, and to monitor tumour progression. However, new approaches are needed to increase the sensitivity of liquid biopsies. What joins all the outstanding questions is the overwhelming need to translate the new knowledge about gliomas into novel strategies for their diagnosis and management.

Compliance with ethical standards

Research Trust, and Inman Charity.

Anwar (Clinical Scientist, KCH) for the MYD88 data.

### **Box 1: Outstanding questions**

**Conflicts of interest** AJK is CEO of Vivisco Limited. GTF is a share-holder and sits on the Advisory Board. Vivisco did not contribute materials or funds to this project.

1. Does control of seizure activity limit glioma growth and invasion?

Ethics approval Clinical data are presented with the patients' consent. Data from one person were collected as part of a larger UK Human Research Authority (https://www.hra.nhs.uk/)-approved study, which had received a favourable opinion from the South West Research Ethics Committee (REC approval code: 18/SW/0022).

2. Is killing glioma stem cells sufficient to reduce glioma growth or must all of the glioma cellular states be attacked?

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- 3. Can new investigations predict which lower grade gliomas will progress into more malignant tumours?
- 4. Can the sensitivity and specificity of liquid biopsy be improved through interventions that increase release of glioma tissue fragments into the blood or CSF?



#### References

- Ostrom QT, Bauchet L, Davis FG et al (2014) The epidemiology of glioma in adults: a "state of the science" review. Neuro Oncol 16:896–913. https://doi.org/10.1093/neuonc/nou087
- Gittleman H, Boscia A, Ostrom QT et al (2018) Survivorship in adults with malignant brain and other central nervous system tumor from 2000 to 2014. Neuro Oncol 20:6–16. https://doi. org/10.1093/neuonc/noy090
- Stupp R, Hegi ME, Mason WP et al (2009) Effects of radiotherapy with concomitant and adjuvant temozolomide versus radiotherapy alone on survival in glioblastoma in a randomised phase III study: 5-year analysis of the EORTC-NCIC trial. Lancet Oncol 10:459–466. https://doi.org/10.1016/S1470-2045(09)70025-7
- Molinaro AM, Hervey-Jumper S, Morshed RA et al (2020) Association of maximal extent of resection of contrast-enhanced and non-contrast-enhanced tumor with survival within molecular subgroups of patients with newly diagnosed glioblastoma. JAMA Oncol 6:495–503. https://doi.org/10.1001/jamaoncol.2019.6143
- Buckner JC, Shaw EG, Pugh SL et al (2016) Radiation plus procarbazine, CCNU, and vincristine in low-grade glioma. N Engl J Med 374:1344–1355. https://doi.org/10.1056/NEJMoa1500925
- Weller M, van den Bent M, Tonn JC et al (2017) European Association for Neuro-Oncology (EANO) guideline on the diagnosis and treatment of adult astrocytic and oligodendroglial gliomas. Lancet Oncol 18:e315–e329. https://doi.org/10.1016/S1470-2045(17)30194-8
- National Institute for Health and Care Excellence (NICE) Guideline [NG 99] (2018) Brain tumours (primary) and brain metastases in adults. https://www.nice.org.uk/guidance/ng99. Accessed 1 Apr 2020
- Ostrom QT, Cioffi G, Gittleman H et al (2019) CBTRUS statistical report: primary brain and other central nervous system tumors diagnosed in the United States in 2012–2016. Neuro Oncol 21:v1–v100. https://doi.org/10.1093/neuonc/noz150
- Rouse C, Gittleman H, Ostrom QT et al (2016) Years of potential life lost for brain and CNS tumors relative to other cancers in adults in the United States, 2010. Neuro Oncol 18:70–77. https:// doi.org/10.1093/neuonc/nov249
- Scherer HJ (1938) Structural development in gliomas. Am J Cancer 3:333–351. https://doi.org/10.1158/ajc.1938.333
- Gardner WJ (1933) Removal of the right cerebral hemisphere for infiltrating glioma. JAMA 101:823–826. https://doi.org/10.1001/ jama.1933.02740360003002
- Patel AP, Tirosh I, Trombetta JJ et al (2014) Single-cell RNA-seq highlights intratumoral heterogeneity in primary glioblastoma. Science 344:1396–1401. https://doi.org/10.1126/science.1254257
- Puchalski RB, Shah N, Miller J et al (2018) An anatomic transcriptional atlas of human glioblastoma. Science 360:660–663. https://doi.org/10.1126/science.aaf2666
- Sottoriva A, Spiteri I, Piccirillo SGM et al (2013) Intratumor heterogeneity in human glioblastoma reflects cancer evolutionary dynamics. Proc Natl Acad Sci USA 110:4009–4014. https://doi.org/10.1073/pnas.1219747110
- Schiff D, van den Bent M, Vogelbaum MA et al (2019) Recent developments and future directions in adult lower-grade gliomas: Society for Neuro-Oncology (SNO) and European Association of Neuro-Oncology (EANO) Consensus. Neuro Oncol 21:837–853. https://doi.org/10.1093/neuonc/noz033
- Reifenberger G, Wirsching H-G, Knobbe-Thomsen CB, Weller M (2017) Advances in the molecular genetics of gliomas—implications for classification and therapy. Nat Rev Clin Oncol 14:434– 452. https://doi.org/10.1038/nrclinonc.2016.204

- 17. Wen PY, Chang SM, Van den Bent MJ et al (2017) Response assessment in neuro-oncology clinical trials. J Clin Oncol 35:2439–2449. https://doi.org/10.1200/JCO.2017.72.7511
- Greaves M, Maley CC (2012) Clonal evolution in cancer. Nature 481:306–313. https://doi.org/10.1038/nature10762
- Nowell PC (1976) The clonal evolution of tumor cell populations.
   Science 194:23–28. https://doi.org/10.1126/science.959840
- Lathia JD, Mack SC, Mulkearns-Hubert EE et al (2015) Cancer stem cells in glioblastoma. Genes Dev 29:1203–1217. https://doi. org/10.1101/gad.261982.115
- Singh SK, Hawkins C, Clarke ID et al (2004) Identification of human brain tumour initiating cells. Nature 432:396–401. https://doi.org/10.1038/nature03128
- Galli R, Binda E, Orfanelli U et al (2004) Isolation and characterization of tumorigenic, stem-like neural precursors from human glioblastoma. Cancer Res 64:7011–7021. https://doi.org/10.1158/0008-5472.CAN-04-1364
- Bao S, Wu Q, McLendon RE et al (2006) Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. Nature 444:756–760. https://doi.org/10.1038/nature05236
- Tirosh I, Venteicher AS, Hebert C et al (2016) Single-cell RNAseq supports a developmental hierarchy in human oligodendroglioma. Nature 539:309–313. https://doi.org/10.1038/nature20123
- Venteicher AS, Tirosh I, Hebert C et al (2017) Decoupling genetics, lineages, and microenvironment in IDH-mutant gliomas by single-cell RNA-seq. Science 355(6332):eaai8478. https://doi.org/10.1126/science.aai8478
- Shackleton M, Quintana E, Fearon ER, Morrison SJ (2009) Heterogeneity in cancer: cancer stem cells versus clonal evolution. Cell 138:822–829. https://doi.org/10.1016/j.cell.2009.08.017
- Clevers H (2011) The cancer stem cell: premises, promises and challenges. Nat Med 17:313–319. https://doi.org/10.1038/ nm.2304
- Kreso A, Dick JE (2014) Evolution of the cancer stem cell model. Cell Stem Cell 14:275–291. https://doi.org/10.1016/j. stem.2014.02.006
- Barthel FP, Wesseling P, Verhaak RGW (2018) Reconstructing the molecular life history of gliomas. Acta Neuropathol 135:649–670. https://doi.org/10.1007/s00401-018-1842-y
- Yan H, Parsons DW, Jin G et al (2009) IDH1 and IDH2 mutations in gliomas. N Engl J Med 360:765–773. https://doi.org/10.1056/ NEJMoa0808710
- 31. Parsons DW, Jones S, Zhang X et al (2008) An integrated genomic analysis of human glioblastoma multiforme. Science 321:1807–1812. https://doi.org/10.1126/science.1164382
- Rohle D, Popovici-Muller J, Palaskas N et al (2013) An inhibitor of mutant IDH1 delays growth and promotes differentiation of glioma cells. Science 340:626–630. https://doi.org/10.1126/scien ce.1236062
- Schumacher T, Bunse L, Pusch S et al (2014) A vaccine targeting mutant IDH1 induces antitumour immunity. Nature 512:324

  –327. https://doi.org/10.1038/nature13387
- 34. Hilf N, Kuttruff-Coqui S, Frenzel K et al (2019) Actively personalized vaccination trial for newly diagnosed glioblastoma. Nature 565:240–245. https://doi.org/10.1038/s41586-018-0810-y
- Keskin DB, Anandappa AJ, Sun J et al (2019) Neoantigen vaccine generates intratumoral T cell responses in phase Ib glio-blastoma trial. Nature 565:234–239. https://doi.org/10.1038/s41586-018-0792-9
- Wang D, Starr R, Chang W-C et al (2020) Chlorotoxin-directed CAR T cells for specific and effective targeting of glioblastoma. Sci Transl Med 12:eaaw2672. https://doi.org/10.1126/scitranslmed.aaw2672



- Chen J, Li Y, Yu T-S et al (2012) A restricted cell population propagates glioblastoma growth after chemotherapy. Nature 488:522–526. https://doi.org/10.1038/nature11287
- Calabrese C, Poppleton H, Kocak M et al (2007) A perivascular niche for brain tumor stem cells. Cancer Cell 11:69–82. https:// doi.org/10.1016/j.ccr.2006.11.020
- Montana V, Sontheimer H (2011) Bradykinin promotes the chemotactic invasion of primary brain tumors. J Neurosci 31:4858–4867. https://doi.org/10.1523/JNEUROSCI.3825-10.2011
- Folkman J (1971) Tumor angiogenesis: therapeutic implications.
   N Engl J Med 285:1182–1186. https://doi.org/10.1056/NEJM197111182852108
- Gilbertson RJ, Rich JN (2007) Making a tumour's bed: glioblastoma stem cells and the vascular niche. Nat Rev Cancer 7:733

  736. https://doi.org/10.1038/nrc2246
- Chinot OL, Wick W, Mason W et al (2014) Bevacizumab plus radiotherapy-temozolomide for newly diagnosed glioblastoma. N Engl J Med 370:709–722. https://doi.org/10.1056/NEJMoa1308 345
- Gilbert MR, Dignam JJ, Armstrong TS et al (2014) A randomized trial of bevacizumab for newly diagnosed glioblastoma. N Engl J Med 370:699–708. https://doi.org/10.1056/NEJMoa1308573
- Wick W, Gorlia T, Bendszus M et al (2017) Lomustine and bevacizumab in progressive glioblastoma. N Engl J Med 377:1954– 1963. https://doi.org/10.1056/NEJMoa1707358
- Day BW, Stringer BW, Al-Ejeh F et al (2013) EphA3 maintains tumorigenicity and is a therapeutic target in glioblastoma multiforme. Cancer Cell 23:238–248. https://doi.org/10.1016/j.ccr.2013.01.007
- Neftel C, Laffy J, Filbin MG et al (2019) An integrative model of cellular states, plasticity, and genetics for glioblastoma. Cell 178:835–849.e21. https://doi.org/10.1016/j.cell.2019.06.024
- Hegi ME, Diserens A-C, Gorlia T et al (2005) MGMT gene silencing and benefit from temozolomide in glioblastoma. N Engl J Med 352:997–1003. https://doi.org/10.1056/NEJMoa043331
- Wenger A, Ferreyra Vega S, Kling T et al (2019) Intratumor DNA methylation heterogeneity in glioblastoma: implications for DNA methylation-based classification. Neuro Oncol 21:616–627. https://doi.org/10.1093/neuonc/noz011
- Pinson H, Hallaert G, Van der Meulen J et al (2020) Weak MGMT gene promoter methylation confers a clinically significant survival benefit in patients with newly diagnosed glioblastoma: a retrospective cohort study. J Neurooncol 146:55–62. https://doi. org/10.1007/s11060-019-03334-5
- Kirby AJ, Lavrador JP, Bodi I et al (2020) Ex vivo ultrasonic samples of human brain tumors in the molecular era. Neurooncol Adv 2:vdaa14. https://doi.org/10.1093/noajnl/vdaa014
- Campbell SL, Buckingham SC, Sontheimer H (2012) Human glioma cells induce hyperexcitability in cortical networks. Epilepsia 53:1360–1370. https://doi.org/10.1111/j.1528-1167.2012.03557.x
- Buckingham SC, Campbell SL, Haas BR et al (2011) Glutamate release by primary brain tumors induces epileptic activity. Nat Med 17:1269–1274. https://doi.org/10.1038/nm.2453
- Venkatesh HS, Tam LT, Woo PJ et al (2017) Targeting neuronal activity-regulated neuroligin-3 dependency in high-grade glioma. Nature 549:533–537. https://doi.org/10.1038/nature24014
- 54. Venkatesh HS, Johung TB, Caretti V et al (2015) Neuronal activity promotes glioma growth through neuroligin-3 secretion. Cell 161:803–816. https://doi.org/10.1016/j.cell.2015.04.012
- Venkatesh HS, Morishita W, Geraghty AC et al (2019) Electrical and synaptic integration of glioma into neural circuits. Nature 573:539–545. https://doi.org/10.1038/s41586-019-1563-y
- Venkataramani V, Tanev DI, Strahle C et al (2019) Glutamatergic synaptic input to glioma cells drives brain tumour progression. Nature 573:532–538. https://doi.org/10.1038/s41586-019-1564-x

- 57. Zeng Q, Michael IP, Zhang P et al (2019) Synaptic proximity enables NMDAR signalling to promote brain metastasis. Nature 573:526–531. https://doi.org/10.1038/s41586-019-1576-6
- Takano T, Lin JH, Arcuino G et al (2001) Glutamate release promotes growth of malignant gliomas. Nat Med 7:1010–1015. https://doi.org/10.1038/nm0901-1010
- Ishiuchi S, Tsuzuki K, Yoshida Y et al (2002) Blockage of Ca(2+)-permeable AMPA receptors suppresses migration and induces apoptosis in human glioblastoma cells. Nat Med 8:971– 978. https://doi.org/10.1038/nm746
- Caragher SP, Hall RR, Ahsan R, Ahmed AU (2018) Monoamines in glioblastoma: complex biology with therapeutic potential. Neuro Oncol 20:1014–1025. https://doi.org/10.1093/neuonc/nox210
- Campbell SL, Robel S, Cuddapah VA et al (2015) GABAergic disinhibition and impaired KCC2 cotransporter activity underlie tumor-associated epilepsy. Glia 63:23–36. https://doi.org/10.1002/ glia.22730
- 62. Yu K, Lin C-CJ, Hatcher A et al (2020) PIK3CA variants selectively initiate brain hyperactivity during gliomagenesis. Nature 578:166–171. https://doi.org/10.1038/s41586-020-1952-2
- Allen NJ, Bennett ML, Foo LC et al (2012) Astrocyte glypicans 4 and 6 promote formation of excitatory synapses via GluA1 AMPA receptors. Nature 486:410–414. https://doi.org/10.1038/ nature11059
- 64. Glantz MJ, Cole BF, Forsyth PA et al (2000) Practice parameter: anticonvulsant prophylaxis in patients with newly diagnosed brain tumors. Report of the Quality Standards Subcommittee of the American Academy of Neurology. Neurology 54:1886–1893. https://doi.org/10.1212/wnl.54.10.1886
- 65. Berntsson SG, Malmer B, Bondy ML et al (2009) Tumor-associated epilepsy and glioma: are there common genetic pathways? Acta Oncol 48:955–963. https://doi.org/10.1080/02841860903104145
- Danfors T, Ribom D, Berntsson SG, Smits A (2009) Epileptic seizures and survival in early disease of grade 2 gliomas. Eur J Neurol 16:823–831. https://doi.org/10.1111/j.1468-1331.2009.02599
- 67. Klein M (2012) Neurocognitive functioning in adult WHO grade II gliomas: impact of old and new treatment modalities. Neuro Oncol 14(Suppl 4):iv17–24. https://doi.org/10.1093/neuonc/nos16
- van Breemen MSM, Rijsman RM, Taphoorn MJB et al (2009) Efficacy of anti-epileptic drugs in patients with gliomas and seizures. J Neurol 256:1519–1526. https://doi.org/10.1007/s0041 5-009-5156-9
- Englot DJ, Berger MS, Barbaro NM, Chang EF (2011) Predictors of seizure freedom after resection of supratentorial low-grade gliomas. A review J Neurosurg 115:240–244. https://doi.org/10.3171/2011.3.JNS1153
- Suzuki H, Aoki K, Chiba K et al (2015) Mutational landscape and clonal architecture in grade II and III gliomas. Nat Genet 47:458–468. https://doi.org/10.1038/ng.3273
- Ceccarelli M, Barthel FP, Malta TM et al (2016) Molecular profiling reveals biologically discrete subsets and pathways of progression in diffuse glioma. Cell 164:550–563. https://doi. org/10.1016/j.cell.2015.12.028
- Bai H, Harmancı AS, Erson-Omay EZ et al (2016) Integrated genomic characterization of IDH1-mutant glioma malignant progression. Nat Genet 48:59–66. https://doi.org/10.1038/ng.3457
- Luchman HA, Stechishin OD, Dang NH et al (2012) An in vivo patient-derived model of endogenous IDH1-mutant glioma. Neuro Oncol 14:184–191. https://doi.org/10.1093/neuonc/nor207
- Kelly JJP, Blough MD, Stechishin ODM et al (2010) Oligodendroglioma cell lines containing t(1;19)(q10;p10). Neuro Oncol 12:745–755. https://doi.org/10.1093/neuonc/noq031



- Müller Bark J, Kulasinghe A, Chua B et al (2019) Circulating biomarkers in patients with glioblastoma. Br J Cancer 122:295–305. https://doi.org/10.1038/s41416-019-0603-6
- Koch CJ, Lustig RA, Yang X-Y et al (2014) Microvesicles as a biomarker for tumor progression versus treatment effect in radiation/temozolomide-treated glioblastoma patients. Transl Oncol 7:752–758. https://doi.org/10.1016/j.tranon.2014.10.004
- Merker JD, Oxnard GR, Compton C et al (2018) Circulating tumor DNA analysis in patients with cancer: American Society of Clinical Oncology and College of American Pathologists Joint Review. Arch Pathol Lab Med 142:1242–1253. https://doi.org/10.5858/ arpa.2018-0901-SA
- 78. Wang Z, Jiang W, Wang Y et al (2015) MGMT promoter methylation in serum and cerebrospinal fluid as a tumor-specific biomarker of glioma. Biomed Rep 3:543–548. https://doi.org/10.3892/br.2015.462
- Miller AM, Shah RH, Pentsova EI et al (2019) Tracking tumour evolution in glioma through liquid biopsies of cerebrospinal fluid. Nature 565:654–658. https://doi.org/10.1038/s41586-019-0882-3

- Zhu L, Cheng G, Ye D et al (2018) Focused ultrasound-enabled brain tumor liquid biopsy. Sci Rep 8:6553. https://doi.org/10.1038/ s41598-018-24516-7
- Fox CP, Phillips EH, Smith J et al (2019) Guidelines for the diagnosis and management of primary central nervous system diffuse large B-cell lymphoma. Br J Haematol 184:348–363. https://doi.org/10.1111/bjh.15661
- Louis DN, Perry A, Reifenberger G et al (2016) The 2016 World Health Organization Classification of tumors of the central nervous system: a summary. Acta Neuropathol 131:803–820. https:// doi.org/10.1007/s00401-016-1545-1
- 83. Brennan CW, Verhaak RGW, McKenna A et al (2013) The somatic genomic landscape of glioblastoma. Cell 155:462–477. https://doi.org/10.1016/j.cell.2013.09.034
- Nakamura T, Tateishi K, Niwa T et al (2016) Recurrent mutations of CD79B and MYD88 are the hallmark of primary central nervous system lymphomas. Neuropathol Appl Neurobiol 42:279–290. https://doi.org/10.1111/nan.12259

