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REVIEW

Glioblastoma multiforme: novel therapeutic targets

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ABSTRACT

Introduction: The increasingly detailed genetic characterization of glioblastoma (GBM) has failed to translate into meaningful breakthroughs in treatment. This is likely to be attributed to molecular heterogeneity of GBM. However, the understanding of the tumor microenvironment in GBM has become more refined and has revealed a wealth of therapeutic targets that may enable the disruption of angiogenesis or immunosuppression.

Areas covered: This review discusses the selective targeting of tumor-intrinsic pathways, therapies that target the GBM tumor microenvironment and relevant preclinical studies and their limitations. Relevant literature was derived from a PubMed search encompassing studies from 1989 to 2020.

Expert opinion: Despite appropriate target engagement, attempts to directly inhibit oncogenic pathways in GBM have yielded little success. This is likely attributed to the molecular heterogeneity of GBM and the presence of redundant signaling that allow for accumulation of adaptive mutations and development of drug resistance. Subsequently, there has been a shift toward therapies modulating the pro-angiogenic, immunosuppressive tumor microenvironment in GBM. The non-transformed cells in the microenvironment which includes endothelial cells, myeloid cells, and T cells, are presumably genetically stable, less susceptible to heterogeneity, and easier to target. This approach offers the highest potential for a therapeutic breakthrough in GBM.

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1. Introduction

Glioblastoma (GBM) is the most common malignant primary brain tumor. After diagnosis, median overall survival (OS) is 15 months despite the current standard of care with surgical resection followed by adjuvant chemoradiation [1,2]. Early clinical trials have primarily focused on delineating the optimal dose and combination of chemotherapy and radiotherapy in conjunction with surgery; however, there has been little progress beyond the 'Stupp protocol' [3]. Indeed, in the past 25 years, only 1 out of 11 phase III clinical trials for GBM showed an increase in overall survival [4]. A broader survey of 44 phase III clinical trials from 1966 to 2004 found only a 7 month improvement in overall survival (from 8 to 15 months) in the experimental groups [5]. This review will compare the two main approaches for targeting GBM - reversing tumorigenesis versus revitalization of the microenvironment - and highlight key preclinical findings. We searched pubmed.gov for the following search terms with various combinations: GBM, therapeutic targets, pathways, immunotherapy, macrophages, personalized medicine, vaccines, oncolytic viruses. We reviewed relevant papers from 1989 to 2020.

2. Selective targeting of oncogenic pathways

Detailed molecular characterization of GBMs identified common mutations in genes such as epidermal growth factor receptor (*EGFR*), tumor protein 53 (*TP53*), isocitrate dehydrogenase 1 (*IDH1*), neurofibromin 1 (*NF1*), and phosphatase and tensin homolog (*PTEN*) that define classical, mesenchymal, and proneural subtypes [6,7]. While pathways affected by these mutations have been targeted therapeutically, these attempts have met with little success.

2.1. EGFR

Recent sequencing data have shown that 57% of GBM show evidence of gain of function mutation and/or focal amplification of EGFR [8]. EGFR is activated by ligands such as EGF, transforming growth factor alpha, heparin-binding EGF-like growth factor, amphiregulin, epiregulin, betacellulin, and epigen [9]. Ligand binding induces receptor dimerization and autophosphorylation by the intracellular tyrosine kinase domain, resulting in recruitment of effector proteins and activation of downstream signaling cascades including phosphoinositide 3-kinase, mitogen-activated protein kinase, and signal transducer and activator of transcription 3 (STAT3) pathways. The most frequently occurring EGFR mutation in GBM, EGFRvIII, contains a deletion within the extracellular domain of the receptor that renders it constitutively active. Importantly, single cell sequencing studies showed that wild-type and mutant forms of EGFR are almost mutually exclusive, with only 1-2% of cells coexpressing wild type EGFR and EGFRvIII [10].

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Article Highlights

- GBM is a molecularly heterogeneous disease with signaling pathway alterations that vary at a single-cell level. This likely accounts for the clinical failures of therapeutic agents directed against single oncogenic pathways, such as peptide vaccine against EGFRvIII.
- The GBM tumor microenvironment is pro-angiogenic. While the role for the FDA approved VEGF inhibitor, bevacizumab, is still being defined, inhibition of integrin, Notch, and Wnt pathways are additionally being explored for anti-angiogenic effects in GBM.
- Immunosuppressive TAMs in GBM offer a wide range of therapeutic targets that affect their migration (e.g. LOX), polarization (e.g. STAT3), and phagocytic capacity. (e.g. CD47).
- Checkpoint inhibition in GBM has met with little success so far, but combination therapy with STING agonists, FGL2 inhibitors, or oncolytic viruses may boost T cell infiltration into GBM and render checkpoint inhibitors more effective.
- Ultimately, targeting the tumor microenvironment in GBM may offer the best chance for a therapeutic breakthrough.

This box summarizes key points contained in the article.

Small molecule inhibitors targeting the tyrosine kinase domain of EGFR have demonstrated efficacy in non-small cell lung cancer. However, clinical trials evaluating these agents in GBM showed disappointing results [11–13]. While tyrosine kinase inhibitors were capable of engaging its target, as evidenced by EGFR dephosphorylation in tumors of treated patients, its downstream signaling pathways were unaffected, suggesting that they are regulated by redundant mechanisms [14]. Indeed, EGFR pathway alterations have been shown to be associated with alterations in other receptor tyrosine kinases, providing an escape route from anti-EGFR therapeutic targeting [15–17]. Similarly, a peptide vaccine against EGFRvIII which showed initial promise in phase I and II trials ultimately demonstrated no survival benefit in a phase III randomized controlled trial [18-20]. Interestingly, 82% of vaccinated patients demonstrated loss of EGFRvIII expression upon GBM recurrence, suggesting a shift toward non-EGFR dependent oncogenic pathways following antigen loss [21].

2.2. IDH1

IDH1 is an enzyme that catalyzes the oxidative decarboxylation of isocitrate to alpha-ketoglutarate as part of the Krebs cycle. IDH1 mutations, most commonly R132 H, are genetic markers of secondary and proneural GBMs. They confer a gain of function in glioma cells, resulting in accumulation of an oncometabolite D-2-hydroxyglutarate (2-HG), which inhibits a key enzyme involved in histone modification and DNA methylation, leading to de-differentiation of GBM cells [22,23]. Consistent with this mechanism of action, IDH1 R132 H inhibitor reduced 2-HG production in IDH1 mutant glioma lines and impaired growth of IDH1 mutant glioma xenografts by promoting astrocytic differentiation, forming the basis for ongoing clinical trials [24]. IDH1 mutations also contain immunogenic epitopes that elicit specific CD4 and humoral responses, and preliminary results of phase I study of an anti- IDH R132 H peptide showed safety and sufficient immunogenicity [25,26].



Figure 1. Overview of the p53 signaling pathway and therapeutic approaches. A schematic of the main p53 signaling pathway is shown. (a). Hypoxia, UV radiation, ionizing radiation, and chemotherapy initiate DNA damage, which recruits ATM and ATR to phosphorylate CHK1 or CHK2, respectively. In unstressed cells, p53 is suppressed by MDM2 and MDM4. Additional downstream components of this pathway, feedback loops, and parallel p53 isoforms are not shown but also play key roles in modulating p53 and gliomagenesis. Several methods to target mutant p53 include genetherapy and vaccination to reintroduce wild-type p53 into cells with mutant p53. Other methods include p53-MDM2 targeted agents (e.g., nutilins, RITA) to interrupt the p53-MDM2 interaction to allow p53 to induce cell senescence. (b) CHK1, CHK2, and DNA damage activate p53 that binds to p53 response elements in the genome to upregulate a variety of genes in GBM. Furthermore, mutant p53 up- and down-regulates a distinct set of genes in GBM. A method targeting mutant p53 includes agents that attach to mutant p53 (e.g., PRIMA-1, PhiKan083, SCH529074, MIRA-3, STIMA-1) and revert it to a wild-type form. By normalizing wild-type p53 or otherwise disrupt the effects of mutant p53, these treatments may impact tumors. ATM ataxia telangiectasia mutated, ATR ataxia telangiectasia related, MIRA-3: mutant p53 reactivation and induction of rapid apoptosis, PRIMA-1 p53 reactivation and induction of massive apoptosis-1, RITA reactivation of p53 and induction of tumor cell apoptosis, STIMA-1: SH group targeting and induction of massive apoptosis. Reproduced with permission (England et al. Current understanding of the role and targeting of tumor suppressor p53 in glioblastoma multiforme, Tumor Biology. August 2013, Volume 34, Issue 4, pp 2063-2074).

2.3. p53

p53 is a classic tumor suppressor that regulates many genes involved in the cell cycle and apoptosis cascades (Figure 1). Inactivation of p53 occurs by a variety of mechanisms, including amplification of p53 inhibitors such as murine double minute (MDM) 2 or MDM4, deletion of p53 stabilizers such as p14/ARF, or mutation in the *TP53* gene which occurs in 85% of GBMs [8,27]. p53 deficiency confers a growth advantage of glioma cells and facilitates malignant transformation of primary cortical astrocytes [28,29]. Strategies attempting to restore the p53 pathway using gene therapy or pharmacological approaches in a variety of cancers, including GBM, have shown little clinical efficacy [30]. However, MDM2 inhibition has emerged as a promising option to restore the p53 pathway, albeit only in 8-10% of GBMs with MDM2 amplification and wild-type *TP53* [31]. Novel nutlin-based agents showed efficacy in MDM2-amplified, *TP53* wild-type GBM xenograft models, with adequate blood-brain barrier (BBB) penetration for clinical trials [32].

Mutant p53 is stabilized by chaperone activity of heat shock protein 90 which in turn is upregulated by histone deacetylase (HDAC) 6 by K294 deacetylation [33]. HDAC inhibitors such as suberoylanilide hydroxamic acid can therefore preferentially degrade mutant p53 and have shown anti-GBM activity in several studies [34–36].

2.4. GBM heterogeneity

Unfortunately, the efforts to target these oncogenic pathways have been hampered by tremendous molecular heterogeneity of GBM. Even individual cells within the same tumor exhibited mosaic expression of receptor tyrosine kinases and showed gene signature reflecting multiple glioblastoma subtypes [10]. Furthermore, cells from different locations of a multifocal GBM, as well as tumors from local and distant recurrences demonstrated divergent genetic profiles and varying drug responses [37]. Therefore, monotherapy targeting a single oncogenic pathway is unlikely to be beneficial in GBM, even with appropriate target engagement, because GBM will adapt to utilize other non-targeted mechanisms, conferring rapid drug resistance. One approach to address this problem is by tailoring treatment based on genetic makeup of the individual GBM [38]. However, this is prohibitively expensive and therapeutic agents targeting the unique genetic alterations may not be available. Another approach is to drive an immune response against a wide array of tumor antigens via dendritic cell vaccination, although the specific therapeutic targets are unclear [39-41].

Given these limitations due to GBM heterogeneity, we believe that the greatest potential for therapeutic breakthrough in GBM lies not in targeting the process of gliomagenesis *per se*, but its pro-angiogenic, immunosuppressive microenvironment. This strategy is unlikely to result in a cure, but may transform GBM into a manageable disease.

3. Selective targeting of tumor microenvironment

3.1. Angiogenesis

GBM is a highly vascularized tumor requiring extensive recruitment of blood vessels to combat hypoxia. Inhibition of vascular endothelial growth factor (VEGF) mediated angiogenesis is a therapeutic strategy at the center of several clinical trials for GBM [4,42,43]. However, a meta-analyses of 14 clinical trials found that these drugs do not improve overall survival (OS) in GBM, either as a single agent or in combination with chemotherapy. While anti-VEGF therapies such as bevacizumab, cediranib, and enzastaurin showed promising radiographic response rates and increased progression-free survival (PFS), they do not impact OS and are currently only offered as salvage therapy in refractory cases at our institution [44].

Besides VEGF, many other pathways and factors play important roles in the angiogenesis cascade. Integrins are membrane bound heterodimeric proteins involved in modulating the proangiogenic vs antiangiogenic microenvironment, activated by ligands of the extracellular matrix (ECM) [45]. Integrins $\alpha\nu\beta3$ and $\alpha\nu\beta5$ have been shown to be highly expressed on activated endothelial cells within the tumor compared to normal brain endothelial cells [46]. These ligated integrins have also been shown to regulate migration, invasion, and survival of endothelial cells. Disruption of the interaction between these integrins and their ligands in *in vitro* as well as *in vivo* promotes vascular regression, making them viable clinical targets [47,48]. Matrix metallopeptidases are also involved in these regulatory processes between the ECM, integrins, and endothelial cells by releasing proangiogenic factors into the ECM by proteolytic cleavage [45].

Other targets include the delta-like ligand 4/Notch pathway, shown to have a proangiogenic effect *in vivo*. A decoy delta-like ligand protein demonstrated reduced tumor angiogenic sprouting, vessel perfusion, pericyte coverage, and tumor growth, showing promise for clinical translation [49]. The Wnt family member 5A (WNT5A) pathway has been shown to mediate GBM stem cell (GSC) differentiation to endothelial cells, which recruit existing endothelial cells to support peritumoral satellite lesions [50]. These impactful preclinical studies show promise for the therapeutic targeting of these pathways.

3.2. Tumor-associated microglia and macrophages (TAMs)

The immune response to GBM is dominated by myeloid cells, consisting of CNS resident microglia and CNS infiltrating peripheral macrophages [51]. It is widely believed that these TAMs belie the immunosuppressive nature of the GBM microenvironment, reflected by the relative dearth of effector lymphocytes. Classically, TAMs were functionally classified into tumor suppressive M1 polarized phenotype in response to interferon γ (IFN γ) or toll-like receptor 4 ligands versus tumor supportive M2 polarized phenotype in response to IL4, IL-10, and/or IL-13 [52]. M2 polarized TAMs, defined by expression of cell surface markers such as CD206 and characterized by production of immunosuppressive IL-10, increases in number with glioma grade [52]. However, single cell sequencing of TAMs has revealed frequent co-expression of both M1 and M2 gene signatures in individual cells, suggesting that TAMs exhibit highly plastic phenotypes in vivo [53]. Classification of TAMs as M1 or M2 based on single marker such as CD206 may be inappropriate [54]. Nonetheless, this notion of M1/M2 TAM polarization has persisted as a useful conceptual framework, forming the basis for attempts to redirect TAMs to possess antitumor activity.

3.2.1. TAM reprogramming

Initial attempts to therapeutically reprogram TAMs in GBM targeted colony stimulating factor 1 receptor (CSF1R), critical for macrophage differentiation and survival. BBB-penetrant small molecule CSF1R inhibitors significantly increased survival in a proneural mouse model and slowed intracranial growth in patient-derived glioma xenografts by decreasing TAM accumulation, inhibiting M2 polarization, and increasing phagocytosis [55]. However, phase 2 trial demonstrated safety but no efficacy [56], which may be due to acquired macrophage expression of insulin-like growth factor 1 (IGF1) that provides

redundant signaling [57]. Future trials investigating CSF1R inhibitors in combination with other immunotherapy are being planned.

Other attempts to elicit M1 polarization have focused on intratumoral delivery of IL-12 using genetically modified viruses. Oncolytic herpes simplex virus (HSV) expressing IL-12 elicited M1 polarization, increased macrophage infiltration, and increased T effector to T regulatory ratios in xenograft and syngeneic models, which in combination with checkpoint inhibitors drastically improved survival [58]. Another study utilized replication incompetent adenovirus that can be induced to express IL-12 by administration of veledimex. In a syngeneic mouse glioma model, this approach led to preferential induction of IL-12 and IFN γ within the tumor, increased infiltration of effector lymphocytes, and tumor regression [59], and recent phase I trial using this approach showed promising results [60].

Another study showed that TAMs facilitate GBM by stimulating GSCs through pleiotrophin (PTN) secretion and activation of receptor protein tyrosine phosphatase receptor type (PTPRZ1). In addition, PTN and PTPRZ1 expression correlated with Cd11b+/CD163M2 TAMs and poor prognosis. Implantation of M2 macrophages without PTN inhibition promoted tumor growth, while implantation with simultaneous PTN inhibition abrogated the pro-tumorigenic activity of the M2 macrophages, highlighting the therapeutic potential of targeting this pathway in TAMs [61].

Accumulating data have demonstrated that STAT3 is a central mediator of immunosuppression in GBM [62-65]. STAT3 pathway is activated by various cytokines (e.g. IL-6 and IL-10) and growth factors (e.g. EGF), leading to the translocation of phosphorylated STAT3 into the nucleus and induction of target genes. In particular, STAT3 activation in TAMs limits their activation whereas STAT3 activation in GSCs promotes IL-10 secretion and inhibition of microglia/macrophage phagocytosis [66,67]. Given such a comprehensive role for STAT3 in the complex interplay between GBM and TAMs, STAT3 is an attractive therapeutic target. Indeed, STAT3 inhibitors were able to restore macrophage activation and their pro-inflammatory state in TAMs isolated from surgical specimens and peripheral blood mononuclear cells [63,65]. In addition, miR-124 inhibited STAT3 activation in vivo and showed promising efficacy in multiple preclinical glioma models [68].

3.2.2. Targeting TAM recruitment

TAM recruitment is mediated by chemokines such as CCL2 and fractalkine (CX3CL1). For instance, induction of CCL2 in a syngeneic rat GBM model promoted TAM accumulation and more aggressive tumor growth [69]. CCL2 has also been implicated in recruitment of monocytes that differentiate into immunosuppressive myeloid-derived suppressor cells (MDSCs) [70]. CX3CL1 signaling also promotes recruitment of TAMs, which is dependent upon CX3CL1 mediated CCL2 transcription and inhibited by the presence of the common V249I *CX3CR1* polymorphism [71,72]. Given the important role of CCL2 in TAM recruitment in GBM, targeting its receptor CCR2 is an attractive therapeutic strategy, with efficacy demonstrated in preclinical models [73].

Combined profiling and functional studies of PTEN deficient GBM models show that PTEN deficiency activates YAP1, which directly upregulates lysyl oxidase (LOX) expression. LOX acts as a chemoattractant for macrophages, which infiltrate and secrete osteopontin (SPP1), sustaining glioma cell survival and stimulating angiogenesis (Figure 2). LOX has been identified as a therapeutic target by a study showing that its inhibition suppresses tumor progression by abrogating macrophage infiltration [74]. SPP1 itself is a key driver of TAM recruitment in GBM, and its receptor integrin $\alpha v\beta 5$ is highly expressed by M2 polarized TAMs [75]. GSCs additionally secrete periostin that recruits M2 TAMs via the integrin $\alpha v\beta 3$ signaling, and disruption of this pathway inhibits glioma progression [76].

3.2.3. Enhancing TAM function

CD47 was identified as a 'don't eat me' signal on tumor cells that binds to SIRPa on macrophages leading to a signal cascade within the macrophages that inhibits phagocytosis [77]. Monoclonal antibodies blocking the CD47-SIRPa axis have shown efficacy treating GBM *in vitro* and *in vivo*, leading to phagocytosis of GBM cells by TAMs. The anti-CD47 treatment led to significantly higher levels of M1 macrophages in the tumor microenvironment, showing that anti-CD47 treatment either increases M1 polarization or enhances M1 macrophage recruitment from the periphery [78]. Despite the M1 bias, however, phagocytosis of tumor cells was enhanced in both M1 and M2 macrophages in response to CD47 blockade.

Recent studies have demonstrated that checkpoint inhibitors against programmed cell death protein 1 (PD-1) and programmed cell death ligand 1 (PD-L1), while traditionally thought to impact cytotoxic lymphocyte – tumor interaction, also affects TAM function as well. PD-1 is expressed by TAMs in a mouse model of colorectal cancer and in samples from colorectal cancer patients. PD-1 positive TAMs had a reduced capacity for phagocytosis of tumor cells, which was restored by PD-1/PD-L1 blockade, resulting in improved survival [79]. In an immunocompetent mouse GBM model, PD-L1 blockade enhanced macrophage phagocytosis of tumor cells and mediated radiation induced abscopal effect [80].

3.3. Checkpoint blockade

PD-1/PD-L1 checkpoint blockade has shown profound efficacy for malignancies such as non-small cell lung cancer, renal cancer, chronic Hodgkin's lymphoma, and gastric cancer [81-84]. PD-1 is expressed by activated T cells and its binding by PD-L1 tumor cells drives T cell apoptosis, anergy, and exhaustion which prevent cytotoxic T cell mediated tumor cell killing [85-87]. High levels of PD-L1 expression in tumors have been demonstrated to predict clinical efficacy of PD-1 inhibition [88-90]. Studies have shown that between 61% and 88% of GBM patients have tumors expressing PD-L1, indicating the potential use for PD-1 blockade in GBM treatment [91,92]. Despite promising success in preclinical studies, clinical trials have initially shown disappointing results for patients receiving PD-1 blockade therapy [93-98]. However, a recent randomized multi-institutional clinical trial showed that patients receiving neoadjuvant pembrolizumab had significantly increased overall survival compared to patients



Figure 2. Infiltrating macrophages secrete SPP1, which sustains glioma cell survival and stimulates angiogenesis. In PTEN-null GBM models, LOX inhibition markedly suppresses macrophage infiltration and tumor progression. Correspondingly, YAP1-LOX and β1 integrin-SPP1 signaling correlates positively with higher macrophage density and lower overall survival in GBM patients. This symbiotic glioma-macrophage interplay provides therapeutic targets specifically for PTEN-deficient GBM. Reproduced with permission (Chen et al. symbiotic macrophage-glioma Cell interactions reveal synthetic lethality in PTEN-null glioma. *Cancer Cell*. Volume 35, Issue 6, 10 June 2019, Pages 868–884.e6.).

randomized to receive adjuvant pembrolizumab. Tumor infiltrating lymphocyte density was shown to be associated with survival, while presurgical tumor volume, postsurgical tumor volume, percentage resection, gross total resection, and dexamethasone dosage at time of registration were not, indicating that the survival benefit is mediated by augmenting the preexisting immune response [99].

Cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) is another checkpoint receptor extensively studied for cancer immunotherapy. CTLA-4 inhibition increases CD28, which allows for sustained T-cell activation [100]. Combination treatment of IL-12 and anti CTLA-4 antibodies in a syngeneic mouse GBM model led to tumor regression even at advanced disease stages in which monotherapy failed. The concurrent treatment led to a significant decrease in forkhead box P3 (FoxP3) positive regulatory T cells and an increase in effector T cells [101].

In the last few years, a number of primary tumors metastatic to the CNS have seen a dramatic improvement in prognosis with the application of various immunotherapies. Most recently, the use of combined nivolumab and ipilimumab in patients with melanoma brain metastases was shown to confer a significant survival benefit [102]. Despite the exciting developments of immunotherapies for other primary malignancies, applications in GBM remain difficult. The unique immunosuppressive profile of GBM is ultimately the greatest challenge to immunotherapy development. Contributing factors are the low number of infiltrating T cells and expression of 'exhaustion' markers in T cells [102–104]. Additionally, the combination standard of care treatment for GBM including chemotherapy, temozolomide, radiotherapy, and corticosteroids is known to further contribute to immunosuppression [104].

Recently, stimulator of IFN genes (STING) agonists has emerged as a promising approach for increasing T cell infiltration into the GBM microenvironment, rendering the tumor more susceptible to checkpoint blockade. In GBM, STING is activated by double-stranded DNA from necrotic tumor cells, driving type I IFN production and upregulation of chemokines that recruit T cells. Intratumoral delivery of STING agonists elicited *Ccl5* and *Cxcl10* upregulation, increased the number of IFN γ producing CD8T cells, and extended survival in a mouse GBM model [105].

Other studies have shown that inhibition of FGL2 is another potential strategy to reverse immunosuppression in GBM and allow for more effective checkpoint blockade. Low levels of FGL2 expression along with high levels of granulocyte-macrophage colony-stimulating are associated with higher cytotoxic T cell infiltration and longer survival, while FGL2 overexpression increases CD4+ foxP3 regulatory T cells and M2 polarization of TAMs (Figure 3) [106,107]. In addition, FGL2 also inhibits dendritic cell maturation, antigen presentation capabilities, and ability to stimulate allogeneic T cell proliferation [107].

3.4. Oncolytic viruses – stimulating the anti-GBM immune response

Oncolytic viruses (OVs) are viruses that have been genetically modified to selectively replicate in and lyse tumor cells, emerging as a new treatment for GBM. Several OVs have entered clinical trials for patients with glioblastoma including herpes simplex virus, adenovirus, polio virus, measles virus, H-parvovirus, reovirus, and Newcastle disease virus [108].



Figure 3. Schematic of FGL2 function. FGL2 activates the FcyRIIB on antigenpresenting cells (APC) which can override PD1 (programmed cell death) blockade, resulting in T-cell inactivation. FcyRIIB blockade will subsequently result in T-cell activation in conjunction with anti-PD1 antibody therapy. Reproduced with permission (Patel et al. Fibrinogen-like protein 2: a potential molecular target for glioblastoma treatment. *Expert Opinion on Therapeutic Targets*. 23:8, 647–649, DOI:10.1080/14728222.2019.1628220). Traditionally, tumor eradication from oncolytic virotherapy was thought to be achieved through direct oncolysis. A recent paradigm shift in OV function now highlights the virally induced immune responses against the tumor that plays a critical role in their efficacy [109].

Combining checkpoint inhibitors with oncolytic viruses is an exciting approach that combines a promising immunotherapeutic strategy with the cytotoxic delivery system of oncolytic viruses. Despite promising results for checkpoint inhibitors in other brain malignancies [110], applications in GBM remain difficult. Possible contributing factors include poor penetration into the microenvironment through the BBB as well as low number of infiltrating T cells [102]. Saha et al. demonstrated the potential for this approach by engineering HSV to express IL-12 as well as the checkpoint inhibitors anti-PD-1 and anti-CTLA-4. This combination cured most mice in two separate glioma models. The authors showed that the treatment was associated with macrophage influx and M1-like polarization as well as increased T effector to T regulatory cell ratios, suggesting that microenvironment manipulation is necessary for the survival benefit [58]. Another study used HSV-1 armed with anti-PD1 antibodies to successfully treat two immunocompetent mouse models of GBM with a durable antitumor response rejecting second challenges of GBM implanted in the contralateral hemisphere [111].

Studies have also explored the use of viruses expressing immunotherapeutic agents other than checkpoint inhibitors. The recombinant polio:rhinovirus chimera virus (PVSRIPO) is currently in clinical trials for use against recurrent glioblastoma. A mechanistic study found that the virus activates the immune system by two separate mechanisms. Cytotoxic infection of malignant cells releases tumor-specific antigens for immune system priming. Additionally, sublethal infection of dendritic cells and macrophages yields type I interferondominant activation as well as development of tumor antigenspecific T cell responses [112]. G47∆ is another virus capable of modulating the immune system. G47A is a modified G207 virus with a47 gene deletion in order to disrupt the ICP47 protein, thus activating major histocompatibility complex class I antigen presentation [113]. Another study used HSV OV as a gene therapy vector carrying the human IL-12 cytokine to enhance T-cell-mediated immunologic effects [114,115]. Ultimately, the immunotherapeutic effects of OVs that stimulate an innate antiviral immune response and an adaptive antitumor T cell are now recognized to be essential to the success of this treatment strategy (Figure 4). Identifying viruses that produce robust immune responses in clinical trials and combining these viruses with neoadjuvant and adjuvant therapies will be central in the future of virus-based immunotherapy.

4. Conclusions

Traditional approaches that target oncogenic pathways that are intrinsic to the tumor have been met with limited success because of the molecular heterogeneity of GBM. Understanding and manipulating the GBM microenvironment is the key to addressing translational gaps. Abrogating the ability of GBM to sustain itself through angiogenesis and immunosuppression is a promising way to circumvent its defense mechanisms.



Figure 4. Schematic picture summarizing the events and factors related to successful oncolytic immunotherapy. From Martikainen and Essen, Virus-based immunotherapy of glioblastoma. Cancers (Basel). 5 February 2019. doi: 10.3390/cancers11020186.

5. Expert opinion

Despite decades of research, overall survival for patients diagnosed with GBM remains dismal. Detailed molecular characterization of GBM revealed several key oncogenic mutations. However, the efforts to target these tumor intrinsic pathways have met with little success, best exemplified by clinical failure of the peptide vaccine against EGFRvIII. Despite achieving its intended effect and driving EGFRvIII antigen loss, the agent failed to extend survival in a phase III trial. This highlighted the greatest barrier to targeting oncogenic pathways in GBM; that is, GBM exhibits tremendous amount of heterogeneity from each tumor cell to tumor cell, from each distinct region of the tumor to another, and between de novo and recurrent tumors. Any attempt to downregulate one particular aspect of gliomagenesis may simply allow for another protumoral signaling to predominate, rapidly resulting in drug resistance. Therefore, while other monotherapeutic agents targeting other oncogenic pathways such as IDH1 mutant inhibitors are in pre-clinical development, it is difficult to imagine that they will be able to overcome this problem. One attempt to circumvent the thorny issue of GBM heterogeneity is 'personalized therapy.' For instance, dendritic cells pulsed with the patient's own tumor lysate or multivalent peptide vaccines have been shown to invoke an anti-tumor immune response against multiple tumor antigens. Personalized therapy is incredibly expensive and given the relatively rare incidence of glioblastoma, it may be cost prohibitive to actualize. It remains to be seen whether this strategy will be effective, but we believe that another promising approach in GBM therapeutics is targeting of the GBM tumor microenvironment.

The cells that comprise the GBM microenvironment – endothelial cells, microglia/macrophages, and T cells – are presumably genetically stable and less impacted by heterogeneity as the tumor cells. In addition, signaling pathways in these nontransformed are more accurately and easily recapitulated in preclinical models than tumor-intrinsic pathways because they do not have to account for the complex GBM heterogeneity, and wide range of genetic tools such as cell-type-specific knockout or reporter mice are available, facilitating clinical translation.

Various therapeutic targets in TAMs affecting their recruitment, polarization, function, and interaction with T cells have been identified, placing this immunosuppressive cell type at the forefront of the quest for effective GBM treatment. For example, STAT3 inhibitors and viral mediated inducible delivery of IL-12 are already in clinical trials. In order to advance the field even further, a few gaps in knowledge need to be addressed. For instance, the relative contribution of CNS resident microglia versus peripheral macrophages is still unclear. Deep immunophenotyping of these two similar cell types using discriminatory markers such as TMEM119 or CCR2 within different GBM niches would address this question. In addition, characterization of TAMs should transition from strict M1/M2 dichotomy to a truly functional definition based, for example, on their secretome or phagocytic capacity.

There has also been burgeoning interest in checkpoint inhibitors for GBM, given the remarkable clinical efficacy of these drugs in several systemic malignancies and their CNS metastases. However, their application in GBM have been hampered by the immunosuppressive microenvironment characterized limited numbers of infiltrating T cells that express high levels of exhaustion markers. Fortunately, STING agonists, FGL2 inhibitors, or oncolytic viruses may be able to reinvigorate this adaptive effector response, and may render GBM susceptible to checkpoint inhibition when used as combination therapy. Ultimately, we believe the greatest potential for alleviating the devastating prognosis of GBM lies in better understanding and therapeutic modulation of the immunosuppressive microenvironment.

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References

Papers of special note have been highlighted as either of interest (+) or of considerable interest (++) to readers.

- Ostrom QT, Gittleman H, Farah P, et al. CBTRUS statistical report: primary brain and central nervous system tumors diagnosed in the United States in 2006-2010. Neuro Oncol. 2013 Nov;15(Suppl 2):ii1–56.
- Stupp R, Mason WP, van den Bent MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. N Engl J Med. 2005 Mar 10;352(10):987–996.
- This was the study that defined the current standard of care for glioblastoma. This paradigm is still in as of this writing.
- Stewart LA. Chemotherapy in adult high-grade glioma: a systematic review and meta-analysis of individual patient data from 12 randomised trials. Lancet (London, England). 2002 Mar 23;359(9311):1011–1018.
- Mandel JJ, Yust-Katz S, Patel AJ, et al. Inability of positive phase II clinical trials of investigational treatments to subsequently predict positive phase III clinical trials in glioblastoma. Neuro Oncol. 2018 Jan 10;20(1):113–122.
- Anderson E, Grant R, Lewis SC, et al. Randomized Phase III controlled trials of therapy in malignant glioma: where are we after 40 years? Br J Neurosurg. 2008 Jun;22(3):339–349.
- Wang Q, Hu B, Hu X, et al. Tumor evolution of glioma-intrinsic gene expression subtypes associates with immunological changes in the microenvironment. Cancer Cell. 2017 Jul 10;32(1):42–56.e6.
- Verhaak RG, Hoadley KA, Purdom E, et al. Integrated genomic analysis identifies clinically relevant subtypes of glioblastoma characterized by abnormalities in PDGFRA, IDH1, EGFR, and NF1. Cancer Cell. 2010 Jan 19;17(1):98–110.
- This paper defined the subtypes of glioblastoma in terms of their gene expression profile.
- 8. Brennan CW, Verhaak RG, McKenna A, et al. The somatic genomic landscape of glioblastoma. Cell. 2013 Oct 10;155(2):462–477.
- 9. Hynes NE, Lane HA. ERBB receptors and cancer: the complexity of targeted inhibitors. Nat Rev Cancer. 2005 May;5(5):341–354.
- Patel AP, Tirosh I, Trombetta JJ, et al. Single-cell RNA-seq highlights intratumoral heterogeneity in primary glioblastoma. Science (New York, NY). 2014 Jun 20;344(6190):1396–1401.
- •• This paper is of considerable interest because it demonstrates the intratumoral heterogeneity of GBM at the single cell level.
- van den Bent MJ, Brandes AA, Rampling R, et al. Randomized phase II trial of erlotinib versus temozolomide or carmustine in recurrent glioblastoma: EORTC brain tumor group study 26034. J Clin Oncol. 2009 Mar 10;27(8):1268–1274.
- Thiessen B, Stewart C, Tsao M, et al. A phase I/II trial of GW572016 (lapatinib) in recurrent glioblastoma multiforme: clinical outcomes, pharmacokinetics and molecular correlation. Cancer Chemother Pharmacol. 2010 Jan;65(2):353–361.
- Reardon DA, Groves MD, Wen PY, et al. A phase I/II trial of pazopanib in combination with lapatinib in adult patients with relapsed malignant glioma. Clin Cancer Res off J Am Assoc Cancer Res. 2013 Feb 15;19(4):900–908.
- 14. Hegi ME, Diserens AC, Bady P, et al. Pathway analysis of glioblastoma tissue after preoperative treatment with the EGFR tyrosine kinase inhibitor gefitinib-a phase II trial. Mol Cancer Ther. 2011 Jun;10(6):1102–1112.
- Stommel JM, Kimmelman AC, Ying H, et al. Coactivation of receptor tyrosine kinases affects the response of tumor cells to targeted therapies. Science (New York, NY). 2007 Oct 12;318(5848):287–290.
- Huang PH, Mukasa A, Bonavia R, et al. Quantitative analysis of EGFRvIII cellular signaling networks reveals a combinatorial therapeutic strategy for glioblastoma. Proc Natl Acad Sci U S A. 2007 Jul 31;104(31):12867–12872.

- Wykosky J, Hu J, Gomez GG, et al. A urokinase receptor-Bim signaling axis emerges during EGFR inhibitor resistance in mutant EGFR glioblastoma. Cancer Res. 2015 Jan 15;75(2):394–404.
- Sampson JH, Archer GE, Mitchell DA, et al. An epidermal growth factor receptor variant Ill-targeted vaccine is safe and immunogenic in patients with glioblastoma multiforme. Mol Cancer Ther. 2009 Oct;8(10):2773–2779.
- Sampson JH, Aldape KD, Archer GE, et al. Greater chemotherapy-induced lymphopenia enhances tumor-specific immune responses that eliminate EGFRvIII-expressing tumor cells in patients with glioblastoma. Neuro Oncol. 2011 Mar;13 (3):324–333.
- Weller M, Butowski N, Tran DD, et al. Rindopepimut with temozolomide for patients with newly diagnosed, EGFRvIII-expressing glioblastoma (ACT IV): a randomised, double-blind, international phase 3 trial. Lancet Oncol. 2017 Oct;18(10):1373–1385.
- Sampson JH, Heimberger AB, Archer GE, et al. Immunologic escape after prolonged progression-free survival with epidermal growth factor receptor variant III peptide vaccination in patients with newly diagnosed glioblastoma. J Clin Oncol. 2010 Nov 1;28 (31):4722–4729.
- Dang L, White DW, Gross S, et al. Cancer-associated IDH1 mutations produce 2-hydroxyglutarate. Nature. 2009 Dec 10;462(7274):739–744.
- 23. Lu C, Ward PS, Kapoor GS, et al. IDH mutation impairs histone demethylation and results in a block to cell differentiation. Nature. 2012 Feb 15;483(7390):474–478.
- Rohle D, Popovici-Muller J, Palaskas N, et al. An inhibitor of mutant IDH1 delays growth and promotes differentiation of glioma cells. Science (New York, NY). 2013 May 3;340(6132):626–630.
- 25. Platten M, Schilling D, Bunse L, et al. ATIM-33. NOA-16: A first-inman multicenter Phase I clinical trial of the German neurooncology working group evaluating a mutation-specific peptide vaccine targeting IDH1R132H in patients with newly diagnosed malignant astrocytomas. Neuro Oncol. 2018;20(suppl_6):vi8–vi9.
- 26. Schumacher T, Bunse L, Pusch S, et al. A vaccine targeting mutant IDH1 induces antitumour immunity. Nature. 2014 Aug 21;512 (7514):324–327.
- Frattini V, Trifonov V, Chan JM, et al. The integrated landscape of driver genomic alterations in glioblastoma. Nat Genet. 2013 Oct;45 (10):1141–1149.
- Sidransky D, Mikkelsen T, Schwechheimer K, et al. Clonal expansion of p53 mutant cells is associated with brain tumour progression. Nature. 1992 Feb 27;355(6363):846–847.
- 29. Bogler O, Huang HJ, Cavenee WK. Loss of wild-type p53 bestows a growth advantage on primary cortical astrocytes and facilitates their in vitro transformation. Cancer Res. 1995 Jul 1;55 (13):2746–2751.
- Duffy MJ, Synnott NC, Crown J. Mutant p53 as a target for cancer treatment. Eur J Cancer (Oxford, England: 1990). 2017 Sep;83: 258–265.
- Reifenberger G, Liu L, Ichimura K, et al. Amplification and overexpression of the MDM2 gene in a subset of human malignant gliomas without p53 mutations. Cancer Res. 1993 Jun 15;53 (12):2736–2739.
- Verreault M, Schmitt C, Goldwirt L, et al. Preclinical efficacy of the MDM2 inhibitor RG7112 in MDM2-amplified and TP53 wild-type glioblastomas. Clin Cancer Res off J Am Assoc Cancer Res. 2016 Mar 1;22(5):1185–1196.
- Scroggins BT, Robzyk K, Wang D, et al. An acetylation site in the middle domain of Hsp90 regulates chaperone function. Mol Cell. 2007 Jan 12;25(1):151–159.
- 34. Yin D, Ong JM, Hu J, et al. Suberoylanilide hydroxamic acid, a histone deacetylase inhibitor: effects on gene expression and growth of glioma cells in vitro and in vivo. Clin Cancer Res off J Am Assoc Cancer Res. 2007 Feb 1;13(3):1045–1052.
- Eyupoglu IY, Hahnen E, Buslei R, et al. Suberoylanilide hydroxamic acid (SAHA) has potent anti-glioma properties in vitro, ex vivo and in vivo. J Neurochem. 2005 May;93(4):992–999.
- 36. Ugur HC, Ramakrishna N, Bello L, et al. Continuous intracranial administration of suberoylanilide hydroxamic acid (SAHA) inhibits

tumor growth in an orthotopic glioma model. J Neurooncol. 2007 Jul;83(3):267–275.

- Lee JK, Wang J, Sa JK, et al. Spatiotemporal genomic architecture informs precision oncology in glioblastoma. Nat Genet. 2017 Apr;49(4):594–599.
- Chi AS, Batchelor TT, Kwak EL, et al. Rapid radiographic and clinical improvement after treatment of a MET-amplified recurrent glioblastoma with a mesenchymal-epithelial transition inhibitor. J Clin Oncol. 2012 Jan 20;30(3):e30–3.
- This paper is important because it demonstrates the feasibility of identifying novel malignant pathways from particular patients and using pathway specific therapies to extend survival.
- Hunn MK, Bauer E, Wood CE, et al. Dendritic cell vaccination combined with temozolomide retreatment: results of a phase I trial in patients with recurrent glioblastoma multiforme. J Neurooncol. 2015 Jan;121 (2):319–329.
- Prins RM, Cloughesy TF, Liau LM. Cytomegalovirus immunity after vaccination with autologous glioblastoma lysate. N Engl J Med. 2008 Jul 31;359(5):539–541.
- 41. Prins RM, Soto H, Konkankit V, et al. Gene expression profile correlates with T-cell infiltration and relative survival in glioblastoma patients vaccinated with dendritic cell immunotherapy. Clin Cancer Res off J Am Assoc Cancer Res. 2011 Mar 15;17(6):1603–1615.
- 42. Herrlinger U, Schafer N, Steinbach JP, et al. Bevacizumab plus irinotecan versus temozolomide in newly diagnosed O6-methylguanine-DNA methyltransferase nonmethylated glioblastoma: the randomized GLARIUS trial. J Clin Oncol. 2016 May 10;34(14):1611–1619.
- Gilbert MR, Dignam JJ, Armstrong TS, et al. A randomized trial of bevacizumab for newly diagnosed glioblastoma. N Engl J Med. 2014 Feb 20;370(8):699–708.
- This paper definitively showed that bevacizumab offered no survival benefit for newly diagnosed patients with glioblastoma.
- 44. Kaka N, Hafazalla K, Samawi H, et al. Progression-free but no overall survival benefit for adult patients with bevacizumab therapy for the treatment of newly diagnosed glioblastoma: a systematic review and meta-analysis. Cancers (Basel). 2019 Nov 4;11(11).
- 45. Seystahl K, Weller M. Is there a world beyond bevacizumab in targeting angiogenesis in glioblastoma? Expert Opin Investig Drugs. 2012 May;21(5):605–617.
- Gladson CL. Expression of integrin alpha v beta 3 in small blood vessels of glioblastoma tumors. J Neuropathol Exp Neurol. 1996 Nov;55(11):1143–1149.
- 47. Gladson CL, Cheresh DA. Glioblastoma expression of vitronectin and the alpha v beta 3 integrin. Adhesion mechanism for transformed glial cells. J Clin Invest. 1991 Dec;88(6):1924–1932.
- Chatterjee S, Matsumura A, Schradermeier J, et al. Human malignant glioma therapy using anti-alpha(v)beta3 integrin agents. J Neurooncol. 2000;46(2):135–144.
- Kangsamaksin T, Murtomaki A, Kofler NM, et al. NOTCH decoys that selectively block DLL/NOTCH or JAG/NOTCH disrupt angiogenesis by unique mechanisms to inhibit tumor growth. Cancer Discov. 2015 Feb;5(2):182–197.
- This paper is of importance because it is the first study to identify the Delta-like ligand 4/Notch pathway as a therapeutic target for clinical translation, identifying an alternative route to angiogenesis suppression.
- 50. Hu B, Wang Q, Wang YA, et al. Epigenetic activation of WNT5A drives glioblastoma stem cell differentiation and invasive growth. Cell. 2016 Nov 17;167(5):1281–1295.e18.
- 51. Hussain SF, Yang D, Suki D, et al. The role of human glioma-infiltrating microglia/macrophages in mediating antitumor immune responses. Neuro Oncol. 2006 Jul;8(3):261–279.
- Mantovani A, Sozzani S, Locati M, et al. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. Trends Immunol. 2002 Nov;23 (11):549–555.
- 53. Muller S, Kohanbash G, Liu SJ, et al. Single-cell profiling of human gliomas reveals macrophage ontogeny as a basis for regional differences in macrophage activation in the tumor microenvironment. Genome Biol. 2017 Dec 20;18(1):234.

- Gabrusiewicz K, Rodriguez B, Wei J, et al. Glioblastoma-infiltrated innate immune cells resemble M0 macrophage phenotype. JCI Insight. 2016;1(2):1-19.
- 55. Pyonteck SM, Akkari L, Schuhmacher AJ, et al. CSF-1R inhibition alters macrophage polarization and blocks glioma progression. Nat Med. 2013 Oct;19(10):1264–1272.
- This paper is of considerable importance because it is the first to identify tumor associated macrophages as powerful therapeutic targets in animal studies.
- 56. Butowski N, Colman H, De Groot JF, et al. Orally administered colony stimulating factor 1 receptor inhibitor PLX3397 in recurrent glioblastoma: an Ivy foundation early phase clinical trials consortium phase II study. Neuro Oncol. 2016 Apr;18(4):557–564.
- 57. Quail DF, Bowman RL, Akkari L, et al. The tumor microenvironment underlies acquired resistance to CSF-1R inhibition in gliomas. Science (New York, NY). 2016 May 20;352(6288):aad3018.
- Saha D, Martuza RL, Rabkin SD. Macrophage polarization contributes to glioblastoma eradication by combination immunovirotherapy and immune checkpoint blockade. Cancer Cell. 2017 Aug 14;32(2):253–267.e5.
- 59. Barrett JA, Cai H, Miao J, et al. Regulated intratumoral expression of IL-12 using a RheoSwitch therapeutic system((R)) (RTS((R))) gene switch as gene therapy for the treatment of glioma. Cancer Gene Ther. 2018 Jun;25(5–6):106–116.
- 60. Chiocca EA, Yu JS, Lukas RV, et al. Regulatable interleukin-12 gene therapy in patients with recurrent high-grade glioma: results of a phase 1 trial. Sci Transl Med. 2019 Aug 14;11(505):1-14.
- Shi Y, Ping YF, Zhou W, et al. Tumour-associated macrophages secrete pleiotrophin to promote PTPRZ1 signalling in glioblastoma stem cells for tumour growth. Nat Commun. 2017 Jun 1;8:15080.
- Doucette TA, Kong LY, Yang Y, et al. Signal transducer and activator of transcription 3 promotes angiogenesis and drives malignant progression in glioma. Neuro Oncol. 2012 Sep;14(9):1136–1145.
- Hussain SF, Kong LY, Jordan J, et al. A novel small molecule inhibitor of signal transducers and activators of transcription 3 reverses immune tolerance in malignant glioma patients. Cancer Res. 2007 Oct 15;67(20):9630–9636.
- 64. Wei J, Barr J, Kong LY, et al. Glioblastoma cancer-initiating cells inhibit T-cell proliferation and effector responses by the signal transducers and activators of transcription 3 pathway. Mol Cancer Ther. 2010 Jan;9(1):67–78.
- 65. Wu A, Wei J, Kong LY, et al. Glioma cancer stem cells induce immunosuppressive macrophages/microglia. Neuro Oncol. 2010 Nov;12(11):1113–1125.
- Lang R, Patel D, Morris JJ, et al. Shaping gene expression in activated and resting primary macrophages by IL-10. J Immunol (Baltimore, Md: 1950). 2002 Sep 1;169(5):2253–2263.
- 67. O'Farrell AM, Liu Y, Moore KW, et al. IL-10 inhibits macrophage activation and proliferation by distinct signaling mechanisms: evidence for Stat3-dependent and -independent pathways. Embo J. 1998 Feb 16;17(4):1006–1018.
- Ponomarev ED, Veremeyko T, Barteneva N, et al. MicroRNA-124 promotes microglia quiescence and suppresses EAE by deactivating macrophages via the C/EBP-alpha-PU.1 pathway. Nat Med. 2011 Jan;17(1):64–70.
- Platten M, Kretz A, Naumann U, et al. Monocyte chemoattractant protein-1 increases microglial infiltration and aggressiveness of gliomas. Ann Neurol. 2003 Sep;54(3):388–392.
- 70. Chang AL, Miska J, Wainwright DA, et al. CCL2 produced by the glioma microenvironment is essential for the recruitment of regulatory T cells and myeloid-derived suppressor cells. Cancer Res. 2016 Oct 1;76(19):5671–5682.
- Rodero M, Marie Y, Coudert M, et al. Polymorphism in the microglial cell-mobilizing CX3CR1 gene is associated with survival in patients with glioblastoma. J Clin Oncol. 2008 Dec 20;26 (36):5957–5964.
- 72. Lee S, Latha K, Manyam G, et al. Role of CX3CR1 signaling in malignant transformation of gliomas. Neuro Oncol. 2020 Apr 1:1-33.
- 73. Flores-Toro JA, Luo D, Gopinath A, et al. CCR2 inhibition reduces tumor myeloid cells and unmasks a checkpoint inhibitor effect to

slow progression of resistant murine gliomas. Proc Natl Acad Sci U S A. 2020 Jan 14;117(2):1129–1138.

- Chen P, Zhao D, Li J, et al. Symbiotic macrophage-glioma cell interactions reveal synthetic lethality in PTEN-null glioma. Cancer Cell. 2019 Jun 10;35(6):868–884.e6.
- 75. Wei J, Marisetty A, Schrand B, et al. Osteopontin mediates glioblastoma-associated macrophage infiltration and is a potential therapeutic target. J Clin Invest. 2019 Jan 2;129(1):137–149.
- Zhou W, Ke SQ, Huang Z, et al. Periostin secreted by glioblastoma stem cells recruits M2 tumour-associated macrophages and promotes malignant growth. Nat Cell Biol. 2015 Feb;17(2):170–182.
- Jaiswal S, Jamieson CH, Pang WW, et al. CD47 is upregulated on circulating hematopoietic stem cells and leukemia cells to avoid phagocytosis. Cell. 2009 Jul 23;138(2):271–285.
- Zhang M, Hutter G, Kahn SA, et al. Anti-CD47 treatment stimulates phagocytosis of glioblastoma by M1 and M2 polarized macrophages and promotes M1 polarized macrophages in vivo. PloS One. 2016;11(4):e0153550.
- 79. Gordon SR, Maute RL, Dulken BW, et al. PD-1 expression by tumour-associated macrophages inhibits phagocytosis and tumour immunity. Nature. 2017 May 25;545(7655):495–499.
- Ene CI, Kreuser SA, Jung M, et al. Anti-PD-L1 antibody direct activation of macrophages contributes to a radiation-induced abscopal response in glioblastoma. Neuro Oncol. 2019 Dec 3:1-13.
- Ledford H, Else H, Warren M. Cancer immunologists scoop medicine Nobel prize. Nature. 2018 Oct;562(7725):20–21.
- Rotte A, D'Orazi G, Bhandaru M. Nobel committee honors tumor immunologists. J Exp Clin Cancer Res. 2018 Oct 30;37(1):262.
- Chen YS, Shen CR. Immune checkpoint blockade therapy: the 2014 Tang prize in biopharmaceutical science. Biomed J. 2015 Jan-Feb;38(1):5–8.
- Xu F, Jin T, Zhu Y, et al. Immune checkpoint therapy in liver cancer. J Exp Clin Cancer Res. 2018 May 29;37(1):110.
- 85. Dai S, Jia R, Zhang X, et al. The PD-1/PD-Ls pathway and autoimmune diseases. Cell Immunol. 2014 Jul;290(1):72–79.
- Twyman-Saint Victor C, Rech AJ, Maity A, et al. Radiation and dual checkpoint blockade activate non-redundant immune mechanisms in cancer. Nature. 2015 Apr 16;520(7547):373–377.
- 87. Sharpe AH, Wherry EJ, Ahmed R, et al. The function of programmed cell death 1 and its ligands in regulating autoimmunity and infection. Nat Immunol. 2007 Mar;8(3):239–245.
- Weber JS, D'Angelo SP, Minor D, et al. Nivolumab versus chemotherapy in patients with advanced melanoma who progressed after anti-CTLA-4 treatment (CheckMate 037): a randomised, controlled, open-label, phase 3 trial. Lancet Oncol. 2015 Apr;16(4):375–384.
- 89. Taube JM, Klein A, Brahmer JR, et al. Association of PD-1, PD-1 ligands, and other features of the tumor immune microenvironment with response to anti-PD-1 therapy. Clin Cancer Res off J Am Assoc Cancer Res. 2014 Oct 1;20(19):5064–5074.
- Nghiem PT, Bhatia S, Lipson EJ, et al. PD-1 blockade with pembrolizumab in advanced merkel-cell carcinoma. N Engl J Med. 2016 Jun 30;374(26):2542–2552.
- Berghoff AS, Kiesel B, Widhalm G, et al. Programmed death ligand 1 expression and tumor-infiltrating lymphocytes in glioblastoma. Neuro Oncol. 2015 Aug;17(8):1064–1075.
- 92. Nduom EK, Wei J, Yaghi NK, et al. PD-L1 expression and prognostic impact in glioblastoma. Neuro Oncol. 2016 Feb;18(2):195–205.
- Buerki RA, Chheda ZS, Okada H. Immunotherapy of primary brain tumors: facts and hopes. Clin Cancer Res off J Am Assoc Cancer Res. 2018 Nov 1;24(21):5198–5205.
- 94. Hung AL, Maxwell R, Theodros D, et al. TIGIT and PD-1 dual checkpoint blockade enhances antitumor immunity and survival in GBM. Oncoimmunology. 2018;7(8):e1466769.
- Dejaegher J, Verschuere T, Vercalsteren E, et al. Characterization of PD-1 upregulation on tumor-infiltrating lymphocytes in human and murine gliomas and preclinical therapeutic blockade. Int J Cancer. 2017 Nov 1;141(9):1891–1900.
- 96. Wainwright DA, Chang AL, Dey M, et al. Durable therapeutic efficacy utilizing combinatorial blockade against IDO, CTLA-4, and

PD-L1 in mice with brain tumors. Clin Cancer Res off J Am Assoc Cancer Res. 2014 Oct 15;20(20):5290–5301.

- Magara A, Buhler R, Moser D, et al. First experience with MR-guided focused ultrasound in the treatment of Parkinson's disease. J Ther Ultrasound. 2014;2:11.
- 98. Reardon DA, Omuro A, Brandes AA, et al. OS10.3 randomized Phase 3 study evaluating the efficacy and safety of Nivolumab vs Bevacizumab in patients with recurrent glioblastoma: checkMate 143. Neuro Oncol. 2017;19(suppl_3):iii21-iii21.
- Cloughesy TF, Mochizuki AY, Orpilla JR, et al. Neoadjuvant anti-PD-1 immunotherapy promotes a survival benefit with intratumoral and systemic immune responses in recurrent glioblastoma. Nat Med. 2019 Mar;25(3):477–486.
- •• This study is of considerable interest because it is one of the first to show promising results using checkpoint inhibitors to treat glioblastoma.
- 100. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer. 2012 Mar 22;12(4):252–264.
- Vom Berg J, Vrohlings M, Haller S, et al. Intratumoral IL-12 combined with CTLA-4 blockade elicits T cell-mediated glioma rejection. J Exp Med. 2013 Dec 16;210(13):2803–2811.
- 102. Li B, Severson E, Pignon JC, et al. Comprehensive analyses of tumor immunity: implications for cancer immunotherapy. Genome Biol. 2016 Aug 22;17(1):174.
- 103. Lim M, Xia Y, Bettegowda C, et al. Current state of immunotherapy for glioblastoma. Nat Rev Clin Oncol. 2018 Jul;15(7):422–442.
- 104. Eil R, Vodnala SK, Clever D, et al. Ionic immune suppression within the tumour microenvironment limits T cell effector function. Nature. 2016 Sep 22;537(7621):539–543.
- 105. Ohkuri T, Ghosh A, Kosaka A, et al. STING contributes to antiglioma immunity via triggering type I IFN signals in the tumor microenvironment. Cancer Immunol Res. 2014 Dec;2(12):1 199–1208.
- Latha K, Yan J, Yang Y, et al. The role of fibrinogen-like protein 2 on immunosuppression and malignant progression in glioma. J Natl Cancer Inst. 2019 Mar 1;111(3):292–300.
- 107. Yan J, Zhao Q, Gabrusiewicz K, et al. FGL2 promotes tumor progression in the CNS by suppressing CD103(+) dendritic cell differentiation. Nat Commun. 2019 Jan 25;10(1):448.
- Foreman PM, Friedman GK, Cassady KA, et al. Oncolytic virotherapy for the treatment of malignant glioma. Neurotherapeutics. 2017 Apr;14(2):333–344.
- 109. Forsyth PA, Abate-Daga D. Oncolytic virotherapy for malignant gliomas. J Clin Oncol. 2018 May 10;36(14):1440–1442.
- 110. Tawbi HA, Forsyth PA, Algazi A, et al. Combined Nivolumab and Ipilimumab in melanoma metastatic to the brain. N Engl J Med. 2018;379(8):722–730.
- 111. Passaro C, Alayo Q, De Laura I, et al. Arming an oncolytic herpes simplex virus type 1 with a single-chain fragment variable antibody against PD-1 for experimental glioblastoma therapy. Clin Cancer Res off J Am Assoc Cancer Res. 2019 Jan 1;25(1):290–299.
- 112. Brown MC, Holl EK, Boczkowski D, et al. Cancer immunotherapy with recombinant poliovirus induces IFN-dominant activation of dendritic cells and tumor antigen-specific CTLs. Sci Transl Med. 2017 Sep 20;9(408):1-31.
- 113. Todo T, Martuza RL, Rabkin SD, et al. Oncolytic herpes simplex virus vector with enhanced MHC class I presentation and tumor cell killing. Proc Natl Acad Sci U S A. 2001 May 22;98(11):63 96–6401.
- 114. Patel DM, Foreman PM, Nabors LB, et al. Design of a Phase I clinical trial to evaluate M032, a genetically engineered HSV-1 expressing IL-12, in patients with recurrent/progressive glioblastoma multiforme, anaplastic astrocytoma, or gliosarcoma. Hum Gene Ther Clin Dev. 2016 Jun;27(2):69–78.
- 115. Parker JN, Gillespie GY, Love CE, et al. Engineered herpes simplex virus expressing IL-12 in the treatment of experimental murine brain tumors. Proc Natl Acad Sci U S A. 2000 Feb 29;97 (5):2208–2213.