

REVIEW

Open Access



Glioma drug development benefits from emerging phase 0 and window-of-opportunity trial paradigm

Shenzhong Jiang^{1*} and Chunlong Zhong^{1*}

Abstract

Clinical drug development is fundamentally difficult for rare and difficult-to-treat solid tumors, for example, glioma. Glioblastoma (GBM), an invariably fatal primary brain tumor, poses a significant challenge in the realm of effective treatments, necessitating an accelerated approach to innovative drug discovery. Investigators keep requiring a process toward obtaining more reliable early-stage signals related to drug activity and a process toward translating those signals into clinical benefits efficiently in late-stage drug development. Besides, these processes could increase the likelihood of benefit in late-stage settings at a lower cost and encourage more opportunities for drug development against other rare and difficult-to-treat cancers. Phase 0 and window-of-opportunity design has been advocated for glioma, aiming to identify and eliminate ineffective therapies early in the specific drug development process, thereby enhancing overall trial quality. However, challenges persist in implementing this trial design including obtaining pre-treatment samples, establishing accurate methodological platforms and biostatistical pipelines, and identifying novel biomarkers based on both clinical and multi-omics information to predict long-term drug responses. In this review, we encapsulate current evidence regarding the window-of-opportunity design in glioma, advocating for its recognition as a standard paradigm in new drug development.

Highlights

Duration between phase 2 to 3 trial on glioma is long and most phase 3 trials fail.
Phase 0 and window-of-opportunity studies should both fulfill PK and PD analysis.
Phase 0 and window-of-opportunity trial helps to avoid ineffective therapies and promote new drug discovery.
Conduction of phase 0 and window-of-opportunity trials needs a comprehensive coordination from a study team.
Obtaining pretreatment samples and identify surrogate biomarkers are still challenging.

Keywords Phase 0, Window-of-opportunity trial, Glioma, Early-stage trial, Drug development

*Correspondence:

Shenzhong Jiang
shenzhongj1@163.com

Chunlong Zhong
drchunlongzhong@tongji.edu.cn

¹Department of Neurosurgery, Shanghai East Hospital, School of Medicine, Tongji University, 150 Jimo Road, Shanghai 200120, China



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

Gliomas are the most common primary central nervous system (CNS) tumors and Glioblastoma (GBM) stands as the most lethal among them. Despite the application of standard and aggressive therapies, the overall survival (OS) for patients remains disheartening [1, 2]. Over the past two decades, the translation of findings from basic and preclinical research into promising treatment methods approved by the United States Food and Drug Administration (FDA) has been limited and challenging. While radiotherapy (RT), chemotherapy (temozolomide (TMZ)), and anti-angiogenesis therapy (bevacizumab) are conventionally utilized in glioma treatment, tumor-treating fields (TTFs) have emerged in clinical trials, showcasing extended OS. However, real-world cost-effectiveness concerns and cautious determination of specific populations are essential considerations. Although conventional immunotherapy, exemplified by immune checkpoint inhibitors (ICIs), has demonstrated remarkable efficacy in various solid tumors such as melanoma, non-small cell lung cancer (NSCLC), and urothelial carcinoma, its clinical benefits are less pronounced in glioma [3, 4]. The key reason is the immunosuppressive tumor microenvironment (TME) of glioma and intratumoral heterogeneity [5]. Addressing this heterogeneity and developing novel GBM-specific yet accurate models for personalized treatments are imperative for both preclinical and clinical studies [6, 7].

Given the deficiency in new drug discovery for GBM, there is ongoing debate regarding the need for improvement in the clinical trial landscape. Notably, the average duration from the initiation of phase II trials to the conclusion of phase III trials stands at 7.2 years, with over 91% of phase III trials for GBM proving unsuccessful [8–10]. In response, clinicians have instituted the early stages of phase 0 or window-of-opportunity trials to expeditiously identify potentially efficacious drugs for advancement to later-stage trials and elucidate their pharmacokinetic (PK) characteristics [8]. In this case, the timely exclusion of ineffective drugs ensures optimal cost-effectiveness. The judicious and ethical design of the study contributes to maintaining the quality of the trial. While phase 0 and window-of-opportunity trials exhibit similarities and disparities. Both can transpire before, during, or even subsequent to early-phase studies. However, a window-of-opportunity trial harbors therapeutic intent that aligns before, alongside, or immediately after phase 1 studies (Fig. 1). Its objective is to ascertain the maximum tolerated dose (MTD), ensure a comprehensive understanding of PK before formally establishing a treatment regimen, and ultimately decide whether to progress to further phase 1 or phase 2 trials.

Pharmacokinetics and pharmacodynamics pave the way for these early clinical trials

GBM evolves and propagates within an organ, impeding accessibility of tumor cells to most systematically administered agents due to the presence of the blood-brain barrier (BBB), which is constituted by an intricate network of endothelial and glial cells that obstruct the majority of systematically administered agents from reaching the CNS compartment [11, 12]. Insufficient penetration into the central nervous system is frequently cited as the primary cause of trial failures in neuro-oncology [13].

The assessment of PK and pharmacodynamics (PD) associated with a novel therapy, be it molecularly targeted or otherwise, constitutes the cornerstone of innovative therapy development. In the context of non-CNS cancers, the response of blood PK often serves as a robust indicator of drug exposure at the targeted site of disease, while PD markers in blood and/or tumor tissue offer confirmation of the anticipated metabolic or biological effects. In CNS cancers, paradoxically, PK values lack substantial significance as indicators for brain exposure since the majority of systematically administered therapeutics exhibit limited penetration into the brain [14]. Even purportedly “brain-penetrant” therapeutics have not consistently undergone evaluation for their efficacy in entering intact brain tissue, largely due to restricted assessments conducted within preclinical models involving the implantation of a tumor in the brain of a rodent [15].

Overview features of window-of-opportunity trials

The window-of-opportunity trials have several principles. The first is short treatment duration. Their treatment duration should be short and limited to a few days and weeks to avoid delaying curative treatment. Additionally, investigations would consider the PK and mechanisms of action of the compound to determine the final duration. In PK stage, the investigated compound should be administered long enough to reach a steady state to generate meaningful results. Modulation of phosphoproteins is generally achieved shortly after drug administration if the drug's blood levels are appropriate. However, for better evaluation of molecular profiles like gene/protein expression or individual immune response, a longer period maybe required. The ideal time to implement a window-of-opportunity design is during the preparatory phase between diagnosis and standard treatment. Therefore, the interval could be allocated for staging procedures, essential physical examinations and the planning of chemotherapy, radiotherapy or immunotherapy [16]. Although the interval between clinical diagnosis and standard treatment is not yet precisely defined, a time-frame of within four weeks is preferable [17, 18] (Fig. 1).

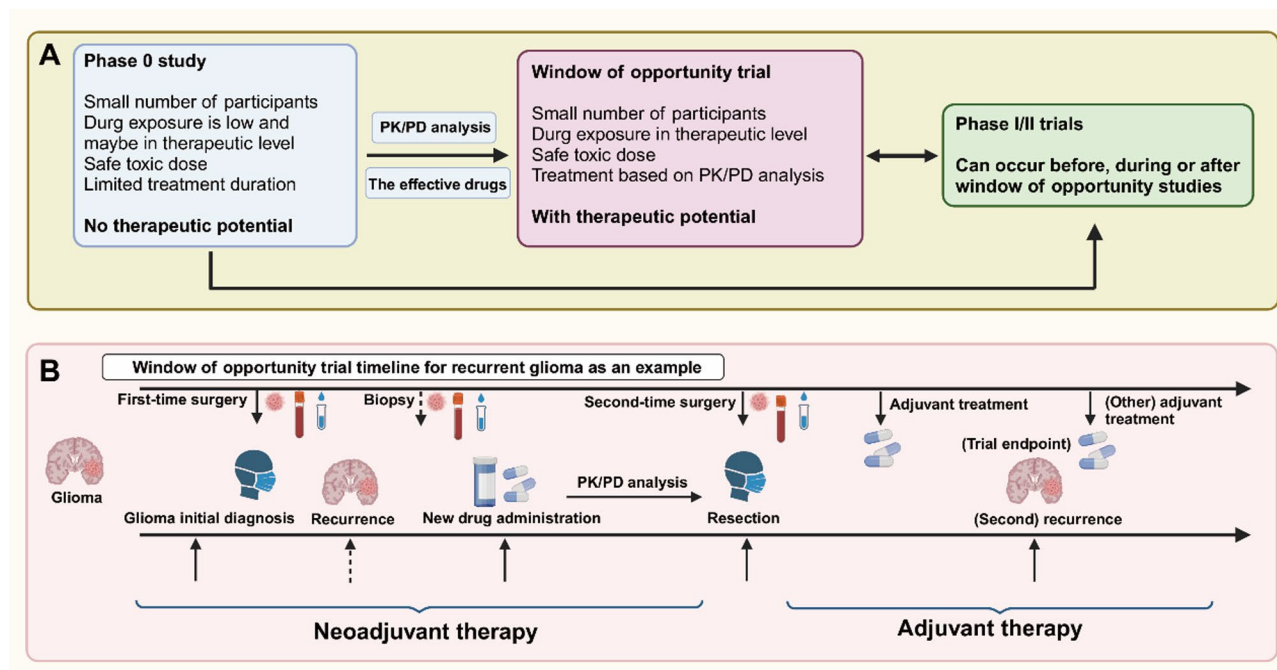


Fig. 1 The overview of phase 0 and window-of-opportunity study for glioma. (A) Phase 0 and window-of-opportunity studies exhibit both differences and similarities. Phase 0 studies involve minimal drug exposure, ensuring a safe toxic dose, while exposure in the window-of-opportunity trial may reach therapeutic levels, with limited treatment duration. The primary goal of a phase 0 study is not therapeutic. Effective drugs identified through PD/PD analysis may progress to a window-of-opportunity trial. In the latter, drug exposure is assessed at therapeutic levels, with careful consideration of safety margins. Treatment duration is determined based on prior PK/PD analyses. Both trials involve a small number of participants, given their early-stage nature. Typically, phase I/II studies follow phase 0, but they can occur before, during, or after window-of-opportunity studies. (B) We present the key procedures and sample collection in the development of a window-of-opportunity trial for recurrent glioma. Before the first recurrence, surgeons can obtain tumor and other samples during the initial surgery after diagnosis. Following the first recurrence, before the second resection, new drugs are administered based on PK/PD analysis, with the treatment duration determined. Biopsy samples may be collected during this phase. Adjuvant treatment follows, employing the same or different drugs until the second recurrence. This entire process highlights that the window-of-opportunity trial provides opportunities to obtain pre-new-drug treatment samples for further analysis or comparison. It heavily relies on PK/PD results and selectively advances drugs with therapeutic potential to subsequent stages or higher-level clinical trials, optimizing efficiency in terms of energy, time, and cost. Abbreviations: PK, pharmacokinetic; PD, pharmacodynamics

The second is primary endpoint should be a molecular or functional imaging parameter as a surrogate marker of treatment efficacy impacting OS and progression-free survival (PFS). The cut-off value for binary endpoints should be sufficiently precise to distinguish responders from non-responders, closely linked to clinical outcomes. While continuous endpoint like Ki-67 was also suitable for specific exploratory analyses, of note, Ki-67 cannot universally serve as a surrogate marker for all compounds and cancers [19, 20]. Other molecular endpoints, such as decreased phosphorylation of the targeted kinase receptor or modulation of specific cell cycle regulators, may also be promising candidates for surrogate markers when investigating compounds, though they still require standardization, validation, and central analysis [21, 22]. When the primary endpoint is based on comparing pre- and post-treatment biopsies, paired biopsies should be conducted under identical conditions and procedures to mitigate the effects of tumor heterogeneity and procedure-induced modifications. Pathological response,

combined with the quantification of viable residual tumor cells in the surgical specimen, could correlate with long-term outcomes and might serve as a valid endpoint [23]. The Response Evaluation Criteria in Solid Tumors (RECIST) criteria have been extensively utilized in clinical trials involving solid tumors; however, anatomic imaging endpoints are impractical due to the abbreviated treatment duration [16, 24–26]. Functional imaging modalities, such as 2-[fluorine-18]-fluoro-2-deoxy-D-glucose positron emission tomography (18 F-FDG-PET), also serve as primary endpoints to identify early metabolic alterations [16, 21]. Similarly, it is advisable to pre-define safety endpoints according to early stopping rules, with a particular emphasis on surgical safety assessed by an independent committee [16, 21]. Additionally, the toxicity profile, MTD, and recommended dose of the investigational compound should be thoroughly characterized before commencing the window study.

Moreover, they should incorporate endpoints encompassing molecular analyses such as

immunohistochemistry (IHC), staining, DNA/RNA investigations, gene expression profiling, or functional imaging techniques. Furthermore, certification of molecular analyses and quality control, along with a centralized review of imaging and standard treatment procedures, is indispensable. Functional imaging measurements heavily rely on imaging standardization levels. Therefore, imaging guidelines should be established beforehand to ensure standardized imaging acquisition [27]. Additionally, the proposed standard treatment must be clearly defined and subjected to rigorous quality control measures.

Overview features of phase 0 clinical trials

Phase 0 studies are acknowledged as pivotal in the development of targeted therapies, particularly where the therapeutic effect hinges on the successful delivery of the drug to the tissue target and instigating a PD response. Consequently, therapies that prove ineffective in modulating the tissue target and eliciting a PD response are promptly eliminated through phase 0 studies [28, 29]. In the realm of neuro-oncology, both “phase 0” and “window-of-opportunity” trials involve administering a potential drug for a brief duration between the “window” of treatment and surgery, followed by the collection of samples to evaluate PD effects. Despite some commonalities, there exist distinctions. Traditional phase 0 trials encompass non-therapeutic microdoses, whereas window-of-opportunity trials involve therapeutic doses albeit for a brief duration [30]. The use of therapeutic doses in window-of-opportunity trials is aimed at ensuring adequate drug penetrance, facilitating a successful assessment of the PD effects of the drug. In the context of neuro-oncology, a phase 0 trial can adopt a window-of-opportunity trial approach by prescribing a higher dose level to ensure BBB penetrance. Typically, the MTD is employed for a brief period to mitigate potential toxicities [31]. Due to the considerable similarities in design, phase 0 and window-of-opportunity trials are occasionally used interchangeably.

Overall, phase 0 trials entail the inclusion of a limited cohort exposed to a singular regimen of a low, non-toxic dosage of a pharmaceutical agent for a constrained duration (typically less than 7 days). The primary objective of phase 0 trials is to scrutinize the PK and PD characteristics of a pharmaceutical agent of considerable interest; typically, these trials do not harbor therapeutic intent. Similarly, but different, a window-of-opportunity trial assesses both PK and PD features while investigating doses and schedules postulated to contribute to a therapeutic effect. The transition from phase 0 to window-of-opportunity studies is plausible; when a drug in the phase 0 stage, or micro-dosing study, exhibits promise, the treatment can progress to the subsequent stage involving higher therapeutic doses.

Sample acquisition in phase 0/window-of-opportunity trials

A crucial procedural stage involves the acquisition of tumor samples prior to initiating treatment, serving as a biological control mechanism to comprehensively grasp the intricacies of the tumor landscape (Fig. 2). While challenges consist in these tissue-based studies, on one hand, samples are not uniformly obtained in accordance with contemporary neurosurgical practices preceding therapeutic interventions. On the other hand, the nomenclature employed to delineate the window-of-opportunity trials lacks uniformity, thereby contributing to technical disparities. Moreover, obtaining tumor tissues from recurrent diseases may appear ostensibly uncomplicated, the duration between sample acquisition and subsequent genotoxic therapies spans several weeks, if not months [32–34]. Evidently, the tumor immune profile, TME, cell state, and response sensitivity undergo substantial alterations during this interim [35]. In light of this, it is advisable to advocate and expand biopsies before the intervention. Stereotactic needle biopsies (SNBs) constitute a routine clinical practice, and although there exists some risk of intracranial hemorrhage, judicious patient stratification enables its performance as an outpatient procedure [36, 37].

Biopsies and surrogate tissues are consistently procured as a baseline before the administration of the investigational drug at a non-therapeutic dose. Subsequently, post-treatment blood samples can also be obtained for PK analyses. Prior to formal resection and post-treatment, there remain viable opportunities to acquire samples through SNB for CNS tumors, thereby facilitating the assessment of PK. For GBM patients, the sampled tissues exhibit notable diversities. For instance, cerebrospinal fluid (CSF) samples can be obtained as surrogate tissue to assess BBB penetrance [38]. During craniotomy, it is advisable to procure tumor samples from the BBB-permeable contrast-enhancing tumor, representing the most active tumor region in gliomas [39–41] as well as from the more diffuse and BBB-intact non-enhancing tumor to capture heterogeneity in the intratumoral TME and BBB penetrance [42].

If the time window between biopsy and resection is notably narrow, there may be insufficient time for an exhaustive investigation of the agent's bioactivity [43]. In window-of-opportunity trials, a pre-treatment biopsy can be reserved for patients exhibiting good physical function and possessing tumors smaller than 50 ml. The median doubling time for tumors typically spans 21–22 days [43]. Consequently, planned resection typically occurs 3–4 weeks subsequent to biopsy and drug administration within the designated time window. The median assessment of tumor growth may not adequately reflect the characteristics of all patients, particularly

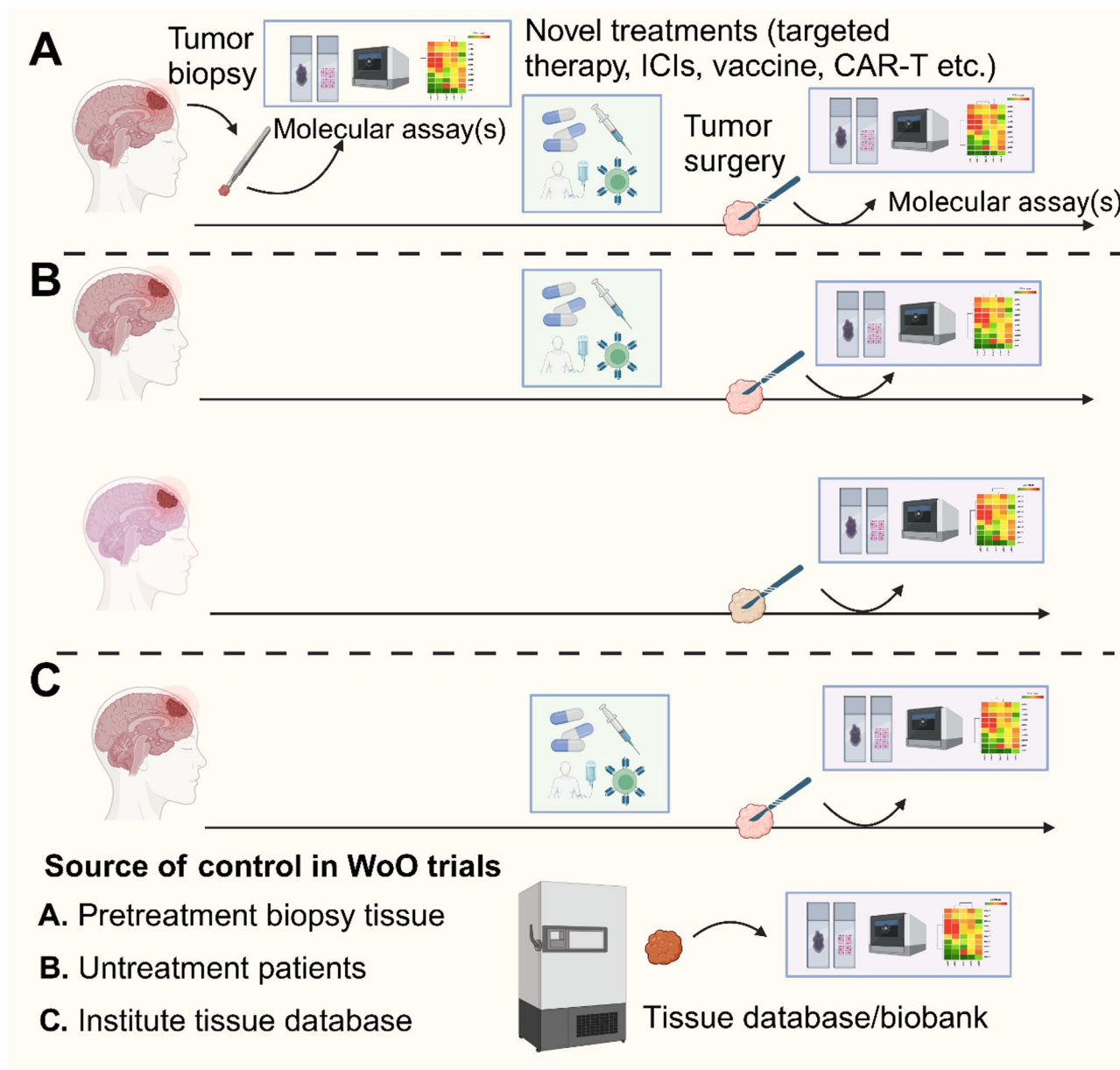


Fig. 2 The source of control for window-of-opportunity trial. (A) The control group is derived from pretreatment biopsy specimens. (B) The control group is from untreated patients. (C) The control group is from cohort/institute tissue biobank. Abbreviations: ICIs, immune checkpoint inhibitors; CAR-T, chimeric antigen receptor T cell; WoO, window-of-opportunity

those susceptible to rapid tumor growth. In such cases, prompt surgery accompanied by diagnostic imaging and cellular proliferation markers is imperative. Although biopsy facilitates sample acquisition before treatment and resection, given the evident intratumoral heterogeneity of gliomas, the biopsy tissues should be of sufficient size to enable accurate detection of histological features and to be representative. Additionally, comparing similar regions of the samples from biopsy and resection is advisable to mitigate the possibility that histological changes resulting from treatment are derived from intratumoral heterogeneity [44].

Molecular assay in phase 0/window-of-opportunity trials

In above part, it was stated that predefined molecular assay should be considered as a crucial component for the desired molecular outcome of window-of-opportunity studies, here we will give more details. When selecting the optimal assay, one fundamental assay that should be incorporated in all studies is a simple assessment of drug levels in the tissue. Indeed, a primary objective of window-of-opportunity trials is to evaluate the therapeutic concentration of the drug in the tumor following its administration at the dosage and schedule intended

for later-stage trials [45]. Moreover, molecular assays can determine whether the drug is interacting with its target(s), particularly when the target is known and an assay for assessing target engagement is available [46, 47]. It is noteworthy that many molecular assays utilized in preclinical laboratory testing can be applied in window-of-opportunity trials. These include IHC [48, 49] western blot (WB) [47] in-suit hybridization [50] cytometry by time-of-flight (CyTOF) [51] and single-cell RNA sequencing (scRNA-seq) [51, 52] (Fig. 3). Preclinical animal studies of the novel agent can establish assays for target engagement and potentially translate these findings into clinical specimens. Beyond verifying efficacy through enhancement in animal survival, another crucial application of preclinical studies is broadening the assay spectrum to clinical specimens [53].

Furthermore, the heterogeneity of these molecular events necessitates careful consideration of control tissue for comparing molecular outcomes. While some

molecular outcomes may not demand control tissue, the baseline levels of most molecular targets exhibit significant variability across tumors. Ideally, comparing untreated to treated tissue is desirable. However, as mentioned above, obtaining untreated tissue is challenging in patients with brain tumors, as it necessitates a biopsy before and after treatment. Many innovative approaches have been developed for acquiring tissues for molecular assays, and several of them have been explored in window studies.

Perioperative trials on brain tumors

For most patients with brain tumors, diagnostic tumor biopsies and safe maximal tumor resections typically occur during the same neurosurgical procedure, providing a limited window-of-opportunity for PD evaluation. However, many patients experience tumor recurrence and undergo a second surgery, as recurrent brain tumors often lack effective second-line treatments.

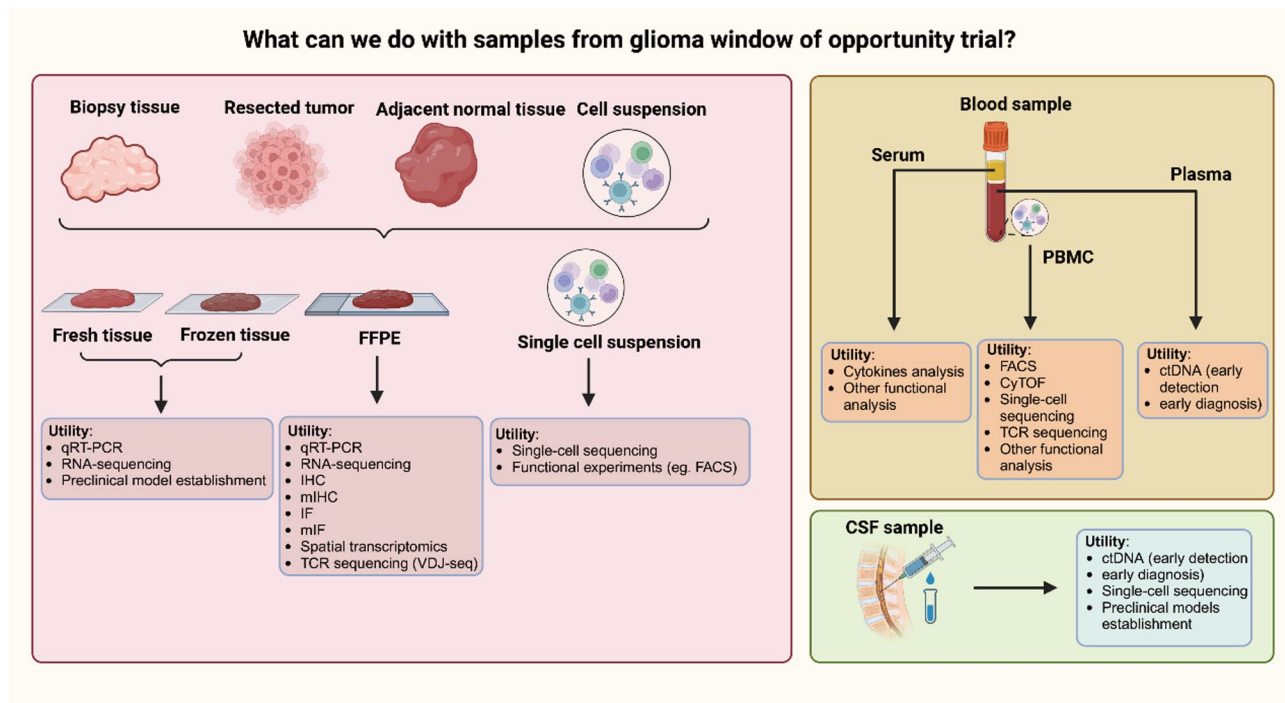


Fig. 3 Samples obtained from window-of-opportunity trial for sequencing and functional analysis. Window-of-opportunity studies provide unique opportunities to obtain multiple samples and tissues before, during, and after the administration of a target drug—an undertaking often challenging in most unresectable cancers. Tumor samples, biopsy tissues, adjacent normal tissue, and single-cell suspensions, both pre- and post-drug treatment, can be preserved in various manners tailored to desired downstream analyses. Specifically, resected tissues can be categorized as fresh tissues, frozen tissues, FFPE, and single-cell suspensions. Fresh, frozen, and FFPE tissues can facilitate sequencing techniques like RNA-seq, spatial transcriptomics, and TCR sequencing, as well as functional experiments such as RT-qPCR, IHC, mIHC, IF, mIF, and preclinical model establishment. Viable cryopreserved enzymatically digested single-cell suspensions enable single-cell sequencing and partial functional experiments like FACS. Peripheral blood samples, collected at sequential timepoints, can be fractionated into plasma, serum, and PBMCs. These samples are valuable downstream for ctDNA analysis, isolated cytokines, functional experiments including FACS and CyTOF, and single-cell sequencing, along with TCR sequencing. Notably, CSF, specific and accessible for brain tumor tissues, offers the identification of ctDNA for early detection and diagnosis. CSF samples also facilitate single-cell analysis and provide the potential for generating preclinical models using cells captured from CSF. Abbreviations: FFPE, formalin-fixed and paraffin-embedded; qRT-PCR, quantitative real-time PCR; IHC, immunohistochemistry; mIHC, multiplex immunohistochemistry; IF, immunofluorescence; mIF, multiplex immunofluorescence; TCR, T cell receptor; FACS, fluorescence activating cell sorter; CyTOF, cytometry by time of flight; ctDNA, circulating tumor DNA; PBMC, peripheral blood mononuclear cells; CSF, cerebrospinal fluid

Consequently, perioperative trials with investigational agents are commonly conducted during recurrence procedures. This presents a crucial opportunity for study participation and enables the assessment of efficacy surrogates such as OS and PFS. As previously mentioned, perioperative trials in neuro-oncology have progressed to randomized settings before surgical treatment [54–56]. The primary advantage of randomization is that patients in both arms have identical inclusion and exclusion criteria, ensuring comparable baseline characteristics before surgery. When analyzing tissue-based comparisons between different treatment arms, randomization instills a higher level of confidence in the results, irrespective of the sample size. Indeed, many investigational therapies necessitate combinations to target the immune suppressive TME or to infiltrate activated T cells into the tumor. Utilizing a neoadjuvant clinical trial platform approach could expedite evaluations, share controls, and harmonize inclusion and exclusion criteria, as well as tissue-based evaluations [57–60]. Beyond considerations for developing and validating PD assays, proper perioperative trials can effectively eliminate ineffective drugs that fail to penetrate into the tumor or engage their targets. Consequently, perioperative trials enhance enthusiasm for developing new agents by meticulously documenting target engagement and additional molecular and cellular effects.

Drugs selection and pharmacodynamics

Not all drugs under investigation are suitable for phase 0 trials, and not all phase 0 trials represent the first in human studies. Candidate drugs selected for phase 0 trials should adhere to the following criteria: (1) there exists an aspect of the drug's mechanism of action that remains unknown; (2) PD considerations are pivotal for the successful development of the drug; (3) preclinical investigations indicate an association between the drug target and an anti-tumor effect; (4) the investigated drug possesses a broad therapeutic window; (5) modulation of the drug target is expected at nontoxic doses and over a short duration of exposure (typically no longer than 7 days); (6) target modulation is likely to be discerned with a small sample size of patients (usually no more than 15 patients). The selection of “promising” agents for phase 0 studies should be evaluated in the specific context of the proposed agents. A phase 0 trial can be conducted for a single agent when it meets the following criteria: (1) evaluating drug efficacy through biopsies obtained pre- and post-exposure; (2) obtaining biomarkers associated with drug effects in tumors, blood, or other surrogate tissues; (3) determining a very safe but potentially effective starting dose based on the number of patients. In the case of combination treatment strategies, investigators often integrate two targeted agents, a targeted agent with

a conventional cytotoxic agent, or a targeted agent with novel immunotherapy. Additionally, phase 0 studies can assist in determining the relative schedule and sequence for monotherapy or combined therapy. Interestingly, “promising” agents can be defined in various ways, but they typically share common characteristics: (1) being first-in-class molecules; (2) having previously unexplored mechanisms in brain tumors; (3) demonstrating validated effects in diseases other than brain tumors; (4) theoretically, possessing mechanistic or toxicity characteristics well-suited for combined drug therapy. The process of drug selection is accompanied by the identification of a suitable biomarker-based PD assay to evaluate drug effects.

Obtaining tumor samples before drug exposure is particularly challenging for CNS tumors due to the heightened risks, costs, time consumption, and the absence of surrogate tissues corresponding to brain tumors. In such phase 0 trials, baseline comparator samples are typically set as prior drug exposure samples. Consequently, phase 0 studies for brain tumor patients to obtain planned biopsies before drug exposure are limited. While these limitations are likely unavoidable in phase 0 trials for brain tumor patients, it is advisable to optimize the study design by initially assessing PD endpoints in a reference population of matched samples from the initial diagnosis and recurrence. Only biomarkers demonstrating stable expressional and functional profiles should be selected as PD endpoints.

Phase 0/window-of-opportunity trials require more biostatistical considerations

Phase 0 trials typically involve limited sample sizes, posing challenges in estimating criteria that incorporate a PD response. In the initial introduction of phase 0 trials, Kummar et al. presented preliminary statistical guidelines and example designs for dichotomized PD, and subsequently [28] Murgo et al. refined the statistical designs by providing calculations to ensure the criteria of PD response along with analysis methods [29]. Rubinstein et al. later contributed to the field by generating new study design estimates for different PD response rates, introducing Simon Optimal and Minimax designs [61]. However, phase 0 trials for gliomas can be viewed as exploratory analyses, and as such, a prior power analysis to determine sample sizes is lacking [31, 42] [62– [64]. The justification for the low sample size lies in its feasibility [64]. Response criteria may be established based on prior preclinical or clinical data, or by simply assessing if there is a relative change compared to archived tissue without a predefined threshold (e.g., one-fold difference) [62, 63]. Nevertheless, the relatively recent development of phase 0 studies in GBM may be conducive to the exploration and implementation of novel statistical innovations.

Current available phase 0/window-of-opportunity trials on glioma

Drawing from a comprehensive review conducted by the Response Assessment in Neuro-Oncology (RANO) working group, the authors initially identified 22 publications pertinent to PD/PK analyses among GBM patients [42]. It is worth noting that not all of these publications align with the criteria of phase 0 or early phase 1 window-of-opportunity trials. A good example is a randomized perioperative phase 1, window-of-opportunity trial (NCT03343197) of 49 patients with isocitrate dehydrogenase-mutant (IDH-mt) gliomas, 2 types of IDH-inhibitors vorasidenib and ivosidenib were administered as trial agents to study dose-specific PD and IDH-related metabolic pathways [65]. There were 28 (+7) treatment window for the patients to be administrated with the intervention prior surgery and postoperative treatment was also allowed until disease progression or unacceptable toxicity. The concentration of 2-hydroxyglutarate (2HG) in resected tumors was the primary endpoint for the trial. The investigators found 2HG was significantly reduced in IDH inhibitors group over the control group, 92.6% in vorasidenib 50 mg q.d level and 91.1% in ivosidenib 500 mg q.d level respectively. Tumor/blood ratio was higher for vorasidenib vs. ivosidenib. Then the objective response rate (ORR) for vorasidenib 50 mg q.d was 42.9% vs. 10% for ivosidenib 10 mg q.d, 12.5% for ivosidenib 250 mg bid, 35.7% for ivosidenib 500 mg q.d. Both the IDH inhibitors were well-tolerated. The promising results were also instrumental in accelerating the success of final phase 3 clinical trial on vorasidenib [66]. While there are also some other examples of phase 0/window-of-opportunity trials on glioma [31, 63, 64]67– [79] (Table 1). The PerSurge (NOA-30) trial is a multicenter, phase 2, randomized, double-blind, placebo-controlled superiority window-of-opportunity study evaluating perampanel, an α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor inhibitor, in 66 progressive and recurrent GBM patients scheduled for surgical resection [74, 80, 81]. Patients are randomized 1:1 to receive perampanel (escalated to 10 mg/day) or placebo for 60 days (30 days pre- and post-surgery). Co-primary endpoints were to assess tumor-neuron connectivity via single nucleus RNA sequencing (snRNA-seq) and AI-based T2/ fluid-attenuated inversion recovery (FLAIR) magnetic resonance imaging (MRI) tumor growth rate changes pre-surgery. Secondary endpoints included contrast-enhanced MRI volumes, health-related quality of life (EORTC QLQ-C30/BN20), cognitive function (MMSE), seizure frequency, PFS, and OS. Besides, translational analyses would explore biomarkers, perampanel tissue concentration levels, and network connectivity via proteomics and sequencing. This trial is still ongoing and results would be expected

in the future. Conclusively, PerSurge tests perampanel's potential to disrupt neuron-glioma synapses hence to reduce tumor proliferation and invasion supported by preclinical evidence [80, 81]. Positive results could inform cancer neuroscience principles and support larger confirmatory trials for GBM network-targeted therapies. Balinda et al. in a phase 0 window-of-opportunity trial (NCT03995706) evaluated sacituzumab govitecan (SG) that selectively delivers SN-38 to tumors to inhibit tumor growth in 13 breast cancer with brain metastasis and 12 recurrent GBM patients undergoing craniotomy [75]. Patients received 10 mg/kg SG intravenously pre-operatively and resumed post-recovery on days 1 and 8 of 21-day cycles. The first aim was to assess intratumoral SN-38 concentration, the secondary aim was to investigate PFS, OS, and safety profile. Intracranial activity was confirmed in xenograft models. SG inhibited tumor growth and prolonged OS in xenografts. Median tissue SN-38 concentration was 104.5 ng/g for recurrent GBM, the PFS, OS and ORR was 2/9.5 months and 29% respectively. Grade ≥ 3 AEs included neutropenia (28%). SG exhibits robust tumor penetration and promising intracranial activity with acceptable toxicity, paving the way for phase II trial (NCT04559230) [75]. Another phase 1 trial (NCT01849146) evaluated Wee1 oral small-molecule inhibitor adavosertib combined with RT and TMZ in newly diagnosed GBM using a 3+3 dose-escalation design across concurrent, adjuvant, and combination cohorts ($n=57$) [76]. Intratumoral drug distribution (IDD) was assessed in recurrent GBM ($n=14$) via tissue homogenates and microdialysis. MTD were 200 mg/day (concurrent RT/TMZ) and 425 mg/day (adjuvant TMZ), but combination yielded unacceptable dose-limiting toxicities (50% rate). IDD showed superior penetration in contrast-enhancing (median 644–3576 ng/g) versus non-enhancing tissue. Median PFS was 8.5 months. Adavosertib exhibited excessive toxicity in concurrent settings; recommended phase II dose is 425 mg/day with adjuvant TMZ. Complementary PK informs future window-of-opportunity designs [76]. A recent phase 0 surgical window-of-opportunity trial (NCT03107780) evaluated the murine double minute homolog 2 (MDM2) inhibitor avtemadlin in 21 patients with recurrent TP53 wild-type GBM. Eligible patients received navtemadlin at 120 mg ($n=10$) or 240 mg ($n=11$) daily for 2 days before surgery, followed by 240 mg post-surgery until tumor progression or unaccepted toxicity [77]. The primary endpoint was set as assessing navtemadlin intratumoral concentration via liquid chromatography coupled with tandem mass spectrometry (LC-MS/MS), secondary endpoints included treatment safety, PD via IHC, RNA-seq, issue-based cyclic immunofluorescence (t-CyCIF) and survival data. Patient-derived GBM neurospheres were used for in vitro functional

Table 1 Current phase 0/early I, window-of-opportunity trials on glioma

Study	Status	Patients	Target drugs	Dose	Schedule and resection	Sample types	Drug level assessment	Biological relevance	Clinical finding and relevance	Registration number	References
Weingart et al., 2007	Finished	rGBM or anaplastic astrocytoma (<i>n</i> = 14)	Carmustine	120 mg/m ² , followed by an infusion of 30 mg/m ²	48 h before final resection	Blood, tumor	Blood and tumor drug concentration, AGT activity	To evaluate MGMT activity	Role of carmustine to inhibit MGMT	NA	[67]
Kuhn et al., 2007	finished	recurrent glioma (<i>n</i> = 6)	Temsirolimus	170–250 mg	1 dose 2 h before final resection	Blood, tumor	LC-MS/MS, blood and tumor drug concentration	Temsirolimus is an ester of sirolimus (rapamycin) that shows anti-glioma efficacy in cell line and orthotopic models	PK and PD analysis	NA	[68]
Rear-don et al., 2012	Finished	rGBM (<i>n</i> = 10)	Ridaforlimus	12.5 mg and 15 mg	4 doses before final resection	Blood, tumor	LC-MS/MS, blood and tumor drug concentration, mTOR activity assessment with p4E-BP1 in blood	pS6 kinase, downstream of mTOR	Efficacy assessed by imaging, safety profile	NA	[69]
Drapatz et al., 2013	Finished	rGBM, anaplastic astrocytoma and oligodendroglioma (<i>n</i> = 9)	GRN1005	30 mg/m ²	6 h before final resection	Blood, tumor	LC-MS/MS, blood and tumor drug concentration, WB to determine LRP-1 expression	GRN1005 binds to LRP-1 and enters brain effectively	Determine MTD, PK and PD analysis	NA	[70]
Xu et al., 2016	Finished	nGBM and anaplastic astrocytoma (<i>n</i> = 21)	RO4929097	10–20 mg daily	7 days before final resection	Both enhancing and non-enhancing tumor, blood	LC-MS/MS, qRT-PCR, IHC, morphometry, RNA sequencing, and FACS, blood and tumor drug concentration	Down-regulation of Notch signaling in enhancing tumor	PK and PD analysis in enhancing tumor and the tumor escape mechanism	NCT01119599	[71]
Sanai et al., 2018	Finished	rGBM (<i>n</i> = 20)	AZD1775	100, 200, 400 mg	4 h, 8 h, 24 h respectively before final resection	Blood, tumor	LC-MS/MS, IHC, DNA sequencing, blood and tumor drug concentration	markers of checkpoint disruption	determine MTD, PK and PD analysis	NCT02207010	[63]

Table 1 (continued)

Study	Status	Patients	Target drugs	Dose	Schedule and resection	Sample types	Drug level assessment	Biological relevance	Clinical finding and relevance	Registration number	References
Tien et al., 2019	Finished	rGBM (n = 12)	Ribociclib	900 mg	5 days daily before final resection	Both enhancing and non-enhancing tumor, blood	AGT activity, blood and tumor drug concentration	Decline in phospho-RB in half patients	Determine MTD, PK and PD analysis	NCT02933736	[31]
Quillin et al., 2020	finished	rGBM (n = 3)	ixazomib	4.0 mg	3 h before final resection	Blood and enhancing tumor	LC-MS/MS, blood and tumor drug concentration	Ixazomib is the second-generation proteasome inhibitor which showed efficacy against multiple myeloma	Safety profile	NCT02630030	[64]
Arrillaga-Romany et al., 2020	Finished	rGBM (n = 6)	ONC201	625 mg once a week	8 days before final resection	Blood and enhancing tumor	LC-MS/MS, tumor drug concentration, rat model validation in blood, tumor and other organs drug concentration	ONC201 as DRD2 antagonist penetrates BBB	PFS, OS, AEs, imaging data	NCT02525692 (part)	[72]
Moreno et al., 2023	Finished	high-grade glioma (n = 20)	Trotaresib	30 mg daily	on day 1–4 before surgery	Blood, CSF, both enhancing and non-enhancing tumor	Blood, tumor and CSF drug concentration, RNA-seq for markers investigation	The bromodomain and extraterminal protein inhibitor trotaresib shows antitumor activity to solid tumors	PK/PD analysis, safety profile, anti-tumor immunity, PFS	NCT04047303	[73]

Table 1 (continued)

Study	Status	Patients	Target drugs	Dose	Schedule and resection	Sample types	Drug level assessment	Biological relevance	Clinical finding and relevance	Registration number	References
Heuer et al., 2024	Unfinished	rGBM (n = 66)	Perampanel	2 mg and dose expansion till 10 mg without severe adverse events	30 days before surgery and 30 days after surgery	Blood, CSF, tumor	Imaging-based screen and molecular analysis	Perampanel is FDA approved AMPA inhibitor that reduce the interaction between glioma and neuron	tumor cell network and imaging-based tumor growth evaluation, adverse events, PFS and OS	2023-503938-52-00 30.11.2023	[74]
Fresendo et al., 2023	Unfinished	rGBM (n = 18)	Adipose-derived mesenchymal stem cells	2 ml fibrinogen, 1 ml thrombin, 1 ml cell mixture	Intra-cavitary treatment after surgery	Blood, CSF, tumor	Blood biochemistry, molecular analysis, ctDNA sequencing, sample IHC	Mesenchymal stem cells migrate and colocalize with tumor cells that remain in the tumor border; they are hypoimmunogenic and possess immunomodulatory properties	toxicity profile, OS and PFS	NCT05789394	[78]
Balinda et al., 2024	Finished	rGBM (n = 12)	Sacituzumab govitecan	10 mg sacituzumab govitecan	Given one day before surgery and continued on day 1, 8 and 21 after surgery	blood, CSF, tumor	PK/PD analysis with xenograft model data, sample immunohistochemistry, molecular analysis, high resolution mass spectrometry for blood, tumor, CSF samples	sacituzumab govitecan is an antibody-drug conjugate targets Trop-2 for selectively delivering SN-38	OS, PFS and AEs	NCT03995706	[75]
Cain et al., 2024	Unfinished	IDH1-mt glioma (n = 15)	safusidenib	250 mg safusidenib	250 mg safusidenib twice daily up to 28 days before surgery, and adjuvant safusidenib after surgery	blood, CSF, tumor	PK/PD analysis like PK concentrations, 2-HG changes and ctDNA analysis in blood and CSF, multi-omics sequencing, and other laboratory tests	Safusidenib is an oral IDH1 small-molecule inhibitor	AEs, imaging response, PFS, time to treatment failure, duration of response	NCT05577416	[79]

Table 1 (continued)

Study	Status	Patients	Target drugs	Dose	Sched- ule and resection	Sam- ple types	Drug level assessment	Biological relevance	Clinical finding and rel- evance	Registration number	Ref- er- enc- es
Lee et al., 2025	Finished	nGBM (n = 57)	adavosertib	200–425 mg (MTD) ada- vosertib daily	200 mg daily with or without RT/TMZ for 6 weeks and 425 mg MTD with adjuvant TMZ in three cohorts	Blood, tumor	PK/PD analysis, molecular analysis	Ada- vosertib (AZD1775) is a selective, ATP-com- petitive, small- molecule inhibitor of Wee1 kinase. The efficacy is demon- strated in preclinical models.	Investi- gate the MTD	NCT01849146	[76]
Rendo et al., 2025	Finished	rGBM (n = 21)	navtemad- lin	120 (n = 11)/240 mg (n = 10) navtemadlin daily	120/240 mg daily for 2 days before surgery and after surgery till tumor progression	Blood, tumor	PK/PD analysis, molecular anal- ysis, IHC, IF and multi-omics sequencing	Navtemad- lin is an oral small- molecule inhibitor for MDM2 that reac- tives TP53	OS and PFS	NCT03107780	[77]

Some phase 0/I trials are summarized from a previous RANO review (Ref [42]), we also updated the results with some new studies

Abbreviation: rGBM, recurrent glioblastoma; AGT, O6-alkylguanine-DNA alkyltransferase; MGMT, O6-methylguanine-DNA methyltransferase; LC-MS/MS, liquid chromatography coupled with tandem mass spectrometry; mTOR, mammalian target of rapamycin; p4E-BP1, phosphorylated 4E-BP1; LRP-1, lipoprotein receptor-related protein-1; DRD2, dopamine receptor D2; qRT-PCR, quantitative reverse transcription polymerase chain reaction; IHC, immunohistochemistry; WB, western blot; FACS, fluorescence-activated cell sorting; BBB, blood brain barrier; CSF, cerebrospinal fluid; OS, overall survival; PFS, progression-free survival; AEs, adverse events; PK, pharmacokinetics; PD, pharmacodynamics; FDA, food and drug administration; AMPA, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid; IDH1, isocitrate dehydrogenase 1; 2-HG, 2-hydroxyglutarate; ATP, adenosine triphosphate; MTD, maximum tolerated dose; MDM2, murine double minute homolog 2; RT, radiotherapy; TMZ, temozolomide; IF, immunofluorescence; NA, not available

assays, including apoptosis, BH3 profiling and scRNA-seq. Both doses achieved PD effects with median PFS of 3.1 months and OS of 10.0 months. Intratumoral drug concentrations exceeded 25 nM in $\geq 80\%$ of patients, with p53 pathway activation (e.g., CDKN1A upregulation) and reduced proliferation (Ki67 decrease). In functional assay, tumor cell death occurred in monotherapy process, combined therapy with TMZ enhanced apoptosis in TP53 wild-type, O[6]-methylguanine-DNA methyltransferase (MGMT)-methylated models while sparing bone marrow cells. Navtemadlin upregulated oligodendrocyte differentiation genes (e.g., OLIG2 enrichment at relapse). No TP53-inactivating mutations were acquired in three recurrent tumors. Navtemadlin penetrates GBM tissues and elicits PD responses at clinically achievable doses, inducing tumor cell death and adaptive oligodendrocyte-like states. Combined treatment with TMZ may enhance apoptosis and promote more durable survival benefits, warranting further clinical trials. Most incorporated studies primarily investigated the efficacy of novel drugs on recurrent tumors, justified by the feasibility of

obtaining pre- and post-resection samples. Over time, analyses expanded beyond tumor tissues to systematically include blood and CSF samples, extending PK and PD evaluations to these fluids to demonstrate comparability with prior results on investigational agents. This comprehensive approach assessed drug-specific target effects across multiple dimensions.

In addition to the studies outlined in Table 1, ongoing clinical trials such as NCT01849146 (phase 1), NCT02101905 (pilot study), NCT02133183 (phase 1), NCT02933736 (early phase 1), NCT03893903 (early phase 1), NCT03122197 (phase 0/1), and others are currently underway, exploring the concept of the window-of-opportunity. In conclusion, the phase 0 window-of-opportunity study design remains an underutilized strategy for the development of systemically administered therapeutics in neuro-oncology. Despite variations in PK and PD analysis approaches and challenges in obtaining samples before neoadjuvant and surgical treatments, the encouragement of tissue-based assessments for the biological effects of treatment is crucial in the early clinical

development of novel therapeutics. This approach facilitates the advancement of highly effective drugs to higher levels of clinical trials [82].

Utility of window-of-opportunity design in phase 1–2 trials

Given the evidence delineated in this review, numerous methodologies have been proposed for tailoring phase 1–2 early-stage trials to incorporate biological endpoints and achieve the objectives of window-of-opportunity trials for brain tumors. Phase 1 clinical investigations are traditionally perceived as the inaugural phase in the clinical advancement of novel drugs. The principal objective of a phase 1 trial is to evaluate the safety profile of the investigational drug, with a secondary focus on assessing its efficacy. Notably, the phase 1 study aims to determine the MTD by progressively escalating dose levels until reaching the pre-defined dose-limiting toxicity (DLT) rate [83]. 3+3 design is always adopted by the classic phase I trial due to its simplicity and accessibility. While an unexpectedly prolonged trial duration could emerge, and predicting the final sample size beforehand for optimal randomization poses a formidable challenge [84, 85].

Designs akin to those proposed for phase 1 trials can likewise be applied to phase 2 trials, acknowledging that these phase 2 trials maintain the objective of determining clinical efficacy while integrating biological endpoints. Ideally, patients would undergo a biopsy before investigational treatment and after which they are treated with therapeutic agent at defined dose in phase 1. Post-treatment specimens are feasible after surgical resection and can be compared with pre-treatment specimens. Molecular assays can be conducted to ascertain whether the agent continues to target its intended site and modifies the TME established in the Phase I setting. Once the agent consistently targets its intended site and elicits the desired molecular effects, it is considered effective [86, 87]. Efficacy analysis can be extended further by administering the experimental agent in the adjuvant setting after surgery to determine PFS and/or OS and recurrence [47]. The window-of-opportunity design in phase 2 trials provides significant biological insights in addition to standard clinical outcome data and informs decision-making regarding whether the agent merits further study. If the agent proves effective, it should progress to phase 3 trials; conversely, if the agent fails to target its intended site or elicit a clinical response, it may be abandoned. However, the window-of-opportunity approach may offer some insights at the molecular and cellular levels into the cause of failure. It is possible that the agent is only partially effective or that the defined dose is not as potent as anticipated in inducing a durable response. Therefore, careful evaluation and discussion of these molecular/cellular results are necessary before completely abandoning the

agent. Subsequently, trial design improvements should be made, the agent should be further developed, and novel targets should be pursued. If there is a phenomenon where the agent demonstrates efficacy in preclinical studies but fails to show robust clinical benefits, one hypothesis is that the target is not an independent driver of tumor growth. In this circumstance, combination therapy with other agents may prove effective, prompting the rational development of the agents in trials with combination arms. Overall, given the fact that new drug discovery efficiency is low for GBM and the median duration from phase 2 to phase 3 trials is quite long of which many fail to promote the new drug into real clinical approval, the window-of-opportunity trial design can be considered as a good paradigm to improve new drug development efficiency, success rate and reduce unexpected cost.

Lessons from phase 0/window-of-opportunity trials on future glioma clinical studies

Although phase 0 and window-of-opportunity studies accelerate the development for novel drug agents against glioma, challenges keep persisting. To optimize glioma drug development, interdisciplinary collaboration and novel technologies must be leveraged with and beyond these novel trial design. First of all, artificial intelligence (AI)-driven predictive modeling can optimize patient selection and dose-escalation strategies reducing trial duration and enhancing success rate [88, 89]. With the state-of-the art machine learning screening algorithms, investigators could identify novel glioma treatment candidates more effectively. Then aggregated big data, imaging data and/or real-world data from multi-centers on glioma patients' clinical response information will facilitate more tailored therapeutic approaches [90, 91]. Next, addressing the collaboration among neurosurgeons, oncologists and pharmacologists help to refine clinical trial methodologies and ensure translational success. Encouraging global partnerships in glioma study benefits to generate more diverse and representative patient cohorts. The diversity and heterogeneity limits representation of glioma patients in current trials, expanding recruitment internationally could enhance the generalizability of findings. Policymakers should also consider developing streamlined regulatory pathways to fast approve early-stage trials. In the future, Early-stage trials should focus on evaluating novel therapies' efficacy in glioma patients, particularly in combination with BBB-penetrating agents. Using gene editing tools such as CRISPR/Cas9 opens up new potentials for precise therapeutic interventions in gliomas [92–94]. Future trials could explore how gene-editing approaches can be incorporated into Phase 0 trials to test target modulation strategies. Innovatively, wearable devices capable of tracking patient biometrics, neurological function, and treatment

responses in real time could play a crucial role in future glioma clinical trials. Those kinds of technologies could provide a non-invasive means of assessing therapy efficacy over extended periods [95, 96].

What the neurosurgical oncologists do on phase 0 trials

Neuro-oncology phase 0 trials are experiencing a surge in the realm of clinical trials within neurosurgery, necessitating collaborative efforts from a substantial number of investigators [97, 98]. Notably, the neurological oncologist plays a pivotal role in the study design, patient accrual, and surgical phases [98]. A comprehensive understanding of the clinical and translational science trial elements is imperative for all team members and constitutes an essential aspect for the neurosurgeon. Initially, the neurosurgical oncologist assumes a critical role in patient selection and study consent [98]. In the context of phase 0 studies, it is crucial to conduct an assessment of tumor operability during the patient selection process. This determination must account for the timing of the planned surgery. Unlike other conventional clinical trials, brain tumor phase 0 studies necessitate a significant lead-in time prior to tumor resection. Molecular entry criteria are standard in phase 0 studies and typically involve 1 to 2 weeks of testing a patient's archived tumor tissues. Overall, eligible patients should be clinically stable, and the neurosurgical oncologist must evaluate the safety of timing an indicated operation to allow for trial pretesting and pretreatment. Timely PK analysis is imperative for the prompt acquisition of blood, CSF, and tumor tissue in phase 0 trials. Therefore, the feasibility of the sample achievement parameter heavily relies on operating room logistics and surgical timing. The key to fulfilling this requirement is to establish deliberate coordination with anesthesiologists, nurses, and surgical technologists, among other operating room personnel, to mitigate potential delays [98]. Furthermore, the neurosurgical oncologists should envisage the impact of data biases and tumor heterogeneity.

Conclusion

The phase 0 and window-of-opportunity clinical trial design, originally derived from the FDA and proposed for general drug development, encounters a slow pace in new drug development for brain tumors. This slowness is attributed not only to the aggressive nature of gliomas but also to challenges such as the absence of predictive animal models, risks associated with tumor acquisition, limitations in microdosing applicability, the presence of the BBB, substantial inter- and intratumoral heterogeneity, an incomplete understanding of the TME, and limited coordination among neurosurgeons, oncologists, and biostatisticians. Implementing these trial paradigms

in brain tumors proves to be an effective strategy for acquiring direct evidence of drug delivery and target modulation. In the future, more steps like utilizing novel AI technologies, incorporating multi-disciplinary cooperation and partnership and linking currently available data to large public, real-world databases should be thoroughly considered to expand the impact from phase 0 and window-of-opportunity trial designs on glioma research. Overall, phase 0 window-of-opportunity trials demonstrate unique advantages for discovering new drugs for gliomas. These innovative trial designs and PK/PD analyses promise more efficient and effective clinical trials, aiming to avoid ineffective therapies and accelerate the discovery of effective treatments for gliomas.

Abbreviations

CNS	Central nervous system
GBM	Glioblastoma
OS	Overall survival
FDA	Food and drug administration
TMZ	Temozolomide
TTF	Tumor-treating field
ICIs	Immune checkpoint inhibitors
NSCLC	Non-small cell lung cancer
TME	Tumor microenvironment
PK	Pharmacokinetic
BBB	Blood-brain barrier
PD	Pharmacodynamics
PFS	Progression-free survival
RECIST	Response evaluation criteria in solid tumors
18F-FDG-PET	[fluorine-18]-fluoro-2-deoxy-D-glucose positron emission tomography
SNBs	Stereotactic needle biopsies
CSF	Cerebrospinal fluid
IHC	Immunohistochemistry
WB	Western blot
CyTOF	Cytometry by time-of-flight
scRNA-seq	Single-cell RNA sequencing
RANO	Response assessment in neuro-oncology
IDH-mt	Isocitrate dehydrogenase-mutant
2HG	2-hydroxyglutarate
ORR	Objective response rate
AMPA	α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid
snRNA-seq	Single nucleus RNA sequencing
FLAIR	Fluid-attenuated inversion recovery
MRI	Magnetic resonance imaging
SG	Sacituzumab govitecan
IDD	Intratumoral drug distribution
MDM2	Murine double minute homolog 2
LC-MS/MS	Liquid chromatography coupled with tandem mass spectrometry
t-CyCIF	Issue-based cyclic immunofluorescence
MGMT	O6-methylguanine-DNA methyltransferase
MTD	Maximum tolerated dose
DLT	Pre-defined dose-limiting toxicity
AI	Artificial intelligence

Acknowledgements

The authors thank the findings revealed by the primary authors on new drug development on glioma and all the scholars in phase 0, window-of-opportunity trials in glioma.

Author contributions

S.J.: writing—original draft preparation; C.Z.: conceptualization, review and editing.

Funding

This study received grants from Shanghai Science and Technology Innovation Initiative, Yangfan Program (24YF2735100), the Key Disciplines Group Construction Project of Shanghai Pudong New Area Health Commission (PWZxq2022-10), the Key Discipline Construction Project of Shanghai East Hospital (2024-DFZD-003S) and the Shanghai East Hospital Youth Research Cultivation Funding (DFPY2024009).

Data availability

No datasets were generated or analysed during the current study.

Declarations

Competing interests

The authors declare no competing interests.

Received: 1 March 2025 / Accepted: 17 August 2025

Published online: 25 September 2025

References

- Ostrom QT, Price M, Neff C, et al. CBRUS statistical report: primary brain and other central nervous system tumors diagnosed in the united States in 2015–2019. *Neurooncology*. 2022;24:v1–95.
- Tan AC, Ashley DM, López GY, Malinzak M, Friedman HS, Khasraw M. Management of glioblastoma: state of the Art and future directions. *CA Cancer J Clin*. 2020;70:299–312.
- Pardoll DM. The Blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer*. 2012;12:252–64.
- Reardon DA, Brandes AA, Omuro A, et al. Effect of nivolumab vs bevacizumab in patients with recurrent glioblastoma: the checkmate 143 phase 3 randomized clinical trial. *JAMA Oncol*. 2020;6:1003–10.
- Wei J, Chen P, Gupta P, et al. Immune biology of glioma-associated macrophages and microglia: functional and therapeutic implications. *Neuro Oncol*. 2020;22:180–94.
- Neftel C, Laffy J, Filbin MG, et al. An integrative model of cellular states, plasticity, and genetics for glioblastoma. *Cell*. 2019;178:835–e849821.
- Louveau A, Herz J, Alme MN, et al. CNS lymphatic drainage and neuroinflammation are regulated by meningeal lymphatic vasculature. *Nat Neurosci*. 2018;21:1380–91.
- Singh K, Hotchkiss KM, Parney IF, et al. Correcting the drug development paradigm for glioblastoma requires serial tissue sampling. *Nat Med*. 2023;29:2402–5.
- Vanderbeek AM, Rahman R, Fell G, et al. The clinical trials landscape for glioblastoma: is it adequate to develop new treatments? *Neuro Oncol*. 2018;20:1034–43.
- Mandel JJ, Yust-Katz S, Patel AJ, et al. Inability of positive phase II clinical trials of investigational treatments to subsequently predict positive phase III clinical trials in glioblastoma. *Neuro Oncol*. 2017;20:113–22.
- Daneman R, Prat A. The blood-brain barrier. *Cold Spring Harb Perspect Biol*. 2015;7:a020412.
- Chernov AN, Alaverdian DA, Galimova ES, Renieri A, Shamova OV. The phenomenon of multidrug resistance in glioblastomas. *Hematology/ Oncology and Stem Cell Therapy*; 2021.
- Steeg PS. The blood–tumour barrier in cancer biology and therapy. *Nat Reviews Clin Oncol* 2021.
- Pardridge WM. The Blood-Brain barrier: bottleneck in brain drug development. *NeuroRx*. 2005;2:3–14.
- Summerfield SG, Yates JWT, Fairman DA. Free drug Theory – No longer just a hypothesis?? *Pharm Res*. 2022;39:213–22.
- Schmitz S, Hamoir M, Reychler H, et al. Tumour response and safety of cetuximab in a window pre-operative study in patients with squamous cell carcinoma of the head and neck. *Ann Oncol*. 2013;24:2261–6.
- Glimelius B, Lahn M. Window-of-opportunity trials to evaluate clinical activity of new molecular entities in oncology. *Ann Oncol*. 2011;22:1717–25.
- Farlow JL, Birkeland AC, Swiechicki PL, Brenner JC, Spector ME. Window of opportunity trials in head and neck cancer. *J Cancer Metastasis Treat* 2019;5.
- Leary A, Evans A, Johnston SRD, et al. Antiproliferative effect of lapatinib in HER2-Positive and HER2-Negative/HER3-High breast cancer: results of the presurgical randomized MAPLE trial (CRUK E/06/039). *Clin Cancer Res*. 2015;21:2932–40.
- Smith I, Robertson J, Kilburn L, et al. Long-term outcome and prognostic value of Ki67 after perioperative endocrine therapy in postmenopausal women with hormone-sensitive early breast cancer (POETIC): an open-label, multicentre, parallel-group, randomised, phase 3 trial. *Lancet Oncol*. 2020;21:1443–54.
- Owonikoko TK, Ramalingam SS, Miller DL, et al. A translational, pharmacodynamic, and Pharmacokinetic phase IB clinical study of everolimus in resectable Non–Small cell lung cancer. *Clin Cancer Res*. 2015;21:1859–68.
- Nogova L, Mattonet C, Scheffler M, et al. Sorafenib and everolimus in patients with advanced solid tumors and KRAS-mutated NSCLC: A phase I trial with early pharmacodynamic FDG-PET assessment. *Cancer Med*. 2020;9:4991–5007.
- Topalian SL, Forde PM, Emens LA, Yarchoan M, Smith KN, Pardoll DM. Neoadjuvant immune checkpoint blockade: A window of opportunity to advance cancer immunotherapy. *Cancer Cell*. 2023;41:1551–66.
- Litière S, Collette S, de Vries EG, Seymour L, Bogaerts J. RECIST - learning from the past to build the future. *Nat Rev Clin Oncol*. 2017;14:187–92.
- Schwartz LH, Seymour L, Litière S, et al. RECIST 1.1 - Standardisation and disease-specific adaptations: perspectives from the RECIST working group. *Eur J Cancer*. 2016;62:138–45.
- Schwartz LH, Litière S, de Vries E, et al. RECIST 1.1-Update and clarification: from the RECIST committee. *Eur J Cancer*. 2016;62:132–7.
- Boellaard R, Delgado-Bolton R, Oyen WJ, et al. FDG PET/CT: EANM procedure guidelines for tumour imaging: version 2.0. *Eur J Nucl Med Mol Imaging*. 2015;42:328–54.
- Kummar S, Kinders R, Rubinstein L, et al. Compressing drug development timelines in oncology using phase 0 trials. *Nat Rev Cancer*. 2007;7:131–9.
- Murgo AJ, Kummar S, Rubinstein L, et al. Designing phase 0 cancer clinical trials. *Clin Cancer Res*. 2008;14:3675–82.
- Aroldi F, Lord SR. Window of opportunity clinical trial designs to study cancer metabolism. *Br J Cancer*. 2020;122:45–51.
- Tien AC, Li J, Bao X, et al. A phase 0 trial of ribociclib in recurrent glioblastoma patients incorporating a tumor Pharmacodynamic- and Pharmacokinetic-Guided expansion cohort. *Clin Cancer Res*. 2019;25:5777–86.
- Magbanua MJM, Swigart LB, Wu HT, et al. Circulating tumor DNA in neoadjuvant-treated breast cancer reflects response and survival. *Ann Oncol*. 2021;32:229–39.
- Magbanua MJM, Brown Swigart L, Ahmed Z, et al. Clinical significance and biology of Circulating tumor DNA in high-risk early-stage HER2-negative breast cancer receiving neoadjuvant chemotherapy. *Cancer Cell*. 2023;41:1091–e11021094.
- Karschnia P, Smits M, Reifenberger G, et al. A framework for standardised tissue sampling and processing during resection of diffuse intracranial glioma: joint recommendations from four RANO groups. *Lancet Oncol*. 2023;24:e438–50.
- Touat M, Li YY, Boynton AN, et al. Mechanisms and therapeutic implications of hypermutation in gliomas. *Nature*. 2020;580:517–23.
- Dhawan S, Venteicher AS, Butler WE, Carter BS, Chen CC. Clinical outcomes as a function of the number of samples taken during stereotactic needle biopsies: a meta-analysis. *J Neurooncol*. 2021;154:1–11.
- Katzendobler S, Do A, Weller J, et al. The value of stereotactic biopsy of primary and recurrent brain metastases in the era of precision medicine. *Front Oncol*. 2022;12:1014711.
- Sanai N. Phase 0 clinical trial strategies for the neurosurgical oncologist. *Neurosurgery*. 2019;85:E967–74.
- Earnest t F, Kelly PJ, Scheithauer BW, et al. Cerebral astrocytomas: histopathologic correlation of MR and CT contrast enhancement with stereotactic biopsy. *Radiology*. 1988;166:823–7.
- Kelly PJ, Dumas-Duport C, Kispert DB, Kall BA, Scheithauer BW, Illig JJ. Imaging-based stereotaxic serial biopsies in untreated intracranial glial neoplasms. *J Neurosurg*. 1987;66:865–74.
- Kelly PJ, Dumas-Duport C, Scheithauer BW, Kall BA, Kispert DB. Stereotactic Histologic Correlations of Computed Tomography- and Magnetic Resonance Imaging-Defined Abnormalities in Patients With Glial Neoplasms. *Mayo Clinic Proceedings* 1987;62:450–459.
- Vogelbaum MA, Krivosheya D, Borghei-Razavi H, et al. Phase 0 and window of opportunity clinical trial design in neuro-oncology: a RANO review. *Neurooncol*. 2020;22:1568–79.

43. Müller DMJ, De Swart ME, Ardon H, et al. Timing of glioblastoma surgery and patient outcomes: a multicenter cohort study. *Neurooncol Adv*. 2021;3:vda053.
44. Karschnia P, Young JS, Dono A, et al. Prognostic validation of a new classification system for extent of resection in glioblastoma: A report of the RANO resect group. *Neuro Oncol*. 2023;25:940–54.
45. Weller M, Wen PY, Chang SM, et al. Glioma. *Nat Rev Dis Primers*. 2024;10:33.
46. Katti A, Vega-Pérez A, Foronda M, et al. Generation of precision preclinical cancer models using regulated in vivo base editing. *Nat Biotechnol*. 2024;42:437–47.
47. Vivanco I, Robins HJ, Rohle D, et al. Differential sensitivity of glioma- versus lung cancer-specific EGFR mutations to EGFR kinase inhibitors. *Cancer Discov*. 2012;2:458–71.
48. Lang FF, Bruner JM, Fuller GN, et al. Phase I trial of adenovirus-mediated p53 gene therapy for recurrent glioma: biological and clinical results. *J Clin Oncol*. 2003;21:2508–18.
49. Lang FF, Conrad C, Gomez-Manzano C, et al. Phase I study of DNX-2401 (Delta-24-RGD) oncolytic adenovirus: replication and immunotherapeutic effects in recurrent malignant glioma. *J Clin Oncol*. 2018;36:1419–27.
50. Marker DF, Pearce TM. Homozygous deletion of CDKN2A by fluorescence in situ hybridization is prognostic in grade 4, but not grade 2 or 3, IDH-mutant Astrocytomas. *Acta Neuropathol Commun*. 2020;8:169.
51. Greenwald AC, Darnell NG, Hofflin R, et al. Integrative Spatial analysis reveals a multi-layered organization of glioblastoma. *Cell*. 2024;187:2485–e25012426.
52. Spitzer A, Gritsch S, Nomura M, et al. Mutant IDH inhibitors induce lineage differentiation in IDH-mutant oligodendroglioma. *Cancer Cell*. 2024;42:904–e914909.
53. Jun HJ, Appleman VA, Wu H-J, et al. A PDGFR α -driven mouse model of glioblastoma reveals a stathmin1-mediated mechanism of sensitivity to vinblastine. *Nat Commun*. 2018;9:3116.
54. Cloughesy TF, Mochizuki AY, Orpilla JR, et al. Neoadjuvant anti-PD-1 immunotherapy promotes a survival benefit with intratumoral and systemic immune responses in recurrent glioblastoma. *Nat Med*. 2019;25:477–86.
55. Mellingshoff IK, Ellingson BM, Touat M, et al. Ivosidenib in isocitrate dehydrogenase 1-Mutated advanced glioma. *J Clin Oncol*. 2020;38:3398–406.
56. Mellingshoff IK, Bent MJvd, Blumenthal DT, et al. Vorasidenib in IDH1- or IDH2-Mutant Low-Grade glioma. *N Engl J Med*. 2023;389:589–601.
57. Mellingshoff IK, Cloughesy TF. Balancing risk and efficiency in drug development for rare and challenging tumors: A new paradigm for glioma. *J Clin Oncol*. 2022;40:3510–9.
58. Zhao B, Li H, Xia Y, et al. Immune checkpoint of B7-H3 in cancer: from immunology to clinical immunotherapy. *J Hematol Oncol*. 2022;15:153.
59. Zhao B, Wu J, Li H, et al. Recent advances and future challenges of tumor vaccination therapy for recurrent glioblastoma. *Cell Commun Signal*. 2023;21:74.
60. Ling AL, Solomon IH, Landivar AM, et al. Clinical trial links oncolytic immunostimulation to survival in glioblastoma. *Nature*. 2023;623:157–66.
61. Rubinstein LV, Steinberg SM, Kummer S, et al. The statistics of phase 0 trials. *Stat Med*. 2010;29:1072–6.
62. Batchelor TT, Gerstner ER, Ye X, et al. Feasibility, phase I, and phase II studies of tandutinib, an oral platelet-derived growth factor receptor- β tyrosine kinase inhibitor, in patients with recurrent glioblastoma. *Neuro Oncol*. 2016;19:567–75.
63. Sanai N, Li J, Boerner J, et al. Phase 0 trial of AZD1775 in First-Recurrence glioblastoma patients. *Clin Cancer Res*. 2018;24:3820–8.
64. Quillin J, Patel R, Herzberg E, et al. A phase 0 analysis of Ixazomib in patients with glioblastoma. *Mol Clin Oncol*. 2020;13:43.
65. Mellingshoff IK, Lu M, Wen PY, et al. Vorasidenib and Ivosidenib in IDH1-mutant low-grade glioma: a randomized, perioperative phase 1 trial. *Nat Med*. 2023;29:615–22.
66. Mellingshoff IK, van den Bent MJ, Blumenthal DT, et al. Vorasidenib in IDH1- or IDH2-Mutant Low-Grade glioma. *N Engl J Med*. 2023;389:589–601.
67. Weingart J, Grossman SA, Carson KA, et al. Phase I trial of Polifeprosan 20 with carmustine implant plus continuous infusion of intravenous O6-benzylguanine in adults with recurrent malignant glioma: new approaches to brain tumor therapy CNS consortium trial. *J Clin Oncol*. 2007;25:399–404.
68. Kuhn JG, Chang SM, Wen PY, et al. Pharmacokinetic and tumor distribution characteristics of Temozolomide in patients with recurrent malignant glioma. *Clin Cancer Res*. 2007;13:7401–6.
69. Reardon DA, Wen PY, Alfred Yung WK, et al. Ridaforolimus for patients with progressive or recurrent malignant glioma: a perisurgical, sequential, ascending-dose trial. *Cancer Chemother Pharmacol*. 2012;69:849–60.
70. Drappatz J, Brenner A, Wong ET, et al. Phase I study of GRN1005 in recurrent malignant glioma. *Clin Cancer Res*. 2013;19:1567–76.
71. Xu R, Shimizu F, Hovinga K, et al. Molecular and clinical effects of Notch Inhibition in glioma patients: A phase 0/I trial. *Clin Cancer Res*. 2016;22:4786–96.
72. Arrillaga-Romany I, Oda Y, Prabhu VV, et al. Biological activity of weekly ONC201 in adult recurrent glioblastoma patients. *Neuro Oncol*. 2019;22:94–102.
73. Moreno V, Manuel Sepúlveda J, Reardon DA, et al. Trovabresin, an oral potent bromodomain and extraterminal inhibitor, in patients with high-grade gliomas: A phase I, window-of-opportunity study. *Neuro Oncol*. 2023;25:1113–22.
74. Heuer S, Burghaus I, Gose M, et al. PerSurge (NOA-30) phase II trial of perampanel treatment around surgery in patients with progressive glioblastoma. *BMC Cancer*. 2024;24:135.
75. Balinda HU, Kelly WJ, Kaklamani VG, et al. Sacituzumab Govitecan in patients with breast cancer brain metastases and recurrent glioblastoma: a phase 0 window-of-opportunity trial. *Nat Commun*. 2024;15:6707.
76. Lee EQ, Alexander BM, Romo CG, et al. Phase I study of Adavosertib with radiotherapy and Temozolomide in newly diagnosed glioblastoma and intratumoral drug levels in recurrent glioblastoma. *Clin Cancer Res*. 2025;31:983–92.
77. Rendo V, Lee EQ, Bossi C, et al. A window-of-opportunity trial reveals mechanisms of response and resistance to Navtemadlin in patients with recurrent glioblastoma. *Sci Transl Med*. 2025;17:eadn6274.
78. Ramos-Fresnedo A, Al-Kharboosh R, Twohy EL, et al. Phase 1, dose escalation, nonrandomized, Open-Label, clinical trial evaluating the safety and preliminary efficacy of allogenic Adipose-Derived mesenchymal stem cells for recurrent glioblastoma: A clinical trial protocol. *Neurosurg Pract*. 2023;4.
79. Cain SA, Topp M, Rosenthal M, et al. A perioperative study of Safusidenib in patients with IDH1-mutated glioma. *Future Oncol*. 2024;20:2533–45.
80. Venkataramani V, Tanev DI, Strahle C, et al. Glutamatergic synaptic input to glioma cells drives brain tumour progression. *Nature*. 2019;573:532–8.
81. Venkatesh HS, Morishita W, Geraghty AC, et al. Electrical and synaptic integration of glioma into neural circuits. *Nature*. 2019;573:539–45.
82. Chen E, Ling AL, Reardon DA, Chiocca EA. Lessons learned from phase 3 trials of immunotherapy for glioblastoma: time for longitudinal sampling? *Neuro Oncol*. 2023.
83. Le Tourneau C, Lee JJ, Siu LL. Dose escalation methods in phase I cancer clinical trials. *J Natl Cancer Inst*. 2009;101:708–20.
84. Reiner E, Paoletti X, O'Quigley J. Operating characteristics of the standard phase I clinical trial design. *Comput Stat Data Anal*. 1999;30:303–15.
85. Yuan Y, Hess KR, Hilsenbeck SG, Gilbert MR. Bayesian optimal interval design: A simple and Well-Performing design for phase I oncology trials. *Clin Cancer Res*. 2016;22:4291–301.
86. Nassiri F, Patil V, Yefet LS, et al. Oncolytic DNX-2401 virotherapy plus pembrolizumab in recurrent glioblastoma: a phase 1/2 trial. *Nat Med*. 2023;29:1370–8.
87. Gállego Pérez-Llarraya J, García-Moure M, Labiano S, et al. Oncolytic DNX-2401 virus for pediatric diffuse intrinsic Pontine glioma. *N Engl J Med*. 2022;386:2471–81.
88. Yu K-H, Healey E, Leong T-Y, Kohane IS, Manrai AK. Medical artificial intelligence and human values. *N Engl J Med*. 2024;390:1895–904.
89. Abbasi AF, Asim MN, Dengel A. Transitioning from wet lab to artificial intelligence: a systematic review of AI predictors in CRISPR. *J Transl Med*. 2025;23:153.
90. Ngiam KY, Khor IW. Big data and machine learning algorithms for health-care delivery. *Lancet Oncol*. 2019;20:e262–73.
91. Wu WT, Li YJ, Feng AZ, et al. Data mining in clinical big data: the frequently used databases, steps, and methodological models. *Mil Med Res*. 2021;8:44.
92. Shalem O, Sanjana NE, Hartenian E, et al. Genome-Scale CRISPR-Cas9 knockout screening in human cells. *Science*. 2014;343:84–7.
93. Endo F, Kasai A, Soto JS, et al. Molecular basis of astrocyte diversity and morphology across the CNS in health and disease. *Science*. 2022;378:eadc9020.
94. Stadtmayer EA, Fraietta JA, Davis MM, et al. CRISPR-engineered T cells in patients with refractory cancer. *Science*. 2020;367:eaba7365.
95. Strain T, Wijndaele K, Dempsey PC, et al. Wearable-device-measured physical activity and future health risk. *Nat Med*. 2020;26:1385–91.
96. Kar A, Ahamed N, Dewani M, Awasthi L, Patil R, Banerjee R. Wearable and implantable devices for drug delivery: applications and challenges. *Biomaterials*. 2022;283:121435.

97. Lang FF, Asher A. Prospective clinical trials of brain tumor therapy: the critical role of neurosurgeons. *J Neurooncol.* 2004;69:151–67.
98. Sanai N. How to build a neurosurgical oncology practice specializing in gliomas. *Neurosurg Clin N Am.* 2019;30:129–36.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.