

Review

# Impact of Nanotechnology on Glioblastoma Studies: From Brain Stem Cells to Neoplastic Development and Cancer Gene Therapy

Maryam Raja<sup>1,2</sup>, Gabriela Quintero<sup>1,3</sup>, Yu-Chun Lone<sup>1,4</sup>, Jerzy Trojan<sup>1,4,5,\*</sup>

<sup>1</sup>CEDEA & ICGT (Center of Oncology Diagnostic and International Cancer Gene Therapy Foundation), 110010 Bogotá D.C., Colombia

<sup>2</sup>Department of Biotechnology, University of Isfahan, 81746-73441 Isfahan, Iran

<sup>3</sup>Faculty of Medicine, Autonomous University, Colombia (UNAB University), 681001 Floridablanca, Colombia

<sup>4</sup>INSERM UMR 1197, Cancer Center & University of Paris/Saclay, 94802 Villejuif, France

<sup>5</sup>National Academy of Medicine–ANM, 75272 Paris, France

\*Correspondence: [genetherapy@hotmail.fr](mailto:genetherapy@hotmail.fr) (Jerzy Trojan)

Academic Editor: Muthu Thiruvengadam

Submitted: 27 February 2025 Revised: 5 April 2025 Accepted: 29 April 2025 Published: 19 December 2025

## Abstract

The brain malignant tumor *Glioblastoma multiforme* (GBM) has a median survival of 14–16 months using current treatments; thus, understanding the pathology of GBM is crucial for proposing new therapies and increasing overall survival outcomes. Therefore, this study aimed to analyze different elements, particularly growth factors and the related signal transduction pathways, which play a role in brain neoplastic development, from stem cells to established solid brain tumors, and the application of current immunology techniques, molecular biology, and nanotechnology. Targeting growth factors, especially insulin-like growth factor-1 (IGF-I) (the principal neoplastic development factor) using anti-gene technologies—antisense and triple helix—has previously been shown to produce an immune anti-tumor response (CD8, CD28) through the TK/PI3K/AKT pathway. This immune response was increased using phytochemicals (phenolics), especially nanoparticles (theranostic nanoparticles), by modulating IGF-I through common pathways (IGF-I-R and TK/PI3K/AKT/TLR/MAPK and JAK/STAT). This review demonstrates how studies on central nervous system neoplastic development progressively led to establishing clinical cancer gene therapies, increasing GBM survival by 20–24 months. The presented studies compare the results of cancer gene therapy with other current immunotherapies. Moreover, this research chapter briefly describes the investigations of nanotechnology related to neurotumorigenesis and GBM therapies. The presented studies relate to nanotechnology and compare the results of cancer gene therapy with other current immunotherapies.

**Keywords:** cancer immuno–gene therapy; brain neoplastic development; stem cells; glioblastoma; IGF-I-induced signal transduction pathways; nanotechnology

## 1. Introduction

Glioblastoma multiforme (GBM) is the most common malignant brain tumor diagnosed in individuals aged 60 years (up to 15 cases per 100,000 people), with a predominantly low survival rate of less than a year [1–4]. The pathogenesis of GBM stems from genetic and growth factor pathway alterations (such as epidermal growth factor (EGF), transforming growth factor-beta (TGF $\beta$ ), insulin-like growth factor-1 (IGF-I)) [5–9]. Despite the ongoing therapeutic efforts, such as radiotherapy, chemotherapy, and recent adoptive T cell (ACT) therapy, GBM remains a lethal brain pathology [10–14]. Thus, understanding the important role of immune mechanisms in anti-tumor treatments is crucial for identifying specific immunomodulatory therapies for targeting growth factors, especially IGF-I or EGF [1,5,15–17]. The effectiveness of anti-IGF-I vaccines has offered promising results in treating GBM due to a strong anti-tumor immune response (TCD8, TCD28) [14,17].

This review aimed to present the research processes leading to central nervous system (CNS)-related tumor ther-

apy, from the hypothesis that neoplastic nervous system differentiation from stem cells to neuroglial differentiation is related to the presence of oncoproteins/growth factors, such as alpha-fetoprotein and IGF-I. Therefore, targeting these factors by suppressing the expression of related factors on a molecular level (transcription and translation) has promoted the development of a new form of cancer treatment: Cancer gene therapy [18]. The cancer gene therapy described in this review is based on anti-gene IGF-1 technology, targeting IGF-I in neoplastic glial cells. The IGF-I receptor induces the IGF-I signal transduction pathway, transforming GBM cells into immunogenic cell vaccines (expressing major histocompatibility complex class I (MHC1) and B7 antigens). This signal transduction pathway was further reinforced using phytochemicals related to IGF-I signaling elements (Toll-like receptor/mitogen-activated protein kinase/nuclear factor kappa-light-chain-enhancer of activated B cells (TLR/MAPK/NF $\kappa$ B)) [18,19]. Moreover, nanotechnology (common TK/PI3K/AKT elements) has recently demonstrated that the IGF-I signal is overexpressed, thus strengthening the immunogenicity of vaccines [20]. In this context, nanotechnology constitutes the most promis-



ing approach to the anti-IGF-I cancer gene therapy employed for GBM [18,21]. Meanwhile, cancer gene therapy can be used in parallel with stem cell therapy [22]. Therefore, our review largely refers to current research on stem cells differentiating into neuroglial cells, followed by mature glial cells, the target of our cancer gene therapy. The neurogenesis leading to glial cells is also described in relation to nanotechnology, which is included in different gene types and immunotherapies [23–25].

## 2. Brain Tumors and Therapy

### 2.1 CNS Malformations and Tumors

Genetic and environmental causes can promote the induction of CNS malformations and tumors [26]. The neurohistogenesis of malformations is closely related to embryonal tumors. Primitive neuroectodermal tumors (PNETs) were classified as central neuroblastoma, ependymoblastoma, medulloblastoma, and supratentorial PNETs—Medulloepithelioma and atypical teratoid/rhabdoid tumors [2,3]. The mouse teratocarcinoma model was employed to study the histogenesis of neuroectodermal tumors behind CNS neoplastic development and their transformation into solid tumors, such as glioblastoma. The histogenesis of teratocarcinoma, derived from pluripotent carcinoma cell line 3 (PCC3) and PCC4 embryonal carcinoma cell lines, reproduced normal CNS development (Fig. 1) [18,27–30], and, in parallel, the different stages of abnormal nervous tissue histogenesis, which can be presented as follows: The neuroectoblast starts as embryonal carcinoma stem cells, which transform into neuroblastic vesicles mimicking the successful development of a neural tube.

These so-called neuroblastic structures follow different stages: (1) Undifferentiated carcino-embryonic structures; (2) medulloepithelial structures (composed of a mixture of ectoblastic and neuroectoblastic components); (3) neuroblastic structures; (4) neuroepithelial structures [2,27,28,31]. These structures present either a diffused growth of neuroblastic stem cells or a mixture of neuronal and glial precursors. Final differentiation results in encephaloid tissue (a positive biomarker in staining for alpha-fetoprotein (AFP), serum albumin (SA), and IGF-I) [27]. Implementation of the mouse teratocarcinoma model, containing stem cells and neuroglial structures, is useful in understanding human CNS tumors, which can differentiate into both neuronal and glial lineages [22,32,33], as well as in GBM-related cancer gene therapy [18].

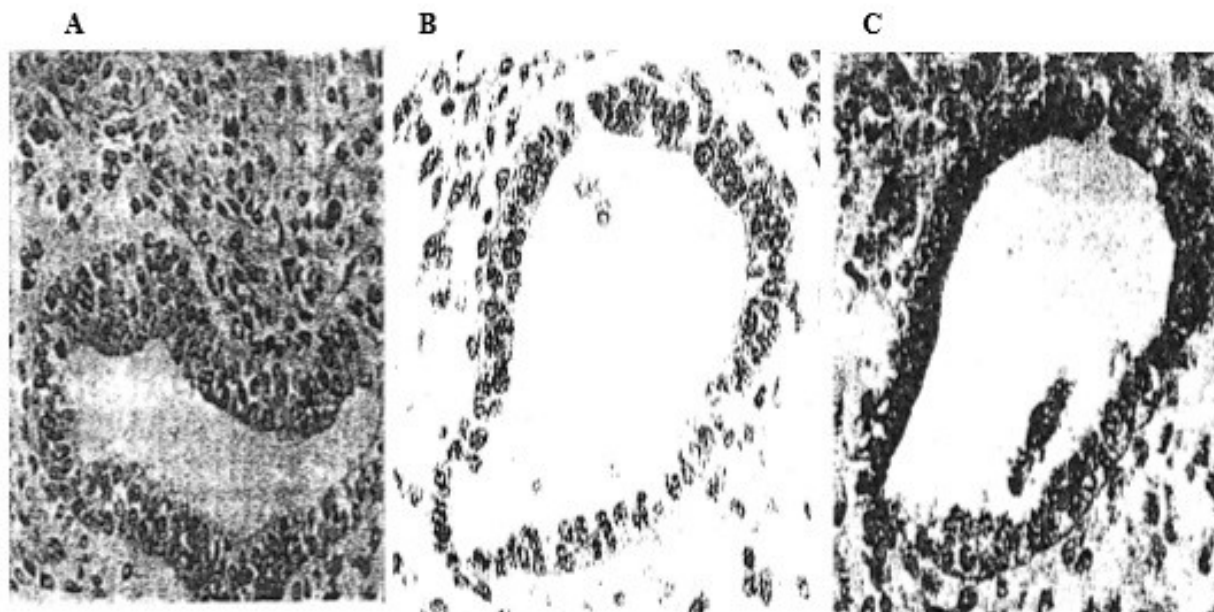
### 2.2 Glioblastoma: Pathogenesis, Diagnosis, and Prognosis

Glioblastomas are usually located in the subcortical white matter and deep grey matter, and the histopathology of GBM is characterized by cellular pleomorphism, nuclear atypia, and areas of hemorrhage and necrosis. Generally, we distinguish between giant cell glioblastomas, gliosar-

comas, and epithelioid glioblastomas [2,34]. GBM pathogenesis involves gene suppression, expression changes, and abnormal dysregulation of growth factors, such as IGF-I, TGF $\beta$ , VEGF, or EGF, and is related to synaptogenesis signal pathways [17].

GBM treatment involves complex molecular characteristics and an immunosuppressive microenvironment [35]. The tumor microenvironment involves immunosuppression, astrogliosis, and angiogenesis. While treatments, such as immune checkpoint inhibitors or chimeric antigen receptor T cell (CAR-T) therapy, have made progress, tumor vaccines based on dendritic cells (DCs) have also appeared in immunotherapy [36]. DCs activate tumor-specific T cells and initiate adaptive immune responses [37,38]. Nevertheless, the immunosuppressive microenvironment in GBM is known to inhibit the function of DCs by secreting immunosuppressive factors, such as TGF- $\beta$  and IL-10 [39]. Although clinical trials using DC vaccines have confirmed an immune response effect, the application of DC vaccines in GBM treatment needs to be optimized [40,41]. Conversely, considering that GBM is composed of glial cells expressing IGF-I, then glial neoplastic cells are included in the astrogliosis phenomenon [19,42–45]. This phenomenon, observed in CNS pathologies such as GBM, is characterized by transforming astrocytes into reactive states. These reactive astrocytes exhibit upregulated intermediate filaments (nestin, vimentin), STAT-3 element of signal transduction, the BDNF growth factor, and CD44 adhesion molecules [45]. The oncogenic transformation of these reactive astrocytes in GBM compromises the blood-brain barrier, enables the entry of immune cells from blood, and can facilitate GBM-related immunotherapies [46]. Regarding angiogenesis, GBM is rich in blood vessels and VEGF, which promotes new blood vessel formation. Anti-angiogenic agents inhibit new blood vessel formation and promote existing vessel regression. Several anti-angiogenic agents have been investigated in clinical trials and have shown promising preliminary results in newly diagnosed and recurrent GBMs [47].

Magnetic resonance imaging (MRI) and computed tomography (CT) exams are used primarily in emergency laboratory diagnoses. Meanwhile, positron emission tomography (PET) and single-photon emission computed tomography (SPECT) facilitate early detection and treatment monitoring [10]. Regarding GBM biomarkers, IGF-I, IDH, P53, olig 2, and S100 are positive, and stem cell markers CD44 and CD133 are generally positive [2,42,43,48,49]. EGFR amplification, the PI3K pathway-related genes, *PTEN* genes, and *RTK* gene alterations should also be considered [50]. For the prognosis of GBM, the giant cell subtype has a better prognosis, whereas gliosarcoma and epithelioid GBM are associated with a poor prognosis; nonetheless, the GBM treatment using temozolomide (MGMT promoter methylated tumor) promotes improved survival [1,13,51]. When considering IGF-I as a diagnostic



**Fig. 1. Histogenesis of neuroectodermal structures in the mouse teratocarcinoma model.** The tumor is derived from PCC4 stem cells. (A) A cyst of nervous origin is a pathological neural tube surrounded by neuroepithelial stem cells, and exhibits characteristics of neuroglia. The cyst shows a basal limit. Hematoxylin and eosin stain (HE) ( $\times 250$ ). (B) The more advanced stage of cyst differentiation is the neuroependymal cyst, which imitates the ependymal canal, which is surrounded by neuroepithelial cells. HE ( $\times 250$ ). (C) AFP labelling of the cyst shown in (B). The wall of the cyst and some more differentiated neuroepithelial cells (upper left) are positively stained. The limiting external of the cyst is visible. Immunoperoxidase counterstained with hematoxylin [30]. HE ( $\times 250$ ). Created using JPG.

and prognostic biomarker, this growth factor also constitutes the principal target for immuno–gene therapies [18]. Moreover, cell-free microRNAs in the blood exhibit potential in diagnosing, prognosticating, and monitoring treatment. The cell-free miRNA profile within CSF demonstrates high potential in delivering precise and specific evaluations of brain tumors [52].

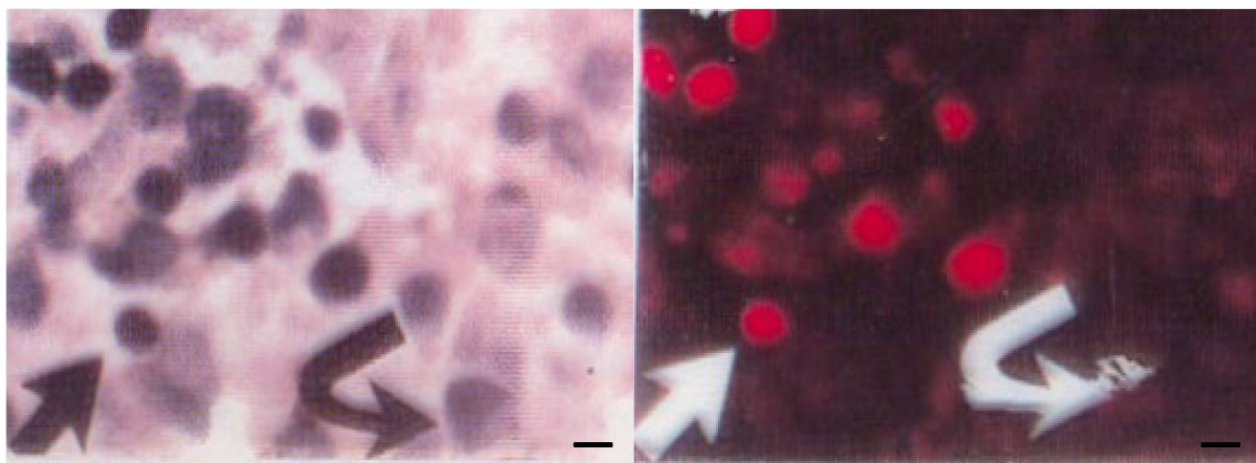
### 2.3 Therapies for Glioblastoma

Classical therapies include surgery, radiotherapy, and chemotherapy; however, the maximalist tumor resection remains the first-line treatment [1]. Nevertheless, high doses of glucocorticoids (GCs) improve neurologic deficits, and patients receiving excessive doses of GCs present high CD8 T cell expression, with the latter playing a role in the anti-tumor response [1,11,12,53]. On the other hand, nuclear medicine has introduced theranostic approaches—the therapeutic potential of targeted radionuclide therapy [54–57]. In parallel with radiotherapy, a postoperative radiation, i.e., chemotherapy, such as temozolomide (TMZ), can be administered to improve the effective survival [58,59]; however, many GBM patients are resistant to temozolomide, affecting the immune response of the host [17,59,60]. Moreover, chemotherapy is often replaced by anti-tumor vaccines such as bevacizumab (Bev) and anti-VEGF-A, al-

though genetic variations in VEGF expression influence treatment resistance [61]. Neoadjuvant Bev treatment decreases the volume of GBM and improves clinical symptoms. Meanwhile, the responsiveness of the patient to Bev might depend on the level of VEGF expression [62]. Other growth factors, such as EGFR and EGFRvIII, can also be considered for therapeutic purposes. EGFR and EGFRvIII amplification are frequently observed in GBMs and contribute to tumorigenesis and progression. However, therapies directed against EGFR and EGFRvIII have yet to present clear clinical benefits; combination therapies are needed to improve outcomes [15]. Moreover, studies have investigated the clinical intravenous delivery of a single dose of autologous T cells redirected toward the EGFRvIII mutation by a chimeric antigen receptor (CAR). These investigations may improve the efficacy of EGFRvIII-directed strategies in GBM [63].

Immunotherapeutic strategies involving immune checkpoint inhibitors, adoptive T cell therapy, and viral immunotherapy have increased interest in immune and virus studies [1,64–69]. The adoptive T cell (ACT) strategy (CAR-T cells), a more recent cancer therapy [66], dramatically improved tumor volume. However, further research is needed to resolve challenges related to the mechanisms involved [14,66,70–72]. In addition to





**Fig. 2. Rat glioma tumor.** (left) The tumor derived from C6 glioma cells one week after subcutaneous injection of transfected IGF-I antisense C6 cells. HE ( $\times 400$ ). (right) An area of tumor showing infiltration of T lymphocytes stained with anti-CD8 antibodies (immunofluorescence produced by rhodamine) [75]. HE ( $\times 400$ ). Scale bar = 25  $\mu\text{m}$ .

technologies, such as immune checkpoint inhibitors (such as anti-PD-1/PD-L1 and CTLA-4 antibodies), CAR-T therapy, and oncolytic viruses, demonstrating progress, tumor vaccines based on DCs have become an important part of the immunotherapy field; nevertheless, individual differences in efficacy suggest that applying DC vaccines to treat GBM requires additional optimization [39–41].

Cancer immunotherapy and nanotechnology have become increasingly intertwined in recent years. Examples of nanotechnology, NPs, used in cancer immunotherapies can be mentioned as follows [21,73]: NPs can be used to deliver immunotherapeutic agents such as checkpoint inhibitors or CAR-T cells to the tumor; NPs can be loaded with tumor antigens that activate dendritic cells; NPs administered in immuno-gene therapy can promote the delivery of agents to APCs, or participate in immunomodulation by delivering cytokines.

#### 2.4 Cancer Gene Therapy and Nanotechnology

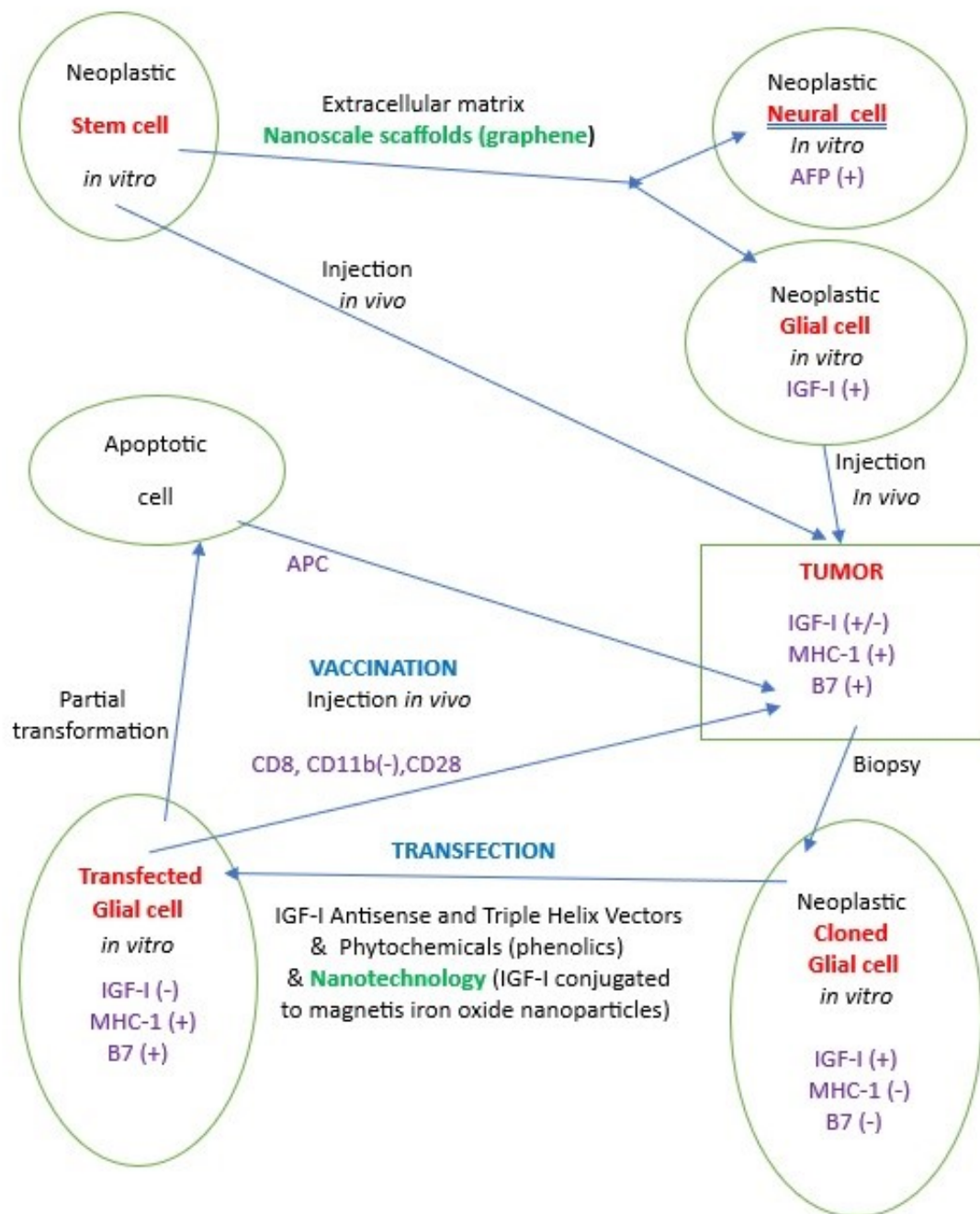
New glioblastoma therapies are mainly based on immune or immuno-gene strategies: cancer immunotherapy was established by Townsend and Allison [74] and cancer gene therapy by Trojan *et al.* [75], followed by others [64–66,70–72,76–78]. Cancer gene therapy targeting growth factor genes, especially *IGF-I*, the principal neoplastic development factor, was performed in tumor cells using antisense [79,80] or triple helix technologies [81,82] (Fig. 2, Ref. [75]).

Other promising approaches involve targeting growth factors such as TGF-beta or VEGF and EGF, their receptors, and their downstream transduction signaling elements [6,16,83,84]. Recent neuro-oncology research has highlighted the role of the PI3K/AKT pathway in glial cells: the simultaneous arrest of at least two links, either

IGF-I, TGF $\beta$ , or VEGF, and GS, and of their pathways, TK/PI3K/AKT/GSK3/GS/BCL-2/mTOR [85–89] could be useful for future clinical gene therapies for GBM.

An anti-gene IGF-I strategy that blocks IGF-I synthesis at the transcription or translation levels [90] has been introduced into clinical trials, with the median survival of patients reaching 18 to 20 months [91]. Immune and gene therapies, particularly anti-gene *IGF-I*/phytochemical immuno-gene treatment, increased the average survival span of GBM patients up to 20–24 months.

These promising clinical results were obtained using cell vaccines (anti-IGF-I antisense and triple helix transfection) presenting strong immunogenicity: CD8 and CD28 molecules. Immunogenicity was induced by a common signal transduction pathway produced by anti-gene and phytochemical technologies (TK/PI3K/AKT and TLR/MAPK/NF $\kappa$ B and JAK/STAT) [17]. Anti-IGF-I therapy, including nanotechnology, is currently being studied to reinforce the signal transduction pathway of IGF-I-R (TK/PI3K/AKT) engaged in the anti-gene *IGF-I*/phytochemical therapy. An increase in signal transduction is decisive to ensuring important cell immunogenicity—the principal mechanism of effective anti-tumor immune response (MHC-I and B7 inducing *in vivo* CD8 response). Nanoparticles—theranostic iron oxide—conjugated to IGF-I and targeting IGF-I receptor (and related signal transduction pathway) have produced an anti-tumor immune effect leading to cancer cell apoptosis [17,92] (Fig. 3, Ref. [17–20,23,27,30,32,75,79,82,90,92–100]).



**Fig. 3. Schematic of the experimental cancer gene therapeutic process (anti-IGF-I/phytochemical/nanotechnology).** The stem cells, i.e., PCC4 cells [30], in the presence of nanoscale scaffolds (graphene), transition to neural and glial cells [23,93]. The stem cells, if injected in a mouse model *in vivo*, induce a teratocarcinoma tumor reproducing CNS neoplastic development containing neural cells (alpha-fetoprotein positive) and glial cells (IGF-I positive) [27,32]. On the other side, the glial cells (i.e., C6), if injected *in vivo*, induce another type of tumor: glioma murine tumor [19,75]. The neoplastic glial cells removed by biopsy from the glioma tumor are transfected *in vitro* using IGF-I antisense or IGF-I triple helix vectors [17,79,82,90,94], combined with phytochemicals derived from essential oils of Acmella and Geraniaceae [18,95,96] and nanotechnology (conjugation of human IGF-I to magnetic iron oxide nanoparticles (IONPs)) [92]. All three technologies combine to increase cell immunogenicity (expression of MHC-1 and B7) due to a common signaling pathway: anti-IGF-I tyrosine kinase—PI3K/AKT/PKC/GSK3 [17,20,97]; phytochemicals (TLR/MAPK/NF $\kappa$ B and JAK/STAT elements [98–100]; nanotechnology (IGF-I-R and PI3K/AKT) [92]. These transfected cells and apoptotic cells originated from these transfected cells, after irradiation, are injected (vaccine) *in vivo*, stimulating CD8 and CD28 mediated T lymphocytes and APC cells, producing an immune anti-tumoral mechanism [17]. Created using JPG.

### 3. Current Brain Tumorigenesis and Nanotechnology

#### 3.1 Stem Cells

New neurons are created throughout life through a process known as neurogenesis [101,102]. Neurogenesis occurs when neural stem cells (NSCs) generate new multipotent cells. This process begins during fetal development and persists in adulthood [103,104]. The first description of the central nervous system development was performed between 1979 and 1981 by applying an alpha-fetoprotein marker using a rat brain fetal model and then a neoplastic model of CNS in mouse teratocarcinoma derived from PCC4 stem cells. A comparison of the two models demonstrated the existence of convergence between embryonic/fetal development and neoplastic development [29,30].

Immunologic factors play an important role in normal and neoplastic development, especially when related to insulin-like growth factor-1 (IGF-I), which plays a role in normal NSCs by inducing differentiation, proliferation, or survival of neurons [105–107]. As the main neoplastic development factor, IGF-I was proposed as a target to treat glioblastoma progression [44,108].

Stem cell research offers great promise for regenerative medicine, tissue engineering, drug screening, and clinical therapies. However, the success of such applications relies on the ability to accurately control the differentiation and growth of stem cells into functional tissues. Meanwhile, the distinction of stem cells can be directed by controlling their microenvironment, including substrate stiffness, topography, and chemical cues. However, methods for controlling traditional microenvironments lack the precision required for successful and reproducible differentiation. Nanotechnology provides a way to precisely control the behavior of stem cells differentiating into neuroglial cells by manipulating their cellular microenvironment at the nanoscale [32].

The unique properties of nanomaterials, including their small size, high surface area, and tunable physicochemical properties, have enabled researchers to manipulate and control cellular behavior with a high degree of precision. In particular, nanomaterials have been developed to mimic the extracellular matrix (ECM) and provide physical and chemical cues to guide stem cell behavior. Additionally, nanoparticles can act as delivery vehicles for therapeutics or genetic material, enabling targeted gene therapy. Furthermore, one of the major applications of nanotechnology in stem cell research is the development of nanoscale scaffolds that mimic the ECM found in the tissues. Nanotechnology has been used to create scaffolds that support the growth and differentiation of neurons and glial cells, with potential applications in treating neurodegenerative diseases and spinal cord injuries. These scaffolds can be designed to provide physical support and chemical and mechanical cues to guide stem cell differentiation and tissue re-

generation. For example, nanofibrous scaffolds have been shown to support the distinction of mesenchymal stem cells into bone, cartilage, and other tissue types [23,24,93,109–113].

#### 3.2 Neuron and Glial Cells

Nanotechnology has also been applied to studying neural cells, including neurons and glial cells. Similar to stem cells, neurons and glial cells require specific microenvironments for growth and differentiation. Thus, nanotechnology has been used to create scaffolds that support the development and differentiation of these cells. Nanofibers can be used as scaffolds to promote neurite outgrowth and synapse formation. Additionally, nanoparticles can be engineered to deliver therapeutic molecules or drugs directly to neurons or glial cells. In neuroscience, nanoelectronics devices are being developed to interface with neural cells and circuits. These devices can record and stimulate neural activity with high spatial and temporal resolution, enabling researchers to improve understanding of brain function. For example, researchers have developed nanowire arrays that can be implanted into the brain to record neural activity. These devices have been used to study various phenomena, including learning and memory, perception, and motor control. Nanoparticles have also been used as contrast agents for imaging techniques such as MRI and PET, allowing researchers to visualize and track neural cells *in vivo* [25,114].

Neurons and glial cells require specific microenvironments for growth and differentiation, and nanotechnology can be used to create scaffolds that support their growth and differentiation. In addition to scaffolds, nanoparticles have also been used to deliver therapeutic agents directly to stem cells or the tissues that require regeneration. These nanoparticles can be functionalized with targeting ligands or imaging agents to improve specificity and efficacy. Moreover, these nanoparticles can be engineered to release drugs or growth factors in a controlled manner, enabling precise temporal and spatial control over the biological response [23,25,115].

Nanomaterials have been developed to mimic the ECM and provide physical and chemical cues to guide stem cell behavior, followed by their ulterior neuroglial differentiation. For example, nanofibers can replicate the collagen fiber structure in the ECM, providing a scaffold for stem cells to attach and grow on. Similarly, nanoparticles can be functionalized with specific ligands that bind to cell surface receptors, directing stem cell differentiation. One example of a nanomaterial used in stem cell research is graphene, a two-dimensional material of carbon atoms arranged in a hexagonal lattice. The unique properties of graphene, including its high surface area, mechanical strength, and electrical conductivity, make it an attractive material for creating scaffolds for tissue engineering applications. Researchers have demonstrated that graphene-based scaffolds can support the growth and differentiation of various cells,

including neural stem cells. In one study, graphene oxide nanosheets were incorporated into a hydrogel scaffold to promote the regeneration of spinal cord tissue in rats following injury. These researchers found that the graphene oxide nanosheets increased the adhesion and proliferation of neural progenitor cells, improving functional recovery in the injured animals [23,32,93,109].

Nanomaterials can also be used as therapeutics or genetic material delivery vehicles, enabling targeted gene therapy and drug delivery. Nanomaterials can be engineered to release drugs or other therapeutic agents in response to specific stimuli, such as changes in pH or temperature. This allows precise control over drug release timing and location, improving therapeutic efficacy and reducing side effects. In one study, researchers developed gold nanoparticles functionalized with a protein that targets cancer cells. When these nanoparticles were loaded with a chemotherapy drug and injected into mice with brain tumors, the nanoparticle–drug combination selectively targeted and killed cancer cells while sparing normal cells. This approach can potentially improve the effectiveness of chemotherapy while minimizing toxicity to healthy tissues [25,110,114,115].

Nanotechnology offers unique opportunities to control stem and neuronal/glial cell behavior by manipulating their microenvironment. Nanomaterials can mimic the ECM and provide physical and chemical cues to guide stem cell behavior, while nanoparticles can act as delivery vehicles for therapeutics or genetic material. Furthermore, nanotechnology has been used to create scaffolds that support the growth and differentiation of neurons and glial cells. Overall, nanotechnology in stem cell and neuronal/glial cell research holds great promise for regenerative medicine, tissue engineering, and drug screening. Meanwhile, integrating nanotechnology with stem cell and neuronal/glial cell research could revolutionize regenerative medicine and neuroscience. Therefore, with continued innovation and development, researchers may be able to create more effective therapies for a wide range of diseases and injuries [17,23,114,115].

### 3.3 Brain Neoplastic Development

Brain development is marked by a complex interplay of molecular, cellular, and structural adaptations, which promote the sophisticated functions performed by the brain. Neoplastic development, such as gliomas and glioblastomas, often exploits pathways similar to those involved in brain evolution, such as cellular plasticity, angiogenesis, and metabolic adaptability. Hence, nanotechnology provides a unique vantage point for studying both the development processes of the brain and the mechanisms underlying tumorigenesis, enabling advancements in diagnostics, research, and therapeutic applications [23,24,109,116].

The human brain is a product of millions of years of evolution, marked by increased cortical size, neural com-

plexity, and functional specialization. Parallely, neoplastic brain diseases, such as glioblastoma, highlight the vulnerabilities of this organ, exploiting developmental and evolutionary mechanisms for malignant growth. Thus, applying nanotechnology offers a dual advantage: Studying the evolutionary intricacies of the brain while addressing its pathological transformations. This chapter delves into the convergence of these fields and the role of nanotechnology in advancing research and treatment paradigms. The evolution of the brain from simpler structures in early vertebrates to the complex human brain involves key genetic, structural, and metabolic changes. Nanotechnology offers tools to dissect these processes with unprecedented precision as follows [116].

**Gene expression studies:** Nanoparticles conjugated with fluorescent probes have enabled gene expression patterns to be traced in ancestral and modern brains. For example, nanoscale sensors track the activity of evolutionarily significant genes across species, such as *FOXP2* (language development) and *SRGAP2* (neural connectivity) [116,117].

**Epigenetics:** gold nanoparticles map histone modifications and DNA methylation, revealing epigenetic changes that underlie brain evolution [110,118].

**Neural circuitry mapping:** (a) Nanoelectrodes. High-resolution nanoelectrodes have mapped evolutionary changes in synaptic structures, highlighting how interspecies variations in connectivity influence cognition and behavior [119]. (b) Brain organoids. Nano-scaffolds facilitate the growth of organoids that mimic ancestral and modern brains, enabling comparative studies on structural and functional evolution [93,120].

Brain tumors, especially gliomas and glioblastomas, share several hallmarks with neoplastic development processes, including (a) cellular plasticity: tumor cells exhibit stem-like behavior, a feature central to neural development during evolution; (b) angiogenesis: tumors coordinate blood vessel formation and developmental pathways; (c) metabolic adaptability: similar to the brain evolutionary processes, tumors have evolved to adapt to limited oxygen and nutrient supplies [18,116].

To understand tumor progression using nanotechnology we can consider (a) the tumor microenvironment: Nanoparticles equipped with biosensors detect pH, oxygen, and nutrient levels, providing real-time data on the tumor microenvironment [121]; (b) genomic mutations: nanoscale sequencing tools can identify key mutations driving tumorigenesis, such as those in the *TP53* and *IDH1/2* genes [122]; (c) three-dimensional tumor models: nano-engineered scaffolds replicate brain tumor growth, enabling studies on tumor invasion and drug resistance mechanisms [93,123]; (d) single-cell analysis: nanoparticle-based techniques isolate and analyze individual tumor cells, uncovering heterogeneity and identifying therapy-resistant populations [124,125].



### 3.4 Brain Tumor Therapies

The insights gained from studying brain tumoral development in relation with tumoral therapies translate into innovative nanotechnology-driven therapies: (a) Drug delivery systems: Nanocarriers bypass the blood–brain barrier to deliver targeted therapies, such as chemotherapeutics or RNA-based treatments, to tumor sites [111,125–127]; (b) gene editing: CRISPR–Cas9 systems, delivered via nanoparticles, target oncogenes while offering insights into gene functions critical for brain evolution [128]; (c) Immunotherapy: Nanoparticles, present tumor antigens to the immune system, enhancing responses against glioblastoma. Immunotherapy performed using nanotechnology constitutes the presently employed promising approach in cancer treatment [21,73,112,125,129–133]; NP-related therapy was recently approved for GBM treatment—NanoTherm™ [134]—and is based on iron oxide NPs and the thermal ablation of the tumor using a magnetic field. In parallel, other types of NPs are being evaluated, including polymer and lipid-based nanoformulations, nanodiscs, dendrimers, metallic, and silica. The advantages of these nanoscale drug carriers include improved penetration across the blood–brain barrier and lower systemic toxicity.

Nanotechnology in relation to signal transduction pathways plays a role in GBM immune and gene treatment mechanisms: (1) Nanostructured scaffolds can mimic the ECM, influencing signaling pathways [109,113,135]; (2) gold NPs, AuNPs, and those conjugated with EGF can activate the EGF receptor [110]; (3) nanocarriers for targeted cancer therapy—the liposomes coated with antibodies against the HER2 receptor—can deliver doxorubicin specifically to HER2-positive cancer cells, and modulate the PI3K/AKT pathway; (4) NPs can deliver small interfering RNAs (siRNAs), either small molecule inhibitors directly to cancer, or reinforce the IGF-I-R signal engaged in anti-IGF-I/phytochemical strategy [109,135]. Ongoing cancer immune and gene therapeutic studies are analyzing nanotechnology mechanisms related to signal transduction pathways and immune response [131,132,136–138].

Future research should focus on: (a) integrating brain organoid studies with evolutionary genomics and tumor modelling; (b) enhancing biocompatibility and minimizing off-target effects; (c) refining nanocarriers for personalized and adaptive therapies, particularly immunotherapies [21,112,125,131,139].

## 4. Conclusion

This review describes long-term research surrounding GBM treatment and analyzes brain neoplastic development from stem cells differentiating into neuroglial cells, followed by the establishment of cancer gene therapy. This cancer gene therapy is based on immunogenic vaccines originating from brain neoplastic glial cells. These vaccines were prepared for the first time according to a strategy using three technologies in parallel: anti-gene, phyto-

chemical, and nanotechnologies, all of which induce immunogenicity. The anti-tumor vaccine immunogenicity (CD8, CD28) is mediated by the overexpression of the common IGF-R signal transduction pathway, which was reinforced by nanotechnology. In the future, this approach will be applied by using a brain neoplastic stem cell therapy in relation to nanotechnology. The ability of nanotechnology to analyze, model, and intervene at the molecular level offers transformative potential for understanding the neoplasia of the brain and combating fatal malignant brain tumors—glioblastomas. Cancer immunotherapy, gene therapy, and nanotechnology have recently become increasingly intertwined, and the inclusion of nanotechnology in GBM immunotherapy is undergoing permanent progress [94,132,140–142].

## Author Contributions

GQ: Conceptualization; Formal analysis; Investigation; Methodology; Roles/Writing — original draft; and Writing — review & editing. MR: Formal analysis; Methodology; Validation; Roles/Writing — original draft. YCL: Formal analysis; Methodology; Validation. JT: Conceptualization; Formal analysis; Investigation; Methodology; Project administration; Supervision; Validation; Roles/Writing — original draft; and Writing — review & editing. All authors have read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

## Ethics Approval and Consent to Participate

Not applicable.

## Acknowledgment

We thank Dr W. Timmer (NIH - NCI, Bethesda, USA) - Program Director, Adult Brain Tumor Consortium), recently deceased, for his help in recognition of cancer gene therapy as a new oncology treatment proposed by J. Trojan thirty years ago. We need to underline the engagement in the described basic CNS research by Drs J. Uriel (CNRS, Villejuif) and in the clinical cancer gene therapy by Dr D. Anthony (CWRU University, USA), H. Kasprzak (Nicolaus Copernic University, Poland), and Pedro J. Penagos (National Cancer Institute, Colombia). We are grateful for a research collaboration in stem cells and neoplastic development to Dr S.J. Bueno (UNAB, Floridablanca, Colombia). We thank Dr A. Trojan (university License in English Literature) for verification of English spelling and grammar. We need to underline the logistic support of W. Arenas (GTF SA, USA) in the publication of previous experimental results.

## Funding

This research received no external funding.



## Conflict of Interest

All authors declare no conflicts of interest. Despite receiving sponsorship from CEDEA & ICGT (Center of Oncology Diagnostic and International Cancer Gene Therapy Foundation), the judgments in data interpretation and writing were not influenced by this relationship.

## References

- [1] Batash R, Asna N, Schaffer P, Francis N, Schaffer M. Glioblastoma Multiforme, Diagnosis and Treatment; Recent Literature Review. *Current Medicinal Chemistry*. 2017; 24: 3002–3009. <https://doi.org/10.2174/0929867324666170516123206>.
- [2] Louis DN, Perry A, Wesseling P, Brat DJ, Cree IA, Figarella-Branger D, *et al.* The 2021 WHO Classification of Tumors of the Central Nervous System: a summary. *Neuro-oncology*. 2021; 23: 1231–1251. <https://doi.org/10.1093/neuonc/noab106>.
- [3] Miller KD, Ostrom QT, Kruchko C, Patil N, Tihan T, Cioffi G, *et al.* Brain and other central nervous system tumor statistics, 2021. *CA: a Cancer Journal for Clinicians*. 2021; 71: 381–406. <https://doi.org/10.3322/caac.21693>.
- [4] Jamshidi P, Brat DJ. The 2021 WHO classification of central nervous system tumors: what neurologists need to know. *Current Opinion in Neurology*. 2022; 35: 764–771. <https://doi.org/10.1097/WCO.0000000000001109>.
- [5] Alifieris C, Trafalis DT. Glioblastoma multiforme: Pathogenesis and treatment. *Pharmacology & Therapeutics*. 2015; 152: 63–82. <https://doi.org/10.1016/j.pharmthera.2015.05.005>.
- [6] Haque S, Morris JC. Transforming growth factor- $\beta$ : A therapeutic target for cancer. *Human Vaccines & Immunotherapeutics*. 2017; 13: 1741–1750. <https://doi.org/10.1080/21645515.2017.1327107>.
- [7] Chetty R. Gene of the month: GLI-1. *Journal of Clinical Pathology*. 2020; 73: 228–230. <https://doi.org/10.1136/jclinpat-2020-206431>.
- [8] Takei N, Yokomaku D, Yamada T, Nagano T, Kakita A, Namba H, *et al.* EGF Downregulates Presynaptic Maturation and Suppresses Synapse Formation In Vitro and In Vivo. *Neurochemical Research*. 2022; 47: 2632–2644. <https://doi.org/10.1007/s11064-021-03524-6>.
- [9] Zegarra-Valdivia J, Nuñez A, Aleman IT. Untangling IGF-I signaling in the aging brain. *Aging*. 2023; 15: 599–600. <https://doi.org/10.18632/aging.204507>.
- [10] Ellingson BM, Chung C, Pope WB, Boxerman JL, Kaufmann TJ. Pseudoprogression, radionecrosis, inflammation or true tumor progression? challenges associated with glioblastoma response assessment in an evolving therapeutic landscape. *Journal of Neuro-oncology*. 2017; 134: 495–504. <https://doi.org/10.1007/s11060-017-2375-2>.
- [11] Noch EK, Ramakrishna R, Magge R. Challenges in the Treatment of Glioblastoma: Multisystem Mechanisms of Therapeutic Resistance. *World Neurosurgery*. 2018; 116: 505–517. <https://doi.org/10.1016/j.wneu.2018.04.022>.
- [12] Wong ET, Swanson KD. Dexamethasone-Friend or Foe for Patients With Glioblastoma? *JAMA Neurology*. 2019; 76: 247–248. <https://doi.org/10.1001/jamaneurol.2018.4530>.
- [13] Wick A, Kessler T, Platten M, Meisner C, Bamberg M, Herlinger U, *et al.* Superiority of temozolomide over radiotherapy for elderly patients with RTK II methylation class, MGMT promoter methylated malignant astrocytoma. *Neuro-oncology*. 2020; 22: 1162–1172. <https://doi.org/10.1093/neuonc/noaa033>.
- [14] Xiong Z, Raphael I, Olin M, Okada H, Li X, Kohanbash G. Glioblastoma vaccines: past, present, and opportunities. *EBioMedicine*. 2024; 100: 104963. <https://doi.org/10.1016/j.ebiom.2023.104963>.
- [15] An Z, Aksoy O, Zheng T, Fan QW, Weiss WA. Epidermal growth factor receptor and EGFRvIII in glioblastoma: signaling pathways and targeted therapies. *Oncogene*. 2018; 37: 1561–1575. <https://doi.org/10.1038/s41388-017-0045-7>.
- [16] Sampson JH, Gunn MD, Fecci PE, Ashley DM. Brain immunology and immunotherapy in brain tumors. *Nature Reviews. Cancer*. 2020; 20: 12–25. <https://doi.org/10.1038/s41568-019-0224-7>.
- [17] Trojan A, Lone YC, Briceno I, Trojan J. Anti-Gene IGF-I Vaccines in Cancer Gene Therapy: A Review of a Case of Glioblastoma. *Current Medicinal Chemistry*. 2024; 31: 1983–2002. <https://doi.org/10.2174/0109298673237968231106095141>.
- [18] Trojan J. Establishment of cancer gene therapy. Cambridge Scholars Publishing: UK. 2023.
- [19] Irais CM, Maria-de-la-Luz SG, Dealmy DG, Agustina RM, Nidia CH, Mario-Alberto RG, *et al.* Plant Phenolics as Pathogen-Carrier Immunogenicity Modulator Haptens. *Current Pharmaceutical Biotechnology*. 2020; 21: 897–905. <https://doi.org/10.2174/1389201021666200121130313>.
- [20] Hakuno F, Takahashi SI. IGF1 receptor signaling pathways. *Journal of Molecular Endocrinology*. 2018; 61: T69–T86. <https://doi.org/10.1530/JME-17-0311>.
- [21] García-Domínguez DJ, López-Enríquez S, Alba G, Garnacho C, Jiménez-Cortegana C, Flores-Campos R, *et al.* Cancer Nano-Immunotherapy: The Novel and Promising Weapon to Fight Cancer. *International Journal of Molecular Sciences*. 2024; 25: 1195. <https://doi.org/10.3390/ijms25021195>.
- [22] Bueno SJ, Trojan A, Santander R, Alvarez A, Guzman A, Rojas C, *et al.* Brain stem cells and IGF-I: implications in development, regeneration and cancer therapeutics. *Integrative Molecular Medicine*. 2018; 5. <https://doi.org/10.15761/MM.1000319>.
- [23] Khan FA, Almohazey D, Alomari M, Almoftly SA. Impact of nanoparticles on neuron biology: current research trends. *International Journal of Nanomedicine*. 2018; 13: 2767–2776. <https://doi.org/10.2147/IJN.S165675>.
- [24] Liu Z, Ji X, He D, Zhang R, Liu Q, Xin T. Nanoscale Drug Delivery Systems in Glioblastoma. *Nanoscale Research Letters*. 2022; 17: 27. <https://doi.org/10.1186/s11671-022-03668-6>.
- [25] Alhodieb FS, Rahman MA, Barkat MA, Alanezi AA, Barkat HA, Hadi HA, *et al.* Nanomedicine-driven therapeutic interventions of autophagy and stem cells in the management of Alzheimer's disease. *Nanomedicine*. 2023; 18: 145–168. <https://doi.org/10.2217/nnm-2022-0108>.
- [26] Love S, Perry A, Ironside J, Budka H. *Greenfield's Neuropathology*. 9th edn. CRC Press/Wolters Kluwer: New York. 2015.
- [27] Trojan J, Naval X, Johnson T, Lafarge-Frayssinet C, Hajeri-Germond M, Farges O, *et al.* Expression of serum albumin and of alphafetoprotein in murine normal and neoplastic primitive embryonic structures. *Molecular Reproduction and Development*. 1995; 42: 369–378. <https://doi.org/10.1002/mrd.1080420402>.
- [28] Trojan J. Brain - from development to neoplasia and gene therapy solution. 1st edn. Lambert Academic Publishers: Saarbrucker, Germany. 2017/2018.
- [29] Trojan J, Uriel J. Intracellular localization of alpha-fetoprotein and serum albumin in the central nervous system of the rat during fetal and postnatal development. *Comptes Rendus des Seances De L'Academie des Sciences. Serie D, Sciences Naturelles*. 1979; 289: 1157–1160. (In French)
- [30] Trojan J, Uriel J. Localisation of alphafetoprotein (AFP) in murine teratocarcinoma. *Biomedicine*. 1981; 34: 140–146.
- [31] Vaz A, Ribeiro I, Pinto L. *Frontiers in Neurogenesis*. *Cells*. 2022; 11: 3567. <https://doi.org/10.3390/cells11223567>.
- [32] Trojan J, Johnson TR, Rudin SD, Blossey BK, Kelley KM, Shevelev A, *et al.* Gene therapy of murine teratocarcinoma: separate functions for insulin-like growth factors I and II in immunogenicity and differentiation. *Proceedings of the National*

- Academy of Sciences of the United States of America. 1994; 91: 6088–6092. <https://doi.org/10.1073/pnas.91.13.6088>.
- [33] Biserova K, Jakovlevs A, Uljanovs R, Strumfa I. Cancer Stem Cells: Significance in Origin, Pathogenesis and Treatment of Glioblastoma. *Cells*. 2021; 10: 621. <https://doi.org/10.3390/cell10030621>.
- [34] Ostrom QT, Cioffi G, Gittleman H, Patil N, Waite K, Kruchko C, *et al.* CBTRUS Statistical Report: Primary Brain and Other Central Nervous System Tumors Diagnosed in the United States in 2012–2016. *Neuro-oncology*. 2019; 21: v1–v100. <https://doi.org/10.1093/neuonc/noz150>.
- [35] Lin H, Liu C, Hu A, Zhang D, Yang H, Mao Y. Understanding the immunosuppressive microenvironment of glioma: mechanistic insights and clinical perspectives. *Journal of Hematology & Oncology*. 2024; 17: 31. <https://doi.org/10.1186/s13045-024-01544-7>.
- [36] Liu B, Zhou H, Tan L, Siu KTH, Guan XY. Exploring treatment options in cancer: Tumor treatment strategies. *Signal Transduction and Targeted Therapy*. 2024; 9: 175. <https://doi.org/10.1038/s41392-024-01856-7>.
- [37] Hilligan KL, Ronchese F. Antigen presentation by dendritic cells and their instruction of CD4+ T helper cell responses. *Cellular & Molecular Immunology*. 2020; 17: 587–599. <https://doi.org/10.1038/s41423-020-0465-0>.
- [38] Perez CR, De Palma M. Engineering dendritic cell vaccines to improve cancer immunotherapy. *Nature Communications*. 2019; 10: 5408. <https://doi.org/10.1038/s41467-019-13368-y>.
- [39] Liu Y, Zhou F, Ali H, Lathia JD, Chen P. Immunotherapy for glioblastoma: current state, challenges, and future perspectives. *Cellular & Molecular Immunology*. 2024; 21: 1354–1375. <https://doi.org/10.1038/s41423-024-01226-x>.
- [40] Yu J, Sun H, Cao W, Song Y, Jiang Z. Research progress on dendritic cell vaccines in cancer immunotherapy. *Experimental Hematology & Oncology*. 2022; 11: 3. <https://doi.org/10.1186/s40164-022-00257-2>.
- [41] Pasqualetti F, Zanotti S. Nonrandomised controlled trial in recurrent glioblastoma patients: the promise of autologous tumor lysate-loaded dendritic cell vaccination. *British Journal of Cancer*. 2023; 129: 895–896. <https://doi.org/10.1038/s41416-023-02194-1>.
- [42] Kiess W, Lee L, Graham DE, Greenstein L, Tseng LY, Rechler MM, *et al.* Rat C6 glial cells synthesize insulin-like growth factor I (IGF-I) and express IGF-I receptors and IGF-II/mannose 6-phosphate receptors. *Endocrinology*. 1989; 124: 1727–1736. <https://doi.org/10.1210/endo-124-4-1727>.
- [43] Pollak MN, Schernhammer ES, Hankinson SE. Insulin-like growth factors and neoplasia. *Nature Reviews. Cancer*. 2004; 4: 505–518. <https://doi.org/10.1038/nrc1387>.
- [44] Zumkeller W. IGFs and IGF-binding proteins as diagnostic markers and biological modulators in brain tumors. *Expert Review of Molecular Diagnostics*. 2002; 2: 473–477. <https://doi.org/10.1586/14737159.2.5.473>.
- [45] Dubois LG, Campanati L, Righy C, D'Andrea-Meira I, Spohr TCDSE, Porto-Carreiro I, *et al.* Gliomas and the vascular fragility of the blood brain barrier. *Frontiers in Cellular Neuroscience*. 2014; 8: 418. <https://doi.org/10.3389/fncel.2014.00418>.
- [46] Zhang H, Zhou Y, Cui B, Liu Z, Shen H. Novel insights into astrocyte-mediated signaling of proliferation, invasion and tumor immune microenvironment in glioblastoma. *Biomedicine & Pharmacotherapy = Biomedicine & Pharmacotherapie*. 2020; 126: 110086. <https://doi.org/10.1016/j.biopha.2020.110086>.
- [47] Ameratunga M, Pavlakakis N, Wheeler H, Grant R, Simes J, Khasraw M. Anti-angiogenic therapy for high-grade glioma. *The Cochrane Database of Systematic Reviews*. 2018; 11: CD008218. <https://doi.org/10.1002/14651858.CD008218.pub4>.
- [48] Sarkar C, Santosh V, Chako G, Mahadevan A. *Essentials of diagnostic surgical neuropathology*. 2nd edn. Thieme (Neurological Society of India): India. 2023.
- [49] Webb LM, Neth BJ, Raghunathan A, Greipp PT, Ida CM, Carabenciov ID, *et al.* A Case of Long-Term Survival After Glioblastoma, IDH-Wild Type. *The Neurologist*. 2024; 29: 254–258. <https://doi.org/10.1097/NRL.0000000000000564>.
- [50] The Cancer Genome Atlas Research Network. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature*. 2008; 455: 1061–1068. <https://doi.org/10.1038/nature07385>.
- [51] Stupp R, Hegi ME, Mason WP, van den Bent MJ, Taphoorn MJB, Janzer RC, *et al.* Effects of radiotherapy with concomitant and adjuvant temozolomide versus radiotherapy alone on survival in glioblastoma in a randomised phase III study: 5-year analysis of the EORTC-NCIC trial. *The Lancet. Oncology*. 2009; 10: 459–466. [https://doi.org/10.1016/S1470-2045\(09\)70025-7](https://doi.org/10.1016/S1470-2045(09)70025-7).
- [52] Beylerli O, Encarnacion Ramirez MDJ, Shumadalova A, Ilyasova T, Zemlyanskiy M, Beilerli A, *et al.* Cell-Free miRNAs as Non-Invasive Biomarkers in Brain Tumors. *Diagnostics*. 2023; 13: 2888. <https://doi.org/10.3390/diagnostics13182888>.
- [53] Kurdi M, Fadul MM, Addas BMJ, Faizo E, Alkhayyat S, Bamağa AK, *et al.* Dynamic interplay between corticosteroid treatment and the role of SRC-1 gene dysregulation in the progression of WHO-Grade 4 Astrocytoma. *Journal of Neurooncology*. 2020; 163: 693–705. <https://doi.org/10.1007/s11060-023-04385-5>.
- [54] Hautiere M, Vivier D, Dorval P, Pineau D, Kereselidze D, Denis C, *et al.* Preoperative PET imaging and fluorescence-guided surgery of human glioblastoma using dual-labeled antibody targeting ETA receptors in a preclinical mouse model: A therapeutic approach. *Theranostics*. 2024; 14: 6268–6280. <https://doi.org/10.7150/thno.98163>.
- [55] Klapproth AP, Shevtsov M, Stangl S, Li WB, Multhoff G. A New Pharmacokinetic Model Describing the Biodistribution of Intravenously and Intratumorally Administered Superparamagnetic Iron Oxide Nanoparticles (SPIONs) in a GL261 Xenograft Glioblastoma Model. *International Journal of Nanomedicine*. 2020; 15: 4677–4689. <https://doi.org/10.2147/IJN.S254745>.
- [56] Nobashi TW, Mayer AT, Xiao Z, Chan CT, Chaney AM, James ML, *et al.* Whole-body PET Imaging of T-cell Response to Glioblastoma. *Clinical Cancer Research: an Official Journal of the American Association for Cancer Research*. 2021; 27: 6445–6456. <https://doi.org/10.1158/1078-0432.CCR-21-1412>.
- [57] Li D, Zhang J, Chi C, Xiao X, Wang J, Lang L, *et al.* First-in-human study of PET and optical dual-modality image-guided surgery in glioblastoma using 68Ga-IRDye800CW-BBN. *Theranostics*. 2018; 8: 2508–2520. <https://doi.org/10.7150/thno.25599>.
- [58] Barani JJ, Larson DA. Radiation therapy of glioblastoma. In Raizer J, Parsa A (eds.) *Current understanding and treatment of gliomas* (pp. 49–73). Springer International Publishing: Switzerland. 2015.
- [59] Hotchkiss KM, Sampson JH. Temozolomide treatment outcomes and immunotherapy efficacy in brain tumor. *Journal of Neurooncology*. 2021; 151: 55–62. <https://doi.org/10.1007/s11060-020-03598-2>.
- [60] Karachi A, Dastmalchi F, Mitchell DA, Rahman M. Temozolomide for immunomodulation in the treatment of glioblastoma. *Neuro-oncology*. 2018; 20: 1566–1572. <https://doi.org/10.1093/neuonc/noy072>.
- [61] Kim MM, Umemura Y, Leung D. Bevacizumab and Glioblastoma: Past, Present, and Future Directions. *Cancer Journal (Sudbury, Mass.)*. 2018; 24: 180–186. <https://doi.org/10.1097/PPO.0000000000000326>.

- [62] Teshigawara A, Kyoichi T, Hasegawa Y, Murayama Y, Tanaka T. Comparative Volumetric Analyses Following Bevacizumab Therapy for a Patient With Concomitant Glioblastoma, Meningioma, and Dural Arteriovenous Fistula: A Case Report and Review of Literature. *Cureus*. 2024; 16: e69794. <https://doi.org/10.7759/cureus.69794>.
- [63] O'Rourke DM, Nasrallah MP, Desai A, Melenhorst JJ, Mansfield K, Morrisette JJD, *et al.* A single dose of peripherally infused EGFRvIII-directed CAR T cells mediates antigen loss and induces adaptive resistance in patients with recurrent glioblastoma. *Science Translational Medicine*. 2017; 9: eaaa0984. <https://doi.org/10.1126/scitranslmed.aaa0984>.
- [64] Preusser M, Lim M, Hafler DA, Reardon DA, Sampson JH. Prospects of immune checkpoint modulators in the treatment of glioblastoma. *Nature Reviews. Neurology*. 2015; 11: 504–514. <https://doi.org/10.1038/nrneurol.2015.139>.
- [65] Medikonda R, Dunn G, Rahman M, Fecci P, Lim M. A review of glioblastoma immunotherapy. *Journal of Neuro-oncology*. 2021; 151: 41–53. <https://doi.org/10.1007/s11060-020-03448-1>.
- [66] Wang J, Shen F, Yao Y, Wang LL, Zhu Y, Hu J. Adoptive Cell Therapy: A Novel and Potential Immunotherapy for Glioblastoma. *Frontiers in Oncology*. 2020; 10: 59. <https://doi.org/10.3389/fonc.2020.00059>.
- [67] McGranahan T, Therkelsen KE, Ahmad S, Nagpal S. Current State of Immunotherapy for Treatment of Glioblastoma. *Current Treatment Options in Oncology*. 2019; 20: 24. <https://doi.org/10.1007/s11864-019-0619-4>.
- [68] Kaufman HL, Kohlhapp FJ, Zloza A. Oncolytic viruses: a new class of immunotherapy drugs. *Nature Reviews. Drug Discovery*. 2015; 14: 642–662. <https://doi.org/10.1038/nrd4663>.
- [69] Koch MS, Zdioruk M, Nowicki MO, Griffith AM, Aguilar-Cordova E, Aguilar LK, *et al.* Perturbing DDR signaling enhances cytotoxic effects of local oncolytic virotherapy and modulates the immune environment in glioma. *Molecular Therapy Oncolytics*. 2022; 26: 275–288. <https://doi.org/10.1016/j.omto.2022.07.009>.
- [70] Choi BD, Gerstner ER, Frigault MJ, Leick MB, Mount CW, Balaj L, *et al.* Intraventricular CARv3-TEAM-E T Cells in Recurrent Glioblastoma. *The New England Journal of Medicine*. 2024; 390: 1290–1298. <https://doi.org/10.1056/NEJMoa2314390>.
- [71] Pan C, Zhai Y, Wang C, Liao Z, Wang D, Yu M, *et al.* Poliovirus receptor-based chimeric antigen receptor T cells combined with NK-92 cells exert potent activity against glioblastoma. *Journal of the National Cancer Institute*. 2024; 116: 389–400. <https://doi.org/10.1093/jnci/djad226>.
- [72] Gardam B, Gargett T, Brown MP, Ebert LM. Targeting the dendritic cell-T cell axis to develop effective immunotherapies for glioblastoma. *Frontiers in Immunology*. 2023; 14: 1261257. <https://doi.org/10.3389/fimmu.2023.1261257>.
- [73] Shams F, Golchin A, Azari A, Mohammadi Amirabad L, Zarein F, Khosravi A, *et al.* Nanotechnology-based products for cancer immunotherapy. *Molecular Biology Reports*. 2022; 49: 1389–1412. <https://doi.org/10.1007/s11033-021-06876-y>.
- [74] Townsend SE, Allison JP. Tumor rejection after direct costimulation of CD8+ T cells by B7-transfected melanoma cells. *Science*. 1993; 259: 368–370. <https://doi.org/10.1126/science.7678351>.
- [75] Trojan J, Johnson TR, Rudin SD, Ilan J, Tykocinski ML, Ilan J. Treatment and prevention of rat glioblastoma by immunogenic C6 cells expressing antisense insulin-like growth factor I RNA. *Science*. 1993; 259: 94–97. <https://doi.org/10.1126/science.8418502>.
- [76] Habib NA. Cancer gene therapy - Post achievements and future challenges. Kluwer Academic/Plenum Publishers: New York. 2002.
- [77] Dietrich PY, Dutoit V, Tran Thang NN, Walker PR. T-cell immunotherapy for malignant glioma: toward a combined approach. *Current Opinion in Oncology*. 2010; 22: 604–610. <https://doi.org/10.1097/CCO.0b013e32833dead8>.
- [78] Trojan A, Kasprzak H, Gutierrez O, Penagos P, Briceno I, Siachoque H, *et al.* Neoplastic brain, glioblastoma and immunotherapy. In Morgan LR, Sarica FB (eds.) *Brain and spinal tumors - primary and secondary*. InTechOpen: UK. 2020.
- [79] Rubenstein JL, Nicolas JF, Jacob F. Nonsense RNA: a tool for specifically inhibiting the expression of a gene in vivo. *Comptes Rendus De L'Academie des Sciences. Serie III, Sciences De La Vie*. 1984; 299: 271–274. (In French)
- [80] Weintraub H, Izant JG, Harland RM. Anti-sense RNA as a molecular tool for ] genetic analysis. *Trends in Genetics*. 1985; 1: 22–25. [https://doi.org/10.1016/0168-9525\(85\)90010-1](https://doi.org/10.1016/0168-9525(85)90010-1).
- [81] Dervan PB. Reagents for the site-specific cleavage of megabase DNA. *Nature*. 1992; 359: 87–88. <https://doi.org/10.1038/359087a0>.
- [82] Hélène C. Control of oncogene expression by antisense nucleic acids. *European Journal of Cancer*. 1994; 30A: 1721–1726. [http://doi.org/10.1016/0959-8049\(93\)e0352-q](http://doi.org/10.1016/0959-8049(93)e0352-q).
- [83] Hau P, Jachimczak P, Schlaier J, Bogdahn U. TGF- $\beta$ 2 signaling in high-grade gliomas. *Current Pharmaceutical Biotechnology*. 2011; 12: 2150–2157. <https://doi.org/10.2174/138920111798808347>.
- [84] Pan Q, Chanthery Y, Liang WC, Stawicki S, Mak J, Rathore N, *et al.* Blocking neuropilin-1 function has an additive effect with anti-VEGF to inhibit tumor growth. *Cancer Cell*. 2007; 11: 53–67. <https://doi.org/10.1016/j.ccr.2006.10.018>.
- [85] Beckner ME, Gobbel GT, Abounader R, Burovic F, Agostino NR, Laterra J, *et al.* Glycolytic glioma cells with active glycogen synthase are sensitive to PTEN and inhibitors of PI3K and gluconeogenesis. *Laboratory Investigation; a Journal of Technical Methods and Pathology*. 2005; 85: 1457–1470. <https://doi.org/10.1038/labinvest.3700355>.
- [86] Premkumar DR, Arnold B, Jane EP, Pollack IF. Synergistic interaction between 17-AAG and phosphatidylinositol 3-kinase inhibition in human malignant glioma cells. *Molecular Carcinogenesis*. 2006; 45: 47–59. <https://doi.org/10.1002/mc.20152>.
- [87] Zeng KW, Wang XM, Ko H, Kwon HC, Cha JW, Yang HO. Hyperoside protects primary rat cortical neurons from neurotoxicity induced by amyloid  $\beta$ -protein via the PI3K/Akt/Bad/Bcl(XL)-regulated mitochondrial apoptotic pathway. *European Journal of Pharmacology*. 2011; 672: 45–55. <https://doi.org/10.1016/j.ejphar.2011.09.177>.
- [88] Fumarola C, Bonelli MA, Petronini PG, Alfieri RR. Targeting PI3K/AKT/mTOR pathway in non small cell lung cancer. *Biochemical Pharmacology*. 2014; 90: 197–207. <https://doi.org/10.1016/j.bcp.2014.05.011>.
- [89] Tao SC, Yuan T, Rui BY, Zhu ZZ, Guo SC, Zhang CQ. Exosomes derived from human platelet-rich plasma prevent apoptosis induced by glucocorticoid-associated endoplasmic reticulum stress in rat osteonecrosis of the femoral head via the Akt/Bad/Bcl-2 signal pathway. *Theranostics*. 2017; 7: 733–750. <https://doi.org/10.7150/thno.17450>.
- [90] Trojan J. Oncoproteins targeting: antibodies, antisense, triple - helix. Case of anti IGF-I cancer immunogene therapy. In Sharad S (ed.) *Antisense therapies*. InTechOpen: UK. 2019. <https://doi.org/10.5772/intechopen.82548>.
- [91] Quintero G, Guzman A, Gomez D, Kasparzk H, Penagos P, Siachoque H, *et al.* Glioblastoma - application of gene therapy during a quarter of a century: Anti - Gene IGF-I strategy. *Acta Scientifica Cancer Biology*. 2020; 4: 38–45.
- [92] Zhou H, Qian W, Uckun FM, Wang L, Wang YA, Chen H, *et al.* IGF1 Receptor Targeted Theranostic Nanoparticles for Targeted and Image-Guided Therapy of Pancreatic Cancer. *ACS Nano*.



- 2015; 9: 7976–7991. <https://doi.org/10.1021/acsnano.5b01288>.
- [93] Almeida SS, Girão AF, Gonçalves G, António Completo A, Marques PAAP. Stimulus responsive graphene scaffolds for tissue engineering. In Gonçalves G, Marques P, Vila M (eds.) *Graphene-based materials in health and environment* (pp. 219–256). Springer Publishing: NY. 2016. [https://doi.org/10.1007/978-3-319-45639-3\\_8](https://doi.org/10.1007/978-3-319-45639-3_8).
- [94] Krause NM, Bains JK, Blechar J, Richter C, Bessi I, Grote P, *et al.* Biophysical Investigation of RNA · DNA : DNA Triple Helix and RNA : DNA Heteroduplex Formation by the IncRNAs MEG3 and Fendrr. *Chembiochem: a European Journal of Chemical Biology*. 2024; 25: e202400049. <https://doi.org/10.1002/cbic.202400049>.
- [95] Rondanelli M, Fossari F, Vecchio V, Braschi V, Riva A, Allegrini P, *et al.* *Acmella oleracea* for pain management. *Fitoterapia*. 2020; 140: 104419. <https://doi.org/10.1016/j.fitote.2019.104419>.
- [96] Jeiter J, Hilger HH, Smets EF, Weigend M. The relationship between nectaries and floral architecture: a case study in Geraniaceae and Hypseocharitaceae. *Annals of Botany*. 2017; 120: 791–803. <https://doi.org/10.1093/aob/mcx101>.
- [97] Gao C, Yuan X, Jiang Z, Gan D, Ding L, Sun Y, *et al.* Regulation of AKT phosphorylation by GSK3 $\beta$  and PTEN to control chemoresistance in breast cancer. *Breast Cancer Research and Treatment*. 2019; 176: 291–301. <https://doi.org/10.1007/s10549-019-05239-3>.
- [98] Chahal DS, Sivamani RK, Isseroff RR, Dasu MR. Plant-based modulation of Toll-like receptors: an emerging therapeutic model. *Phytotherapy Research: PTR*. 2013; 27: 1423–1438. <https://doi.org/10.1002/ptr.4886>.
- [99] Pan W, Yu H, Huang S, Zhu P. Resveratrol Protects against TNF- $\alpha$ -Induced Injury in Human Umbilical Endothelial Cells through Promoting Sirtuin-1-Induced Repression of NF-KB and p38 MAPK. *PloS One*. 2016; 11: e0147034. <https://doi.org/10.1371/journal.pone.0147034>.
- [100] Trojan J, Raja M, Quintero G, Alvarez A, Siachoque HO, Lone Y-C, *et al.* Gene therapy of brain, liver and colon malignancies using anti – gene IGF-I approach. In Cosmi E (ed.) *Medicine and medical research: new perspectives* (vol. 6, pp. 85–103). BP International: UK. 2024. <https://doi.org/10.9734/bpi/mmmp/v6/2108>.
- [101] Ming GL, Song H. Adult neurogenesis in the mammalian brain: significant answers and significant questions. *Neuron*. 2011; 70: 687–702. <https://doi.org/10.1016/j.neuron.2011.05.001>.
- [102] Gage FH, Temple S. Neural stem cells: generating and regenerating the brain. *Neuron*. 2013; 80: 588–601. <https://doi.org/10.1016/j.neuron.2013.10.037>.
- [103] Gebara E, Bonaguidi MA, Beckervordersandforth R, Sultan S, Udry F, Gijis PJ, *et al.* Heterogeneity of Radial Glia-Like Cells in the Adult Hippocampus. *Stem Cells* (Dayton, Ohio). 2016; 34: 997–1010. <https://doi.org/10.1002/stem.2266>.
- [104] Pino A, Fumagalli G, Bifari F, Decimo I. New neurons in adult brain: distribution, molecular mechanisms and therapies. *Biochemical Pharmacology*. 2017; 141: 4–22. <https://doi.org/10.1016/j.bcp.2017.07.003>.
- [105] Daughaday WH, Rotwein P. Insulin-like growth factors I and II. Peptide, messenger ribonucleic acid and gene structures, serum, and tissue concentrations. *Endocrine Reviews*. 1989; 10: 68–91. <https://doi.org/10.1210/edrv-10-1-68>.
- [106] Yuan H, Chen R, Wu L, Chen Q, Hu A, Zhang T, *et al.* The regulatory mechanism of neurogenesis by IGF-1 in adult mice. *Molecular Neurobiology*. 2015; 51: 512–522. <https://doi.org/10.1007/s12035-014-8717-6>.
- [107] Ziegler AN, Levison SW, Wood TL. Insulin and IGF receptor signalling in neural-stem-cell homeostasis. *Nature Reviews. Endocrinology*. 2015; 11: 161–170. <https://doi.org/10.1038/nrendo.2014.208>.
- [108] Schlenska-Lange A, Knüpfer H, Lange TJ, Kiess W, Knüpfer M. Cell proliferation and migration in glioblastoma multiforme cell lines are influenced by insulin-like growth factor I in vitro. *Anticancer Research*. 2008; 28: 1055–1060.
- [109] Davis ME, Chen ZG, Shin DM. Nanoparticle therapeutics: an emerging treatment modality for cancer. *Nature Reviews. Drug Discovery*. 2008; 7: 771–782. <https://doi.org/10.1038/nrd2614>.
- [110] Giljohann DA, Seferos DS, Daniel WL, Massich MD, Patel PC, Mirkin CA. Gold nanoparticles for biology and medicine. *Ange wandte Chemie (International Ed. in English)*. 2010; 49: 3280–3294. <https://doi.org/10.1002/anie.200904359>.
- [111] Li L, Guo Q, Liu Y, Lu M, Yang J, Ge Y, *et al.* Targeted combination therapy for glioblastoma by co-delivery of doxorubicin, YAP-siRNA and gold nanorods. *Journal of Material Science Technology*. 2021; 63: 81–90. <https://doi.org/10.1016/j.jmst.2020.03.009>.
- [112] Gharatape A, Sadeghi-Abdonsari H, Seifalian A, Faridi-Majidi R, Basiri M. Nanocarrier-based gene delivery for immune cell engineering. *Journal of Materials Chemistry. B*. 2024; 12: 3356–3375. <https://doi.org/10.1039/d3tb02279j>.
- [113] Angelopoulou A. Nanostructured Biomaterials in 3D Tumor Tissue Engineering Scaffolds: Regenerative Medicine and Immunotherapies. *International Journal of Molecular Sciences*. 2024; 25: 5414. <https://doi.org/10.3390/ijms25105414>.
- [114] Schaub NJ, Johnson CD, Cooper B, Gilbert RJ. Electrospun Fibers for Spinal Cord Injury Research and Regeneration. *Journal of Neurotrauma*. 2016; 33: 1405–1415. <https://doi.org/10.1089/neu.2015.4165>.
- [115] El-Husseiny HM, Mady EA, Doghish AS, Zewail MB, Abdelfatah AM, Noshay M, *et al.* Smart/stimuli-responsive chitosan/gelatin and other polymeric macromolecules natural hydrogels vs. synthetic hydrogels systems for brain tissue engineering: A state-of-the-art review. *International Journal of Biological Macromolecules*. 2024; 260: 129323. <https://doi.org/10.1016/j.ijbiomac.2024.129323>.
- [116] Lancaster MA. Unraveling mechanisms of human brain evolution. *Cell*. 2024; 187: 5838–5857. <https://doi.org/10.1016/j.cell.2024.08.052>.
- [117] Rilling JK. Comparative primate neuroimaging: insights into human brain evolution. *Trends in Cognitive Sciences*. 2014; 18: 46–55. <https://doi.org/10.1016/j.tics.2013.09.013>.
- [118] Zhou Y, Song H, Ming GL. Genetics of human brain development. *Nature Reviews. Genetics*. 2024; 25: 26–45. <https://doi.org/10.1038/s41576-023-00626-5>.
- [119] Hejazi M, Tong W, Ibbotson MR, Prawer S, Garrett DJ. Advances in Carbon-Based Microfiber Electrodes for Neural Interfacing. *Frontiers in Neurosciences*. 2021; 15: 658703. <https://doi.org/10.3389/fnins.2021.658703>.
- [120] Lancaster MA. Brain organoids: A new frontier of human neuroscience research. *Seminars in Cell & Developmental Biology*. 2021; 111: 1–3. <https://doi.org/10.1016/j.semedb.2020.10.011>.
- [121] Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011; 144: 646–674. <https://doi.org/10.1016/j.cell.2011.02.013>.
- [122] Dakal TC, Kakde GS, Maurya PK. Genomic, epigenomic and transcriptomic landscape of glioblastoma. *Metabolic Brain Disease*. 2024; 39: 1591–1611. <https://doi.org/10.1007/s11011-024-01414-8>.
- [123] Neufeld L, Yeini E, Pozzi S, Satchi-Fainaro R. 3D bioprinted cancer models: from basic biology to drug development. *Nature Reviews. Cancer*. 2022; 22: 679–692. <https://doi.org/10.1038/s41568-022-00514-w>.
- [124] Tirosh I, Izar B, Prakadan SM, Wadsworth MH, 2nd, Treacy D, Trombetta JJ, *et al.* Dissecting the multicellular ecosystem of metastatic melanoma by single-cell RNA-seq. *Science*. 2016;

- 352: 189–196. <https://doi.org/10.1126/science.aad0501>.
- [125] Alrushaid N, Khan FA, Al-Suhaimi EA, Elaissari A. Nanotechnology in Cancer Diagnosis and Treatment. *Pharmaceutics*. 2023; 15: 1025. <https://doi.org/10.3390/pharmaceutics15031025>.
- [126] Jadhav V, Roy A, Kaur K, Rai AK, Rustagi S. Recent advances in nanomaterial-based drug delivery systems. *Nano-Structures & Nano-Objects*. 2024; 37: 101103. <https://doi.org/10.1016/j.nanos.2024.101103>.
- [127] Salazar A, Pérez-de la Cruz V, Muñoz-Sandoval E, Chavarria V, Garcia Morales MDL, Espinosa-Bonilla A, *et al.* Potential Use of Nitrogen-Doped Carbon Nanotube Sponges as Payload Carriers Against Malignant Glioma. *Nanomaterials*. 2021; 11: 1244. <https://doi.org/10.3390/nano11051244>.
- [128] Katti A, Diaz BJ, Caragine CM, Sanjana NE, Dow LE. CRISPR in cancer biology and therapy. *Nature Reviews. Cancer*. 2022; 22: 259–279. <https://doi.org/10.1038/s41568-022-00441-w>.
- [129] Zhang P, Meng J, Li Y, Yang C, Hou Y, Tang W, *et al.* Nanotechnology-enhanced immunotherapy for metastatic cancer. *Innovation*. 2021; 2: 100174. <https://doi.org/10.1016/j.xinn.2021.100174>.
- [130] Pandian SR, Rencilin CF, Sundar K. Emerging nanomaterials for cancer immunotherapy. *Exploration of Medicine*. 2021; 2: 208–231. <https://doi.org/10.37349/emed.2021.00043>.
- [131] Zhu X, Li S. Nanomaterials in tumor immunotherapy: new strategies and challenges. *Molecular Cancer*. 2023; 22: 94. <https://doi.org/10.1186/s12943-023-01797-9>.
- [132] Akabari AH, Patel S, Vaghela N, Ramani V, Shah DP. Recent application of nanotechnology for cancer immunotherapy and its future prospects. *International Journal of Immunology and Immunotherapy*. 2023; 10: 69. <https://doi.org/10.23937/2378-3672/1410069>.
- [133] Yadav D, Puranik N, Meshram A, Chavda V, Lee PCW, Jin JO. How Advanced are Cancer Immuno-Nanotherapeutics? A Comprehensive Review of the Literature. *International Journal of Nanomedicine*. 2023; 18: 35–48. <https://doi.org/10.2147/IJN.S388349>.
- [134] Grzegorzewski J, Michalak M, Wołoszczuk M, Bulicz M, Majchrzak-Celińska A. Nanotherapy of Glioblastoma-Where Hope Grows. *International Journal of Molecular Sciences*. 2025; 26: 1814. <https://doi.org/10.3390/ijms26051814>.
- [135] Rad DM, Nazari H, Naei VY, Lotfi M, Aref AR, Warkiani ME. Cancer nanotechnology: a new approach to upgrade cancer diagnosis and therapy. In Barabadi H (ed.). *Functionalized Nanomaterials for Cancer Research*. Elsevier Inc: USA. 2024. <https://doi.org/10.1016/B978-0-443-15518-5.00013-6>.
- [136] Chaturvedi VK, Singh A, Singh VK, Singh MP. Cancer Nanotechnology: A New Revolution for Cancer Diagnosis and Therapy. *Current Drug Metabolism*. 2019; 20: 416–429. <https://doi.org/10.2174/1389200219666180918111528>.
- [137] de Santana WMOS, Surur AK, Momesso VM, Lopes PM, Santilli CV, Fontana CR. Nanocarriers for photodynamic-gene therapy. *Photodiagnosis and Photodynamic Therapy*. 2023; 43: 103644. <https://doi.org/10.1016/j.pdpdt.2023.103644>.
- [138] Habeeb M, Vengateswaran HT, You HW, Saddhono K, Aher KB, Bhavar GB. Nanomedicine facilitated cell signaling blockade: difficulties and strategies to overcome glioblastoma. *Journal of Materials Chemistry. B*. 2024; 12: 1677–1705. <https://doi.org/10.1039/d3tb02485g>.
- [139] Kang S, Duan W, Zhang S, Chen D, Feng J, Qi N. Muscone/RI7217 co-modified upward messenger DTX liposomes enhanced permeability of blood-brain barrier and targeting glioma. *Theranostics*. 2020; 10: 4308–4322. <https://doi.org/10.7150/thno.41322>.
- [140] Zhao X, He J, Chen Y, Zheng J, Li X, Fu T, *et al.* Transferin receptor-targeted aptamer-drug conjugate overcomes blood-brain barrier for potent glioblastoma therapy. *Bioconjugate Chemistry*. 2025; 3: 1288–1298. <https://doi.org/10.1021/acs.bioconjchem.5c00137>.
- [141] Liaw K, Sharma R, Sharma A, Salazar S, Appiani La Rosa S, Kannan RM. Systemic dendrimer delivery of triptolide to tumor-associated macrophages improves anti-tumor efficacy and reduces systemic toxicity in glioblastoma. *Journal of Controlled Release: Official Journal of the Controlled Release Society*. 2021; 329: 434–444. <https://doi.org/10.1016/j.jconrel.2020.12.003>.
- [142] Zhu Y, Liang J, Gao C, Wang A, Xia J, Hong C, *et al.* Multifunctional ginsenoside Rg3-based liposomes for glioma targeting therapy. *Journal of Controlled Release: Official Journal of the Controlled Release Society*. 2021; 330: 641–657. <https://doi.org/10.1016/j.jconrel.2020.12.036>.