

Commentary

Glioblastoma Stem Cells as Targets for Emerging Precision Immunotherapies and Molecular Treatments

Dennis A. Steindler ^{1,2,3,*}  and Katherine Karakoula ⁴

¹ Steindler Consulting, Boston, MA, USA

² Round Table Research, Inc., Research Triangle Park, Chapel Hill, NC 27709, USA

³ The Eshelman Institute for Innovation, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA

⁴ School of Pharmacy and Life Sciences, Research Institute of Healthcare Sciences, University of Wolverhampton, City Campus, Wolverhampton WV1 1LY, UK; a.karakoula@wlv.ac.uk

* Correspondence: stemcellguy@gmail.com

Abstract

Advances in organoid and other three-dimensional culture systems, single-cell and spatial transcriptomics, multi-omics, and high-resolution imaging are reshaping our understanding of the cellular origins and evolutionary trajectories of glioblastoma. When integrated with modern data science approaches, these technologies enable the construction of increasingly detailed molecular biographies of normal neural stem and progenitor cells as well as malignant stem-like cellular states. Such molecular biographies illuminate how developmental programs, cellular plasticity, and microenvironmental cues are co-opted during gliomagenesis. At the same time, progress in machine learning, immunotherapy, and precision molecular targeting is beginning to translate these biological insights into therapeutic strategies that specifically disrupt glioblastoma stem-like states. Together, these converging approaches provide a conceptual and technological framework for improved tumor modeling, earlier detection, and increasingly personalized therapies for malignant gliomas.

Keywords: neurogenesis; glioma; glioblastoma; cancer stem cell; molecular and immunotherapies; data science

1. Introduction: Normal and Pathological Stem Cell Neurogenesis

The concept of persistent cell genesis in the adult brain—“neurogenesis”—has been debated for more than a century [1]. Early work by Ramón y Cajal argued that the adult mammalian brain lacked regenerative capacity, establishing a long-standing belief that no new neurons were produced after development [2]. This view began to shift in the 1990s with the discovery of neural stem/progenitor cells in the adult rodent and human subependymal zone (SEZ) and hippocampus, demonstrating that limited neurogenesis persists throughout life [3–6]. These findings launched a modern re-evaluation of adult neurogenesis, supported by studies identifying proliferative progenitors in neurogenic niches of the human brain.

Yet the extent and functional relevance of adult neurogenesis remain controversial. Some studies report robust neurogenesis in the adult hippocampus [4,6], while others describe a sharp decline to near undetectable levels after childhood [7]. Recent single-cell and spatial transcriptomic analyses have revived the debate by identifying rare but persistent



Academic Editor: Pablo Martín-Vasallo

Received: 10 February 2026

Revised: 20 April 2026

Accepted: 24 April 2026

Published: 26 April 2026

Copyright: © 2026 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the [Creative Commons Attribution \(CC BY\)](https://creativecommons.org/licenses/by/4.0/) license.

neural progenitors even in the aged human hippocampus [8–10], suggesting that neurogenesis may be more heterogeneous and context-dependent than previously appreciated.

In contrast, hematopoiesis provides a clear example of lifelong stem cell-driven tissue renewal, highlighting the unique regenerative limitations of the adult brain and heart. While epithelial and mesenchymal tissues maintain substantial regenerative capacity, the central nervous system remains largely refractory to repair following injury or disease [11].

2. Stem Cell Pathologies: “Good” and “Bad” Stem Cells

The modern concept of a stem cell originates from pioneering work by Stevens, Martin, Evans, Reynolds, and Weiss, who established the foundations of embryonic and adult stem cell biology [12]. Subsequent studies identified multipotent astrocytic stem/progenitor cells (MASCs) and adult human neural progenitor cells (AHNPs) in the human brain, particularly within the olfactory system and hippocampus [13–15]. These populations exhibit stem-like properties but contribute primarily to gliogenesis rather than neurogenesis during adult life. This leads to an important conceptual framework: Adult stem cells can be “good” or “bad,” depending on whether they maintain tissue homeostasis or contribute to pathology.

Normal adult stem cells may support limited gliogenesis during aging, respond to injury with restricted regenerative potential and fail to replace neurons lost to trauma or neurodegenerative disease. Conversely, pathological stem cells may proliferate excessively, acquire mutations, generate aberrant progeny and initiate neoplastic growth.

This duality forms the basis of stem cell pathologies, in which stem/progenitor cells either fail to repair damaged tissue or contribute to hyperplasia and tumorigenesis [16]. The inability of adult neural stem/progenitor cells to restore circuitry lost to stroke, traumatic injury, or neurodegenerative disease (e.g., Parkinson’s disease) exemplifies one form of stem cell pathology [17]. The opposite extreme—excessive proliferation and malignant transformation—underlies the emergence of brain tumors such as glioblastoma.

The same neural stem and progenitor cell populations that support limited adult neurogenesis can, under pathological conditions, become the cells of origin for gliomas. Understanding how normal stem cell biology intersects with malignant transformation requires examining the ontogeny, lineage relationships, and microenvironmental influences that drive gliomagenesis.

3. Ontogeny and Cells of Origin of Gliomas

3.1. From Normal Stem Cells to Cancer Stem Cells

The cancer stem cell (CSC) concept was first established in hematologic malignancies [18,19], where rare stem-like cells were shown to initiate leukemia following transplantation. This framework was soon extended to solid tumors, including breast cancer—coinciding with the identification of normal mammary stem cells—and subsequently to osteosarcoma, colon cancer, prostate cancer, and gliomas [20–25]. In gliomas, multiple groups identified stem-like populations capable of self-renewal, multilineage differentiation, and tumor initiation in xenograft models [22–24].

Glioma-initiating cells (GICs) are thought to arise from glial lineage stem or progenitor cells that retain latent proliferative capacity. In growth-permissive microenvironments—such as those shaped by inflammation, injury, or reactive gliosis—these cells may re-enter the cell cycle and accumulate oncogenic mutations. Candidate cells include multipotent astrocytic stem/progenitor cells (MASCs) [1,12,13], adult human neural progenitor cells (AHNPs) [13], vestigial boundary astrocytes in the gray matter and persistently reactive white matter astrocytes [25], and oligodendrocyte precursor cells (OPCs/NG2+ cells). Although these progenitor populations are not able to regenerate neuronal or glial populations—a limitation

sometimes described as a “stem cell pathology”—their retained developmental programs and proliferative potential make them susceptible to malignant transformation.

3.2. Neurogenic Niches as Tumorigenic Niches

The developmental subventricular zone (SVZ) and the adult subependymal zone (SEZ) [6] are lifelong neurogenic niches enriched in extracellular matrix components, morphogens, and growth factors. Normal neurogenesis in these regions involves periods of rapid proliferation during which neuronal progenitor cells are vulnerable to genetic instability. During differentiation, progenitors may undergo chromosomal instability, including aneuploidy [26], providing opportunities for oncogenic lesions to arise.

This vulnerability is heightened during periods of inflammation, aging, injury, and reactive gliosis, all of which generate microenvironments that promote progenitor activation and cycling. These conditions support the view that gliomagenesis often emerges from endogenous progenitors placed into pathological states of proliferation. OPCs (NG2+ cells), which normally generate oligodendrocytes and astrocytes, have been strongly implicated as cells of origin for IDH-mutant gliomas [14]. Likewise, astrocytic stem/progenitor cells residing in the SVZ/SEZ can give rise to tumors such as subependymal giant cell astrocytoma (SEGA), particularly in the setting of TSC1/TSC2 mutations [27]. The SEZ exemplifies how a lifelong neurogenic niche—rich in proliferative cues—can support the emergence and maintenance of tumorigenic stem-like cells.

The precise origins of IDH-mutant versus IDH wild-type gliomas remain an area of active investigation and debate [28]. It is increasingly clear that gliomas may arise from multiple progenitor pools depending on developmental timing, regional context, and mutational events. Identifying and therapeutically targeting the true cells of origin remain challenging and will require longitudinal human tissue studies, including xenotransplantation of patient-derived CSC populations into rodent “avatar” models [29]. Such approaches allow researchers to observe tumor initiation, evolution, and therapeutic response in vivo, providing critical insights into the earliest events of gliomagenesis.

3.3. Lineage Tracing, Clonal Evolution, and Tumor Heterogeneity

Lineage tracing studies in genetically engineered mouse models, including NF1- and PDGF-driven glioblastomas, demonstrate that glial progenitors are capable of initiating high-grade tumors [30]. Genetic barcoding approaches further reveal early clonal extinction, intercellular competition, and dynamic clonal evolution during glioma formation [31]. These processes underlie the profound intratumoral and intertumoral heterogeneity characteristic of glioblastoma.

Recent single-cell RNA sequencing, spatial transcriptomics, and multimodal longitudinal profiling have refined our understanding of glioma ontogeny [32–35]. Distinct cellular states—including stem-like, mesenchymal, oligodendrocyte-like, and astrocyte-like populations—coexist within individual tumors and shift in response to therapy. Spatial niches such as perivascular and hypoxic regions impose selective pressures that sustain particular GIC states, influence clonal competition, and contribute to therapeutic resistance.

Taken together, these findings support a model in which gliomas arise from a restricted set of glial lineage stem and progenitor populations residing within specialized neurogenic or inflammatory niches. Yet identifying the initial cell of origin is only the first step. Translating ontogeny into therapy requires reconstructing the “molecular biography” of glioma-initiating cells: the sequential genetic lesions they acquire, the developmental programs they retain, the microenvironmental cues that maintain them, and the immune interactions that shape their evolution. We propose a conceptual framework in which

glioblastoma evolves from neural ontogeny through the acquisition of a tumor-specific molecular biography toward precision immuno-molecular intervention (Figure 1).

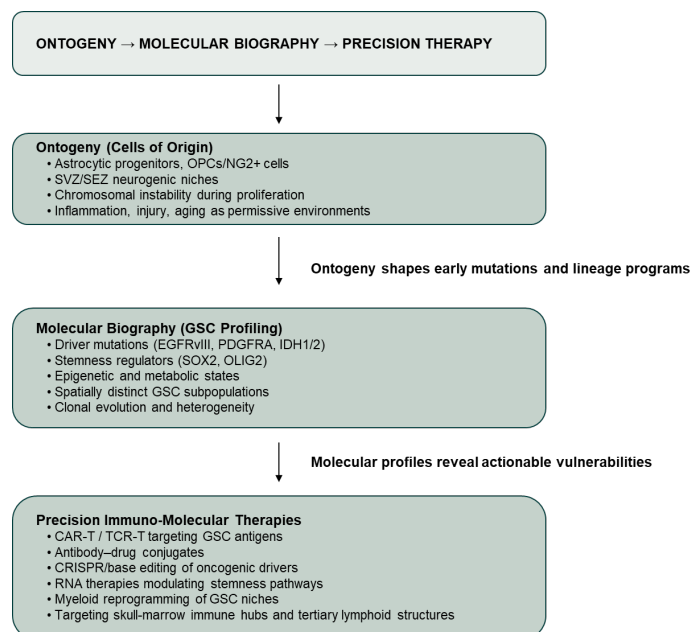


Figure 1. Ontogeny to precision therapy in glioblastoma. Glioblastoma is conceptualized as a progressive transformation from neural ontogeny, through acquisition of a tumor-specific molecular biography, toward therapeutically tractable stem-like states. Cells of origin within astrocytic and oligodendrocyte lineage progenitor pools, shaped by permissive microenvironments, influence early mutational trajectories and lineage programs. Molecular profiling of glioblastoma stem cell (GSC) populations reveals genetic, epigenetic, metabolic and spatial vulnerabilities that inform precision immuno-molecular therapeutic strategies.

4. Molecular Biographies of Glioma Cells as a Foundation for Immunotherapy and Precision Medicine

The expanding molecular and cellular “biographies” of glioma stem and progenitor cells have revealed a complex landscape of genetic, epigenetic, and microenvironmental determinants that shape gliomagenesis. These insights raise an essential question: if we now possess a detailed understanding of the molecular phenotype of glioma-initiating cells, why do effective targeted therapies remain elusive?

Meaningful progress is being made. Standard-of-care DNA-alkylating agents such as temozolomide remain foundational, and targeted therapies directed at oncogenic pathways—including EGFR, STAT3, and Wnt—continue to evolve. Novel EGFR variants such as EGFRx maintain glioblastoma stemness through STAT5 activation [36], while STAT3 remains essential for proliferation and multipotency [37]. Wnt-mediated endothelial-to-mesenchymal transitions contribute to chemoresistance and tumor progression [38]. Importantly, these pathways are not merely generic oncogenic drivers; they are central regulators of glioblastoma stem cell (GSC) self-renewal, lineage plasticity, and apoptotic resistance. Consequently, pathway-directed therapies are increasingly informed by the biology of stem-like cellular states rather than by mutational status alone. Contemporary therapeutic strategies therefore converge on GSC states through complementary mechanisms targeting intrinsic survival programs, immune-mediated cytotoxicity and niche- and system-level modulators (Figure 2)

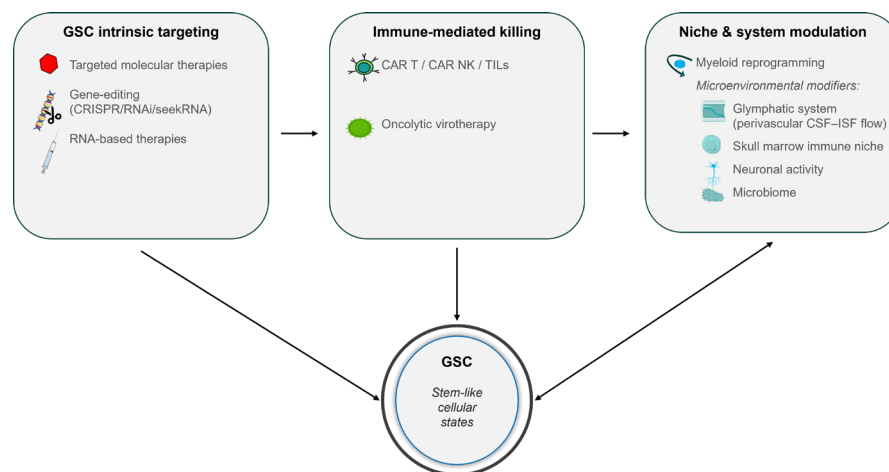


Figure 2. Therapeutic convergence on glioblastoma stem-like cellular states.

Contemporary precision therapies for glioblastoma increasingly target stem-like cellular states that sustain tumor propagation, cellular plasticity and therapeutic resistance. GSC-intrinsic strategies include targeted molecular therapies, RNA-based platforms and emerging gene-editing approaches that disrupt self-renewal and survival programs. Immune-mediated killing is achieved through cellular immunotherapies, including chimeric antigen receptor (CAR) T cells, CAR natural killer (NK) cells and tumor-infiltrating lymphocytes (TILs), as well as oncolytic virotherapy. In parallel, niche- and system-level modulation—including myeloid reprogramming, glymphatic transport (perivascular cerebrospinal fluid–interstitial fluid flow), skull marrow immune niches, neuronal activity, and microbiome-associated influences—shapes GSC maintenance and therapeutic responsiveness. Together, these complementary approaches emphasize a state-centric model of therapeutic intervention in glioblastoma.

Within this framework, therapeutic innovation in glioblastoma increasingly emphasizes coordinated disruption of stem-like cellular states through direct targeting of GSC-intrinsic programs, immune-mediated killing and modulation of permissive niches and systemic regulators.

In parallel, immunotherapy platforms have expanded considerably, including antibody–drug conjugates (ADCs) targeting EGFR, HER2, and other glioma-associated antigens offering selective cytotoxic delivery [39]. RNA-based immunotherapies—including mRNA vaccines, miRNA modulators, and RNA aggregates—are rapidly advancing. RNA aggregates, in particular, can engage innate danger-signal pathways to drive potent anti-tumor immunity [40]. Engineered extracellular vesicles, such as TRAIL-loaded exosomes derived from induced neural stem cells, provide a biocompatible delivery system for RNA and protein therapeutics [41].

Advances in base editing and CRISPR–Cas9 technologies have progressed from *in vitro* applications to *in vivo* gene correction and oncogene disruption in glioma models [42]. Hybrid guide RNAs improve the specificity and efficiency of adenine base editing [43], while emerging systems such as SeekRNA offer programmable RNA-guided editing with reduced off-target activity [44]. These platforms allow for direct *in vivo* modification of oncogenic drivers or restoration of tumor suppressors, offering unprecedented precision in targeting the molecular lesions that sustain GSCs.

Adoptive cellular therapies also continue to advance. CAR T cells targeting IL13R α 2, EGFRvIII, HER2, and GD2 have demonstrated safety and early signs of efficacy in glioblastoma clinical trials [45–47]. CAR NK cells offer additional advantages, including reduced toxicity and the possibility of universal donor-derived products. Tumor-infiltrating lym-

phocyte (TIL) therapies are gaining traction as single-cell profiling reveals resident tumor-reactive lymphocyte subsets. Despite limited efficacy in unselected GBM populations, immune checkpoint inhibitors remain central to immunotherapy research; PD-1, PD-L1, CTLA-4, TIM-3, and LAG-3 blockade may be effective when combined with vaccines, oncolytic viruses, or myeloid-modulating therapies [48,49].

Oncolytic virotherapy—including adenovirus DNX-2401, poliovirus chimera PVS-RIPO, and HSV-based vectors—induces immunogenic tumor cell death, reshapes the tumor microenvironment, and synergizes with ICIs [50,51]. Given that glioblastoma is dominated by immunosuppressive myeloid cells, strategies targeting CSF1R, CD47–SIRP α , TREM2, and other myeloid checkpoints aim to reprogram these populations toward antitumor phenotypes [52].

With single-cell sequencing, spatial transcriptomics, and lineage tracing in their prime, a detailed catalog is emerging of both normal neurogenic and aberrantly neurogenic glioma-like states. These datasets reveal how adult brain stem cells, when forced back into proliferative states—whether through aging, injury, inflammation, or spontaneous mutation—become vulnerable to oncogenic transformation. Additional insights into activity-dependent tumorigenesis [53], the influence of the glymphatic system on inflammatory tone [54], and the discovery of skull-marrow immune niches containing complete lymphoid and myeloid lineages [55,56] further expand the therapeutic landscape. Newly identified tertiary-lymphoid-like structures connected to the tumor via meningeal bridges [56] represent novel targets for immunomodulation and drug delivery.

5. Future Directions: Toward Precision and Personalization

Recurrent gliomas exemplify the evolutionary capacity of malignant systems, frequently undergoing profound genotypic and phenotypic remodeling marked by the re-emergence of stem cell-like states [57] and the adoption of noncanonical modes of intercellular communication. Transcellular transfer of organelles—including mitochondria and RNA—between tumor cells and stromal elements [58] has emerged as a potent mechanism by which gliomas enhance metabolic fitness, evade therapy, and increase tumorigenicity. These processes reinforce the view of gliomas as adaptive, cooperative cellular ecosystems rather than collections of independently evolving clones.

These adaptive, spatially structured stem-like states generate levels of biological complexity that exceed the capacity of linear biomarker-based approaches, providing a strong rationale for AI-driven integrative models in glioblastoma.

Beyond tumor-intrinsic programs, systemic modifiers of gliomagenesis are gaining increasing attention. The gut and brain microbiomes influence immune tone, metabolism and therapeutic responsiveness, suggesting that nutritional status and microbial composition may shape both tumor initiation and treatment outcomes [59]. Integrating systemic physiology with tumor and microenvironmental biology broadens the conceptual framework of neuro-oncology and suggests new opportunities for metabolic, immune and microbiome-directed interventions.

Artificial intelligence provides a unifying framework to translate this multiscale complexity into actionable therapeutic strategies. Machine learning approaches integrating multi-omic, spatial, and imaging data are increasingly capable of identifying druggable dependencies, modeling cellular states, and stratifying patients based on evolving molecular and immunological features [60]. Active learning frameworks coupled with transcriptomic profiling and advances in spatial omics and high-resolution imaging further enable region-specific analysis of tumor architecture, lineage relationships and immune infiltration, supporting rational, adaptive deployment of combination therapies [61,62].

Innovations in data science will increasingly determine the timing, selection, and combination of immune-based, molecular, and cell-directed therapies for glioblastoma and related cancer stem cell driven diseases.

Author Contributions: K.K. and D.A.S. both contributed equally to the ideas and writing of the paper. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: No new data were created or analyzed in this study.

Acknowledgments: Steindler, D.A. would like to thank his family, his mentors and collaborators in neuroscience and developmental biology, Parkinson's, Alzheimer's disease, heart failure, oncology and AI, who have given him the tools and knowledge to try and help all who are challenged by degenerative diseases and cancer.

Conflicts of Interest: The authors declare no conflicts of interest. Steindler, D.A. is a founder, shareholder and Chief Scientific Officer of Round Table Research Inc., RTP, North Carolina. There are no conflicts of interest with that work and the work presented here.

References

1. Steindler, D.A.; Pincus, D. Stem cells and neurogenesis in the adult human brain. *Lancet* **2002**, *359*, 1047–1054. [[CrossRef](#)]
2. Ramon y Cajal, S. *Degeneration and Regeneration of the Nervous System*; May, R.M., Translator; Oxford University Press, 1928.
3. Reynolds, B.A.; Weiss, S. Generation of neurons and astrocytes from isolated cells of the adult mammalian central nervous system. *Science* **1992**, *255*, 1707–1710. [[CrossRef](#)]
4. Eriksson, P.S.; Perfilieva, E.; Björk-Eriksson, T.; Alborn, A.M.; Nordborg, C.; Peterson, D.A.; Gage, F.H. Neurogenesis in the adult human hippocampus. *Nat. Med.* **1998**, *4*, 1313–1317. [[CrossRef](#)]
5. Roy, N.S.; Wang, S.; Jiang, L.; Kang, J.; Benraiss, A.; Harrison-Restelli, C.; Fraser, R.A.; Couldwell, W.T.; Kawaguchi, A.; Okano, H.; et al. In vitro neurogenesis by progenitor cells isolated from the adult human hippocampus. *Nat. Med.* **2000**, *6*, 271–277. [[CrossRef](#)]
6. Kukekov, V.G.; Laywell, E.D.; Suslov, O.; Davies, K.; Scheffler, B.; Thomas, L.B.; O'Brien, T.F.; Kusakabe, M.; Steindler, D.A. Multipotent stem/progenitor cells arise from two neurogenic regions of adult human brain. *Exp. Neurol.* **1999**, *156*, 333–344. [[CrossRef](#)]
7. Sorrells, S.F.; Paredes, M.F.; Cebrian-Silla, A.; Sandoval, K.; Qi, D.; Kelley, K.W.; James, D.; Mayer, S.; Chang, J.; Auguste, K.I.; et al. Human hippocampal neurogenesis drops sharply in children to undetectable levels in adults. *Nature* **2018**, *555*, 377–381. [[CrossRef](#)] [[PubMed](#)]
8. Boldrini, M.; Fulmore, C.A.; Tartt, A.N.; Simeon, L.R.; Pavlova, I.; Poposka, V.; Rosoklija, G.B.; Stankov, A.; Arango, V.; Dwork, A.J.; et al. Human hippocampal neurogenesis persists throughout aging. *Cell Stem Cell* **2018**, *22*, 589–599.e5. [[CrossRef](#)] [[PubMed](#)]
9. Moreno-Jiménez, E.P.; Flor-García, M.; Terreros-Roncal, J.; Rábano, A.; Cafini, F.; Pallas-Bazarra, N.; Ávila, J.; Llorens-Martín, M. Adult hippocampal neurogenesis is abundant in neurologically healthy subjects. *Nat. Med.* **2019**, *25*, 554–560. [[CrossRef](#)]
10. Tobin, M.K.; Musaraca, K.; Disouky, A.; Shetti, A.; Bheri, A.; Honer, W.G.; Kim, N.; Dawe, R.J.; Bennett, D.A.; Arfanakis, K.; et al. Human Hippocampal Neurogenesis Persists in Aged Adults and Alzheimer's Disease Patients. *Cell Stem Cell* **2019**, *24*, 974–982.e3. [[CrossRef](#)] [[PubMed](#)]
11. Weissman, I.L.; Shizuru, J.A. Origins of hematopoietic stem cells. *Blood* **2008**, *112*, 3543–3553. [[CrossRef](#)]
12. Laywell, E.D.; Rakic, P.; Kukekov, V.G.; Holland, E.; Steindler, D.A. Identification of a multipotent astrocytic stem cell. *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 13883–13888. [[CrossRef](#)]
13. Walton, N.M.; Sutter, B.M.; Chen, H.X.; Chang, L.J.; Roper, S.N.; Scheffler, B.; Steindler, D.A. Derivation and large-scale expansion of multipotent astroglial neural progenitors from adult human brain. *Development* **2006**, *133*, 3671–3681. [[CrossRef](#)]
14. Park, J.W.; Kwak, J.; Kim, K.W.; Jung, S.; Nam, C.H.; Kim, H.J.; Lee, S.M.; Choi, C.; Ahn, Y.; Park, J.H.; et al. IDH-mutant gliomas arise from glial progenitor cells harboring the initial driver mutation. *Science* **2026**, *391*, eadt0559. [[CrossRef](#)]
15. Ignatova, T.N.; Kukekov, V.G.; Laywell, E.D.; Suslov, O.N.; Vrionis, F.D.; Steindler, D.A. Human cortical glial tumors contain neural stem like cells. *Glia* **2002**, *39*, 193–206. [[CrossRef](#)] [[PubMed](#)]

16. Steindler, D.A.; Okun, M.; Scheffler, B. Stem cell pathologies and neurological disease. *Mod. Pathol.* **2012**, *25*, 157–162. [[CrossRef](#)] [[PubMed](#)]
17. Wang, S.; Okun, M.S.; Suslov, O.; Zheng, T.; McFarland, N.R.; Vedam-Mai, V.; Foote, K.D.; Roper, S.N.; Yachnis, A.T.; Siebzehnrubl, F.A.; et al. Neurogenic potential of progenitor cells isolated from postmortem human Parkinsonian brains. *Brain Res.* **2012**, *1464*, 61–72. [[CrossRef](#)] [[PubMed](#)]
18. Lapidot, T.; Sirard, C.; Vormoor, J.; Murdoch, B.; Caceres-Cortes, J.; Minden, M.; Paterson, B.; Caligiuri, M.A.; Dick, J.E. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature* **1994**, *367*, 645–648. [[CrossRef](#)]
19. Dontu, G.; Liu, S.; Wicha, M.S. Mammary stem cells in carcinogenesis. *Cell Rev.* **2005**, *1*, 207–213. [[CrossRef](#)]
20. Gibbs, C.P.; Kukekov, V.G.; Reith, J.D.; Tchigrinova, O.; Suslov, O.N.; Scott, E.W.; Ghivizzani, S.C.; Ignatova, T.N.; Steindler, D.A. Stem-like cells in bone sarcomas: Implications for tumorigenesis. *Neoplasia* **2005**, *7*, 967–976. [[CrossRef](#)]
21. Carpentino, J.E.; Hynes, M.J.; Appelman, H.D.; Zheng, T.; Steindler, D.A.; Scott, E.W.; Huang, E.H. Aldehyde dehydrogenase-expressing colon stem cells contribute to tumorigenesis in the transition from colitis to cancer. *Cancer Res.* **2009**, *69*, 8208–8215. [[CrossRef](#)]
22. Bae, K.M.; Su, Z.; Frye, C.; McClellan, S.; Allan, R.W.; Andrejewski, J.T.; Kelley, V.; Jorgensen, M.; Steindler, D.A.; Vieweg, J.; et al. Expression of pluripotent stem cell reprogramming factors by prostate tumor initiating cells. *J. Urol.* **2010**, *183*, 2045–2053. [[CrossRef](#)]
23. Singh, S.K.; Clarke, I.D.; Terasaki, M.; Bonn, V.E.; Hawkins, C.; Squire, J.; Dirks, P.B. Identification of a cancer stem cell in human brain tumors. *Cancer Res.* **2003**, *63*, 5821–5828. [[PubMed](#)]
24. Riddick, G.; Kotliarova, S.; Rodriguez, V.; Kim, H.S.; Linkous, A.; Storaska, A.J.; Ahn, S.; Walling, J.; Belova, G.; Fine, H.A. A core regulatory circuit in glioblastoma stem cells links MAPK activation to transcriptional program of neural stem cell identity. *Sci. Rep.* **2017**, *7*, 43605. [[CrossRef](#)]
25. Steindler, D.A. Glial boundaries in the developing nervous system. *Annu. Rev. Neurosci.* **1993**, *16*, 445–470. [[CrossRef](#)]
26. Walton, N.M.; Snyder, G.E.; Park, D.; Kobeissy, F.; Scheffler, B.; Steindler, D.A. Gliotypic neural stem cells transiently adopt tumorigenic properties during normal differentiation. *Stem Cells* **2009**, *27*, 280–289. [[CrossRef](#)] [[PubMed](#)]
27. Avgeris, S.; Fostira, F.; Vagena, A.; Ninios, Y.; Delimitsou, A.; Vodicka, R.; Vrtel, R.; Youroukos, S.; Stravopodis, D.J.; Vlassi, M.; et al. Mutational analysis of TSC1 and TSC2 genes in Tuberous Sclerosis Complex patients from Greece. *Sci. Rep.* **2017**, *7*, 16697. [[CrossRef](#)] [[PubMed](#)]
28. Lucas, C.G.; Al-Adli, N.N.; Young, J.S.; Gupta, R.; Morshed, R.A.; Wu, J.; Ravindranathan, A.; Shai, A.; Oberheim Bush, N.A.; Taylor, J.W.; et al. Longitudinal multimodal profiling of IDH-wildtype glioblastoma reveals the molecular evolution and cellular phenotypes underlying prognostically different treatment responses. *Neuro Oncol.* **2025**, *27*, 89–105. [[CrossRef](#)]
29. Feng, Y.; Haupt, B.; Huynh, T.T.; Meshaw, R.; Martin-Regalado, A.; Thakur, A.; Duffy, J.T.; Alzeer, A.; Siegel, D.A.; Barnes, A.; et al. Longitudinal Imaging Reveals Tumor Uptake and Prolonged Retention of Bispecific T Cell-Engaging Antibody in GBM via Passive and Active Mechanisms. *Clin. Cancer Res.* **2025**, *31*, 3537–3549. [[CrossRef](#)]
30. Hambardzumyan, D.; Cheng, Y.K.; Haeno, H.; Holland, E.C.; Michor, F. The probable cell of origin of NF1- and PDGF-driven glioblastomas. *PLoS ONE* **2011**, *6*, e24454. [[CrossRef](#)]
31. Ceresa, D.; Alessandrini, F.; Lucchini, S.; Marubbi, D.; Piaggio, F.; Mena Vera, J.M.; Ceccherini, I.; Reverberi, D.; Appolloni, I.; Malatesta, P. Early clonal extinction in glioblastoma progression revealed by genetic barcoding. *Cancer Cell* **2023**, *41*, 1466–1479. [[CrossRef](#)]
32. Quiñones-Hinojosa, A.; Chaichana, K. The human subventricular zone: A source of new cells and a potential source of brain tumors. *Exp. Neurol.* **2007**, *205*, 313–324. [[CrossRef](#)]
33. Neftel, C.; Laffy, J.; Filbin, M.G.; Hara, T.; Shore, M.E.; Rahme, G.J.; Richman, A.R.; Silverbush, D.; Shaw, M.L.; Hebert, C.M.; et al. An Integrative Model of Cellular States, Plasticity, and Genetics for Glioblastoma. *Cell* **2019**, *178*, 835–849.e21. [[CrossRef](#)]
34. Bhaduri, A.; Di Lullo, E.; Jung, D.; Müller, S.; Crouch, E.E.; Espinosa, C.S.; Ozawa, T.; Alvarado, B.; Spatzza, J.; Cadwell, C.R.; et al. Outer Radial Glia-like Cancer Stem Cells Contribute to Heterogeneity of Glioblastoma. *Cell Stem Cell* **2020**, *26*, 48–63.e6. [[CrossRef](#)] [[PubMed](#)]
35. Ravi, V.M.; Will, P.; Kueckelhaus, J.; Sun, N.; Joseph, K.; Salié, H.; Vollmer, L.; Kuliesiute, U.; von Her, J.; Benotmane, J.K.; et al. Spatially resolved multi-omics deciphers bidirectional tumor-host interdependence in glioblastoma. *Cancer Cell* **2022**, *40*, 639–655.e13. [[CrossRef](#)] [[PubMed](#)]
36. Huang, W.; Li, J.; Zhu, H.; Qin, X.; Chen, C.; Wang, B.; Wei, J.; Song, Y.; Lu, X.; Li, Z.; et al. A novel EGFR variant EGFRx maintains glioblastoma stem cells through STAT5. *Neuro Oncol.* **2024**, *26*, 85–99. [[CrossRef](#)]
37. Sherry, M.M.; Reeves, A.; Wu, J.K.; Cochran, B.H. STAT3 is required for proliferation and maintenance of multipotency in glioblastoma stem cells. *Stem Cells* **2009**, *27*, 2383–2392. [[CrossRef](#)]
38. Huang, M.; Zhang, D.; Wu, J.Y.; Xing, K.; Yeo, E.; Li, C.; Zhang, L.; Holland, E.; Yao, L.; Qin, L.; et al. Wnt-mediated endothelial transformation into mesenchymal stem cell-like cells induces chemoresistance in glioblastoma. *Sci. Transl. Med.* **2020**, *12*, eaay7522. [[CrossRef](#)]

39. Parakh, S.; Nicolazzo, J.; Scott, A.M.; Gan, H.K. Antibody drug conjugates in glioblastoma – Is there a future for them. *Front. Oncol.* **2021**, *11*, 718590. [[CrossRef](#)] [[PubMed](#)]
40. Mendez-Gomez, H.R.; DeVries, A.; Castillo, P.; von Roemeling, C.; Qdaisat, S.; Stover, B.D.; Xie, C.; Weidert, F.; Zhao, C.; Moor, R.; et al. RNA aggregates harness the danger response for potent cancer immunotherapy. *Cell* **2024**, *187*, 2521–2535.e21. [[CrossRef](#)]
41. Zhang, X.; Taylor, H.; Valdivia, A.; Dasari, R.; Buckley, A.; Bonacquisti, E.; Nguyen, J.; Kanchi, K.; Corcoran, D.L.; Herring, L.E.; et al. Auto-loaded TRAIL-exosomes derived from induced neural stem cells for brain cancer therapy. *J. Control. Release* **2024**, *372*, 433–445. [[CrossRef](#)]
42. Al-Sammarraie, N.; Ray, S.K. Applications of CRISPR-Cas9 Technology to Genome Editing in Glioblastoma Multiforme. *Cells* **2021**, *10*, 2342. [[CrossRef](#)] [[PubMed](#)]
43. Whittaker, M.N.; Testa, L.C.; Quigley, A.; Brooks, D.L.; Grandinette, S.A.; Said, H.; Dwivedi, G.; Jindal, I.; Volpp, D.; Hacker, J.L.; et al. Improved adenine base editing with hybrid gRNAs. *Nat. Biomed. Eng.* **2025**. [[CrossRef](#)]
44. Siddiquee, R.; Pong, C.H.; Hall, R.M.; Ataide, S.F. A programmable seekRNA guides target selection by IS1111 and IS110 type insertion sequences. *Nat. Commun.* **2024**, *15*, 5235. [[CrossRef](#)]
45. Brown, C.E.; Alizadeh, D.; Starr, R.; Weng, L.; Wagner, J.R.; Naranjo, A.; Ostberg, J.R.; Blanchard, M.S.; Kilpatrick, J.; Simpson, J.; et al. Regression of Glioblastoma after Chimeric Antigen Receptor T-Cell Therapy. *N. Engl. J. Med.* **2016**, *375*, 2561–2569. [[CrossRef](#)] [[PubMed](#)]
46. O'Rourke, D.M.; Nasrallah, M.P.; Desai, A.; Melenhorst, J.J.; Mansfield, K.; Morrissette, J.J.D.; Martinez-Lage, M.; Brem, S.; Maloney, E.; Shen, A.; et al. A single dose of peripherally infused EGFRvIII-directed CAR T cells mediates antigen loss and induces adaptive resistance in patients with recurrent glioblastoma. *Sci. Transl. Med.* **2017**, *9*, eaaa0984. [[CrossRef](#)]
47. Ahmed, N.; Brawley, V.; Hegde, M.; Bielamowicz, K.; Kalra, M.; Landi, D.; Robertson, C.; Gray, T.L.; Diouf, O.; Wakefield, A.; et al. HER2-Specific Chimeric Antigen Receptor-Modified Virus-Specific T Cells for Progressive Glioblastoma: A Phase 1 Dose-Escalation Trial. *JAMA Oncol.* **2017**, *3*, 1094–1101. [[CrossRef](#)]
48. Yang, T.; Kong, Z.; Ma, W. PD-1/PD-L1 immune checkpoint inhibitors in glioblastoma: Clinical studies, challenges and potential. *Hum. Vaccin. Immunother.* **2021**, *17*, 546–553. [[CrossRef](#)]
49. Cloughesy, T.F.; Mochizuki, A.Y.; Orpilla, J.R.; Hugo, W.; Lee, A.H.; Davidson, T.B.; Wang, A.C.; Ellingson, B.M.; Rytlewski, J.A.; Sanders, C.M.; et al. Neoadjuvant anti-PD-1 immunotherapy promotes a survival benefit with intratumoral and systemic immune responses in recurrent glioblastoma. *Nat. Med.* **2019**, *25*, 477–486. [[CrossRef](#)]
50. Desjardins, A.; Gromeier, M.; Herndon, J.E., 2nd; Beaubier, N.; Bolognesi, D.P.; Friedman, A.H.; Friedman, H.S.; McSherry, F.; Muscat, A.M.; Nair, S.; et al. Recurrent Glioblastoma Treated with Recombinant Poliovirus. *N. Engl. J. Med.* **2018**, *379*, 150–161. [[CrossRef](#)]
51. Lang, F.F.; Conrad, C.; Gomez-Manzano, C.; Yung, W.K.A.; Sawaya, R.; Weinberg, J.S.; Prabhu, S.S.; Rao, G.; Fuller, G.N.; Aldape, K.D.; et al. Phase I Study of DNX-2401 (Delta-24-RGD) Oncolytic Adenovirus: Replication and Immunotherapeutic Effects in Recurrent Malignant Glioma. *J. Clin. Oncol.* **2018**, *36*, 1419–1427. [[CrossRef](#)] [[PubMed](#)]
52. Pyonteck, S.M.; Akkari, L.; Schuhmacher, A.J.; Bowman, R.L.; Sevenich, L.; Quail, D.F.; Olson, O.C.; Quick, M.L.; Huse, J.T.; Teijeiro, V.; et al. CSF-1R inhibition alters macrophage polarization and blocks glioma progression. *Nat. Med.* **2013**, *19*, 1264–1272. [[CrossRef](#)]
53. Monje, M. The neuroscience of brain cancers. *Neuron* **2025**, *113*, 2734–2739. [[CrossRef](#)]
54. Dobsalske, C.; Rauschenbach, L.; Hua, Y.; Berliner, C.; Steinbach, A.; Grüneboom, A.; Kokkaliaris, K.D.; Heiland, D.H.; Berger, P.; Langer, S.; et al. Cranioencephalic functional lymphoid units in glioblastoma. *Nat. Med.* **2024**, *30*, 2947–2956. [[CrossRef](#)]
55. Cooper, E.; Posner, D.A.; Lee, C.Y.C.; Hu, L.; Bonner, S.; Taylor, J.T.; Baldwin, O.; Jimenez-Guerrero, R.; Masih, K.E.; Rahrmann, K.W.; et al. Childhood brain tumors instruct cranial hematopoiesis and immunotolerance. *Nat. Genet.* **2026**, *58*, 317–328. [[CrossRef](#)]
56. Soumbasis, A.; Ueno, A.; Elliott, D.; Lama, S.; Edwards, S.; Starreveld, Y.P.; Zhang, Y.; Federico, P.; Sutherland, G.R.; LeVan, P.; et al. The glymphatic system and glioblastoma. *Brain* **2025**, awaf449. [[CrossRef](#)] [[PubMed](#)]
57. Piyadasa, H.; Oberlton, B.; Ribi, M.; Leow, K.; Ranek, J.S.; Averbukh, I.; Amouzgar, M.; Liu, C.C.; Franchina, D.G.; Greenwald, N.F.; et al. Multi-omic landscape of human gliomas from diagnosis to treatment and recurrence. *Cancer Cell* **2026**, *44*, 112–128.e6. [[CrossRef](#)] [[PubMed](#)]
58. Pinto, G.; Saenz-de-Santa-Maria, I.; Chastagner, P.; Perthame, E.; Delmas, C.; Toulas, C.; Moyal-Jonathan-Cohen, E.; Brou, C.; Zurzolo, C. Patient-derived glioblastoma stem cells transfer mitochondria through tunneling nanotubes in tumor organoids. *Biochem. J.* **2021**, *478*, 21–39. [[CrossRef](#)] [[PubMed](#)]
59. Mohamed, Z.S.; Wu, Q.; Jacome, M.A.; Chen, Z.; Etame, A.B. Gut microbiome in GBM evolution. *Int. J. Mol. Sci.* **2025**, *26*, 2935. [[CrossRef](#)]
60. Xu, C.; Yang, J.; Xiong, H.; Cui, X.; Zhang, Y.; Gao, M.; He, L.; Fang, Q.; Han, C.; Liu, W.; et al. Machine learning and multi-omics analysis reveal key regulators of proneural-mesenchymal transition in glioblastoma. *Sci. Rep.* **2025**, *15*, 19731. [[CrossRef](#)]

61. DeMeo, B.; Nesbitt, C.; Miller, S.A.; Burkhardt, D.B.; Lipchina, I.; Fu, D.; Holderrieth, P.; Kim, D.; Kolchenko, S.; Szalata, A.; et al. Active learning framework leveraging transcriptomics identifies modulators of disease phenotypes. *Science* **2025**, *390*, eadi8577. [[CrossRef](#)]
62. Chen, W.; Zhang, P.; Tran, T.N.; Xiao, Y.; Li, S.; Shah, V.V.; Cheng, H.; Brannan, K.W.; Youker, K.; Lai, L.; et al. A visual-omics foundation model to bridge histopathology with spatial transcriptomics. *Nat. Methods* **2025**, *22*, 1568–1582. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.